

Compendium

Clinical Publications and Scientific Validation for CaNa_2 EDTA Chelation Suppositories (Detoxamin[®])

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Compendium of Scientific and Clinical Studies on CaNa₂ EDTA Chelation Suppositories (Detoxamin[®])

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Compendium of Scientific and Clinical Studies on CaNa₂ EDTA Chelation Suppositories (Detoxamin[®])

Introduction

The following is a compilation of research documents and publications on pharmacokinetics, safety and efficacy of Detoxamin chelation suppositories. These studies were conducted by the following teams of researchers and physicians:

| | |
|----------------------------------|--|
| Rita Ellithorpe, MD..... | Tustin Longevity Center, Tustin, CA |
| Brett Jacques, ND..... | “ “ “ |
| Tony Jimenez, MD..... | Hope4Cancer Institute, Baja California, MX |
| Theodore Rosema, MD..... | ACAM President Elect, Landrum, South Carolina |
| Garth Nicolson, PhD..... | Inst. for Molecular Medicine, Laguna Beach, CA |
| Robert Settineri, MS..... | Sierra Research, Irvine, CA |
| Larry Clapp, PhD, JD..... | Prostate Health Resources, Santa Monica, CA |
| M Pelletier, MD, CCN, FACOG..... | Living Longer Inst., Cincinnati, Ohio |
| James Lavelle, ND, RPh, CCN..... | “ “ “ |
| Michael Schmidt, PhD, CCN | “ “ “ |
| Ross Pelton, RPh, PhD, CCN | “ “ “ |
| Harold Ravins, DDS..... | Center for Holistic Dentistry, Los Angeles, CA |
| Paul Mazur, PhD..... | Biological Test Center (BTC), Irvine, CA |
| Glenwood Gum, PhD..... | “ “ “ |
| Ernest. H. Pfadenhauer, MS..... | “ “ “ |
| Gerry Button, BS..... | “ “ “ |
| Julian Le, BS..... | “ “ “ |

ORIGINAL RESEARCH

Comparison of the Absorption, Brain and Prostate Distribution, and Elimination of CaNa_2 EDTA of Rectal Chelation Suppositories to Intravenous Administration

Rita Ellithorpe, MD,¹ Paul Mazur, PhD, Glenwood Gum, PhD,² Gerry Button, BS,²
Julian Le, BS,² Ernest H. Pfadenhauer, MS,³ Robert A. Settineri, MS,³ Garth Nicolson, PhD⁴

1. Tustin Longevity Center, Tustin, California

2. Biological Test Center, Irvine, California

3. Research Consultant, Sierra Research, Irvine, California

4. The Institute for Molecular Medicine, Laguna Beach, California



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Comparison of the Absorption, Brain and Prostate Distribution, and Elimination of CaNa₂ EDTA of Rectal Chelation Suppositories to Intravenous Administration

Rita Ellithorpe, MD,^{1*} Paul Mazur, PhD, Glenwood Gum, PhD,² Gerry Button, BS,² Julian Le, BS,² Ernest H. Pfadenhauer, MS,³ Robert A. Settineri, MS,³ Garth Nicolson, PhD⁴

1. Tustin Longevity Center, Tustin, California

2. Biological Test Center, Irvine, California

3. Research Consultant, Sierra Research, Irvine, California

4. The Institute for Molecular Medicine, Laguna Beach, California

ABSTRACT

Rectal suppositories were compared to IV administration of C14-labeled calcium disodium ethylenediaminetetraacetate (CaNa₂EDTA) to evaluate the absorption, brain and prostate tissue distribution, and excretion in rats. The absolute bioavailability of CaNa₂EDTA in blood following rectal dosing was 36.3% of the IV dose route, which confirmed that rectal dosing is an efficient method for delivering ethylenediaminetetraacetic acid (EDTA) to tissues. The ratio of radioactive residues of EDTA in tissues compared to blood, following IV or rectal dosing of C14 labeled CaNa₂EDTA, showed negligible brain localization. However, prostate tissues were found to have a mean ratio of 3.69 via the IV route and 13.6 rectally. The total recovery of C14 EDTA expressed as percent of administered dosed IV was a mean of 47.3% and 30.3% rectally at eight hours when the test was concluded. The suppository formulation of CaNa₂ appears to be well absorbed, delivering high levels of EDTA to prostate tissue.

INTRODUCTION

Heavy metal exposures in the twenty-first century are an established global health concern. The FDA has approved EDTA as a chelation agent for the removal of heavy metals. It has been placed on the FDA "Generally Recognized as Safe" (GRAS) list for the past sixty years. Extensive national and international clinical experiences demonstrate that acute and chronic human exposure to a wide range of heavy metals can be treated with considerable efficacy using EDTA. It is widely administered, with considerable cost to the patient, as an intravenous (IV) solution, which entails 15 to 30 sessions in a physician's office, taking two to five hours per visit. The transrectal delivery of several pharmacological agents is well established. Therefore, using a rat animal model, we set out to determine if the rectal administration of EDTA is absorbed, resulting in significant blood and tissue levels.

The pharmacodynamic effects of therapeutic agents differ widely in their route of administration, penetration, absorption, and distribution in body tissues. For medicinal agents to act, they must be absorbed and transported to the appropriate tissue or organ, penetrate to the responding cell surface or sub-cellular and interstitial space, and elicit a response or alter ongoing processes.¹ The parenteral and intramuscular forms of EDTA are well absorbed, but not very practical for routine usage.² Oral forms of EDTA have been shown to be poorly absorbed (2% to 5%), and topical and subcutaneous forms have been reported as not being

* Correspondence:

Rita Ellithorpe, MD

Tustin Longevity Center

13422 Newport Avenue # L

Tustin, CA 92780

Phone: 714-544-1521 Fax: 714-544-1904

E-mail: edaly@tlcmd.net

absorbed at all.^{3,4,5,6,7,8,9,10} A relatively new alternative and more convenient route of administration is rectal suppository delivery of a proprietary suppository formula of EDTA (CaNa₂ EDTA, Detoxamin,[®] World Health Products, Draper, Utah), which is the basis of this pharmacokinetic (PK) study. Although IV EDTA dosing is well characterized and has been used for decades, little is known about the absorption of rectal suppositories.

In an effort to elucidate the absorption characteristics of CaNa₂ EDTA in a suppository form, a rat model was chosen. ¹⁴C-labeled EDTA Calcium Disodium salt was administered as a tracer in the suppository and in intravenous forms; blood, urine, and selected tissue levels were evaluated over eight hours.

MATERIALS AND METHODS

¹⁴C-labeled EDTA free acid (11.7 mCi/mmol, Lot No. 63151012, purity greater than 98%) was obtained from MP Biomedicals (Irvine, CA). For the IV dosing solution, ¹⁴C-labeled EDTA was added to normal saline to achieve concentrations needed to deliver a final dose of 7.53 µCi in approximately 1 gram. The rectal suppository (a proprietary suppository formula of EDTA, CaNa₂ EDTA, Detoxamin[®] Health Products, East Draper, Utah, Lot No. 228-190-0117) was prepared by adding ¹⁴C-labeled EDTA solution from Moravsek to molten suppository. For the animal dose, approximately 100 µL of the mixture (containing 23.7 µCi per dose) was taken up in a cylindrical glass pipette equipped with a plunger and allowed to cool to room temperature, where it re-solidified.

The radioactive concentration of the IV dosing solution was calculated by Liquid Scintillation Counting (LSC). The prepared dosing solutions were stored and refrigerated.

Ten male Sprague Dawley rats were obtained from Taconic, Oxnard, CA. Animals were 6 to 7 weeks old and weighed 157 to 187 grams on Day 1. The animal experiments were performed at the Biological Test Center (BTC), in Irvine, CA. Quarantine and care of animals were performed per BTC Standard Operating Procedures.

Prior to dosing, 10 animals were weighed. Cannulated animals (six animals to undergo IV dosing) were random-

ized for placement into Group A or B. Uncannulated animals (four animals to undergo rectal dosing) were not randomized and were placed into Group C. Treatment groups are presented below.

Animals were fasted (food withheld) for 16.5 to 19.5 hours before ¹⁴C-EDTA administration. Prior to dosing, rats were anesthetized with an intramuscular combination injection of ketamine hydrochloride (40-90 mg/kg) and xylazine (5-10 mg/kg). Water and feed were withheld from animals for four hours after ¹⁴C-EDTA administration, and then food and water were given *ad libitum*.

For Group C, the contents of the colon were removed before dosing by flushing with normal saline heated to 37°C. Rectal doses were administered via a 100 µL glass cylindrical tube, gently heated to allow partial liquefaction of the suppository material. Blood samples of approximately 100 µL were taken. Each sample was placed in combustion cones and stored frozen prior to combustion and LSC analysis. The time of blood collection was recorded.

A terminal blood sample was collected from all animals via heart puncture (1 hour ± 5 minutes after dosing for Group A animals; 8 hours ± 15 minutes after dosing for Group B and C animals). Each animal was anesthetized with an intramuscular combination injection of ketamine hydrochloride (40-90 mg/kg) and xylazine (5-10 mg/kg), and euthanized by exsanguination following heart puncture. As much blood as possible was collected from each rat into heparinized tubes. The time of blood collection was recorded. Four 100-µL aliquots of whole blood were transferred to combustion cones. Two of the aliquots were combusted for determination of radioactivity by LSC, and two were kept frozen as reserve samples.

Absorbent paper was placed in the restrainers to collect urine 0 to 4 hours after dosing. Urine was collected from the individual metabolism cages 4 to 8 hours after dosing. For urine samples collected in absorbent paper, water was added to the paper and urine extracted. For urine samples collected from metabolism cages, the urine was freeze-trapped to avoid atmospheric oxidation, evaporation, and bacterial degradation, and the urine collection pan was rinsed with water.

Following euthanasia by exsanguination, the brain and prostate were collected from each animal. Prior to collec-

| Group | No. | Treatment | ¹⁴ C-EDTA Dose (µCi) | Route | Blood Collection Time points (Time After Dosing) ¹ |
|-------|----------------|----------------------|---------------------------------|--------|---|
| A | 2 | ¹⁴ C-EDTA | 10 | IV | 1 hour |
| B | 4 | ¹⁴ C-EDTA | 10 | IV | 5, 15, 30 minutes; 1, 2, 4, 8 hours |
| C | 3 ² | ¹⁴ C-EDTA | 20 | Rectal | 5, 15, 30 minutes; 1, 2, 4, 8 hours |

1. Blood collection times were ± 1 minute for the 5-minute time point; ± 3 minutes for the 15- and 30-minute time points; ± 5 minutes for the 1-hour time point; and ± 15 minutes for the 2-, 4-, and 8-hour time points.

2. The fourth animal in group C, animal, No. 55905, was dead (attributed to anesthesia) 15 minutes after dosing.

tion, the brain was perfused with approximately 5 mL of saline via the carotid artery. Both organs were stored at -20° C. Following completion of blood kinetics analysis, brains and prostates were combusted for determination of radioactivity by LSC. Brains were homogenized prior to combustion, while prostates were directly combusted.

Duplicate aliquots of each urine sample (0.1 mL) and cage rinse sample (1 mL) were transferred to liquid scintillation counting vials and the amount of radioactivity determined by LSC; Insta-Gel was used as the scintillation fluid. Each of the rectal dosing solution samples, tail vein blood samples, and heart puncture blood samples in combustion cones were combusted. Brain and prostate samples were combusted. Combusted samples were trapped in Carbon-14 Cocktail (R.J. Harvey, Hillsdale, NJ) present in liquid scintillation counting vials, and the amount of radioactivity was determined by LSC.

Sample combustion was performed using a Harvey Sample Oxidizer, Model OX300 (Harvey Instrument, Hillsdale, NJ). All radioactivity measurements were performed using a Beckman Liquid Scintillation Spectrometer. Any radioactivity measurement of less than 100 dpm was considered close to background and was not repeated.

When applicable, summary statistics (mean and standard deviation) were prepared to characterize the data (i.e., radioactivity measurement and percent dose). PK parameters, including Area under the Curve (AUC), half-life, Maximum Concentration in blood (C_{max}), Time to Maximum Concentration (T_{max}), and bioavailability, were calculated using WinNonlin (Pharsight Corporation, Mountain View, CA).

RESULTS

Individual and mean (\pm SD) body weights and administered ^{14}C -EDTA doses are presented in Table 1. Radioactivity recovered from blood at different time intervals is presented in Figures 1 and 2. As shown in Figure 2, the absorption phase occurring within the first two hours after dosing for all three rectally-dosed animals was maximal, and the apparent biphasic absorption may have been related to additional material being released from the rectal suppository; the blood levels from the IV doses did not show a biphasic response.

Mean AUC, half-life, C_{max} , T_{max} , and bioavailability of derived radioactivity in blood are presented in Table 2. The T_{max} of EDTA following intravenous dosing occurred at 0.083 hours. The T_{max} of EDTA following rectal dosing occurred at 0.417 hours. The half-life of EDTA following intravenous dosing was 1.50 hours, and the half-life of EDTA following rectal dosing could not be calculated since a terminal elimination phase could not be determined. The absolute bioavailability of EDTA in blood following rectal

dosing was 36.3 compared to the IV bolus of 100%. Radioactivity recovered from urine at different time intervals is presented in Table 3. Following intravenous dosing, the amount of radioactivity excreted in urine decreased over the 8-hour study period (46.3% of dosed radioactivity excreted at the 0 to 4 hour interval, and 0.935% of dosed radioactivity excreted at the 4 to 8 hour interval). Following rectal dosing, the amount of radioactivity excreted in urine remained relatively constant over the 8-hour study period (15.8% of dosed radioactivity excreted at the 0 to 4 hour interval, and 14.4% of dosed radioactivity excreted at the 4 to 8 hour interval).

Radioactivity recovered from tissues (brain and prostate) expressed as a ratio of the radioactivity in blood is

Figure 1. EDTA levels in blood over time following intravenous administration of ^{14}C -EDTA.

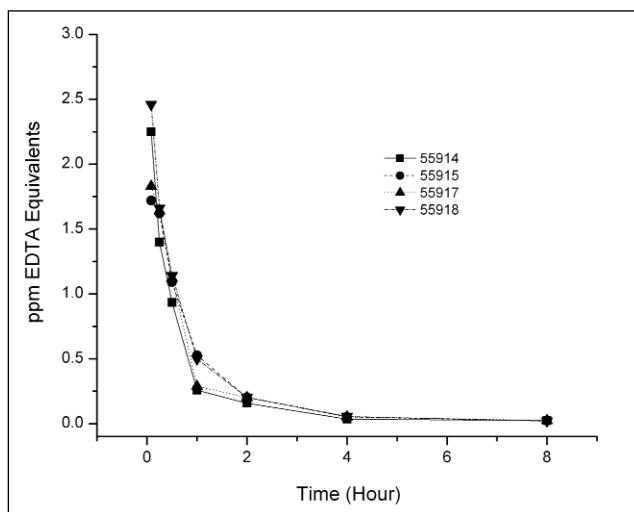


Figure 2. EDTA levels in blood over time following rectal administration of ^{14}C -EDTA

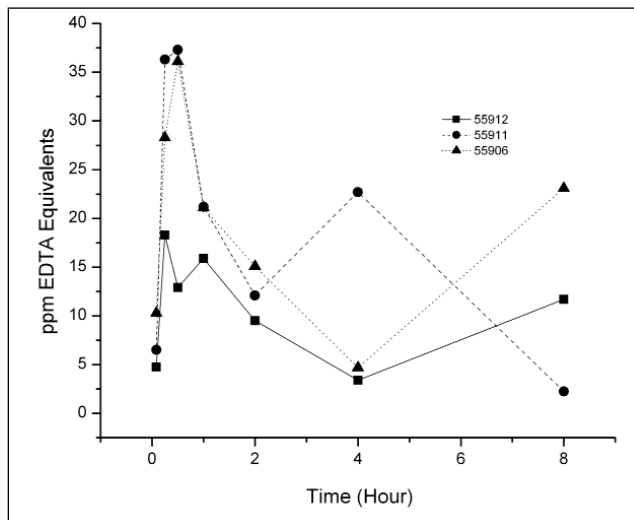


Table 1. Body Weights and Administered ¹⁴C-EDTA Doses

| Group | Animal Number | Body Weight (kg) | Dosage Weight (g) | Dose (mg/kg) | Total Dose (μCi) | Total Dose (dpm) |
|-------|---------------|------------------|-------------------|--------------|------------------|------------------|
| A | 55921 | 0.187 | 1.0019 | 1.31 | 7.64 | 16,954,958 |
| A | 55920 | 0.184 | 0.9906 | 1.31 | 7.55 | 16,763,730 |
| | Mean: | 0.186 | | 1.31 | 7.59 | 16,859,344 |
| | ± SD: | 0.002 | | 0.00 | 0.06 | 135,218 |
| B | 55918 | 0.166 | 0.9899 | 1.45 | 7.55 | 16,751,884 |
| B | 55915 | 0.175 | 0.9776 | 1.36 | 7.45 | 16,543,734 |
| B | 55917 | 0.177 | 0.9718 | 1.34 | 7.41 | 16,445,582 |
| B | 55914 | 0.183 | 0.9804 | 1.31 | 7.47 | 16,591,118 |
| | Mean: | 0.175 | | 1.37 | 7.47 | 16,583,079 |
| | ± SD: | 0.007 | | 0.06 | 0.06 | 127,819 |
| C | 55912 | 0.167 | 0.1100 | 214.4 | 24.6 | 54,534,057 |
| C | 55911 | 0.157 | 0.0970 | 201.1 | 21.7 | 48,089,123 |
| C | 55906 | 0.162 | 0.1120 | 225.0 | 25.0 | 55,525,585 |
| | Mean: | 0.162 | | 213.5 | 23.7 | 52,716,255 |
| | ± SD: | 0.005 | | 12.0 | 1.8 | 4,037,765 |

Table 2. Mean AUC, half-life, C_{max}, T_{max}, and bioavailability of EDTA in blood following intravenous or rectal administration of ¹⁴C-EDTA

| Group | Route | Stat. | Dose (mg/kg) | AUC (μg x Hr/mL) | AUC Inf (μg x Hr/mL) | Half-life (Hour) | C _{max} (μg/mL) | T _{max} (Hour) | Absolute Bioavailability (%) |
|-------|-------------|-------|--------------|------------------|----------------------|------------------|--------------------------|-------------------------|------------------------------|
| B | Intravenous | MEAN | 1.37 | 1.86 | 1.91 | 1.50 | 2.07 | 0.083 | N/A |
| | | SD | 0.06 | 0.20 | 0.19 | 0.34 | 0.35 | 0.000 | |
| | | N | 4 | 4 | 4 | 4 | 4 | 4 | |
| C | Rectal | MEAN | 213.5 | 105.8 | 307.3 | N/A ¹ | 30.6 | 0.417 | 36.3 |
| | | SD | 12.0 | 32.2 | 225.6 | N/A ¹ | 10.6 | 0.144 | |
| | | N | 3 | 3 | 3 | 3 | 3 | 3 | |

N/A = not applicable

$$\text{Absolute bioavailability (\%)} = \frac{(\text{AUC}_{\text{test}} \times \text{Dose}_{\text{ref}})}{(\text{AUC}_{\text{ref}} \times \text{Dose}_{\text{test}})} \times 100$$

Where “test” data is the rectal data, and “ref” (reference) data is the intravenous data.

1. The terminal elimination phase was not observed, therefore, the half-life could not be calculated.

References for the above formula are as follows:

- 1) Kwon Y. *Handbook of Essential Pharmacokinetics, Pharmacodynamics, and Drug Metabolism for Industrial Scientists*. New York: Kluwer Academic/Plenum Publishers, 2001.
- 2) Shargel L, Yu A. *Applied Biopharmaceutics and Pharmacokinetics*, 4th ed. Norwalk, Connecticut: Appleton & Lange, 1999

presented in Table 4 and Figure 3. The prostate retained higher levels of radioactivity than the brain following both intravenous and rectal dosing, with the highest level of radioactivity found in the prostate following rectal dosing. The total recovery of radioactivity from urine and tissues expressed as percent of dose is presented in Tables 5 and 6. Total recovery represents the combined total percent of dose in urine and tissues. Following intravenous dosing, 41.4% and 47.3% of the radioactive dose was recovered 1 hour and 8 hours after dosing, respectively; of which virtually all was in urine. Following rectal dosing, 30.3% of the radioactive dose was recovered 8 hours after dosing, of which virtually all was also in urine.

DISCUSSION

This study has shown that the proprietary formula of Ca Na₂ EDTA has been effectively absorbed from the lower enteral route in rats, through the anal portal into the rectum or lower intestine to reach blood and tissue levels via rectal sup-

Figure 3. Ratio of tissue and blood radioactive residues (ppm) following intravenous or rectal administration of ¹⁴C-EDTA.

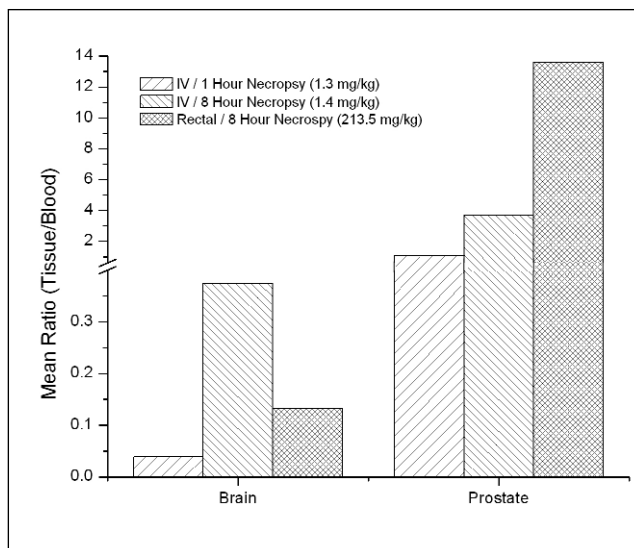


Table 3. ¹⁴C-EDTA-derived radioactivity excreted in urine expressed as percent of administered dose following intravenous or rectal administration of ¹⁴C-EDTA

| Group A: IV (1 hour) Time Interval (Hour) | Animal No. 55921 | | Animal No. 55920 | | % Dose | | Cum. % Dose | |
|--|------------------|-------------|------------------|-------------|-------------|--------|-------------|--------|
| | % Dose | Cum. % Dose | % Dose | Cum. % Dose | Mean Values | ± S.D. | Mean Values | ± S.D. |
| | | | | | | | | |
| 0-1 | 38.1 | 38.1 | 44.7 | 44.7 | 41.4 | 4.7 | 41.4 | 4.67 |
| Total | 38.1 | | 44.7 | | | | 41.4 | 4.67 |

| Group B: IV (8 Hour) Time Interval (Hour) | Animal No. 55918 | | Animal No. 55915 | | Animal No. 55917 | | Animal No. 55914 | | % Dose | | Cum. % Dose | |
|--|------------------|-------------|------------------|-------------|------------------|-------------|------------------|-------------|-------------|--------|-------------|--------|
| | % Dose | Cum. % Dose | % Dose | Cum. % Dose | % Dose | Cum. % Dose | % Dose | Cum. % Dose | Mean Values | ± S.D. | Mean Values | ± S.D. |
| | | | | | | | | | | | | |
| 0 - 4 | 58.5 | 58.5 | 53.4 | 53.4 | 38.2 | 38.2 | 35.2 | 35.2 | 46.3 | 11.4 | 46.3 | 11.4 |
| 4 - 8 | 1.20 | 59.7 | 0.78 | 54.1 | 0.97 | 39.2 | 0.79 | 36.0 | 0.935 | 0.197 | 47.3 | 11.5 |
| Total | 59.7 | | 54.1 | | 39.2 | | 36.0 | | | | 47.3 | 11.5 |

| Group C: Rectal (8 Hour) Time Interval (Hour) | Animal No. 55912 | | Animal No. 55911 | | Animal No. 55906 | | % Dose | | Cum. % Dose | |
|--|------------------|-------------|------------------|-------------|------------------|-------------|-------------|--------|-------------|--------|
| | % Dose | Cum. % Dose | % Dose | Cum. % Dose | % Dose | Cum. % Dose | Mean Values | ± S.D. | Mean Values | ± S.D. |
| | | | | | | | | | | |
| 0 - 4 | 20.0 | 20.0 | 16.7 | 16.7 | 10.8 | 10.8 | 15.8 | 4.67 | 15.8 | 4.67 |
| 4 - 8 | 2.53 | 22.5 | 19.7 | 36.4 | 21.1 | 31.9 | 14.4 | 10.3 | 30.3 | 7.08 |
| Total | 22.5 | | 36.4 | | 31.9 | | | | 30.3 | 7.08 |

positories. Bioavailability has now been established for this mode of administration in an animal model and is strong evidence that EDTA suppositories are an adequate and medically acceptable approach to providing the benefits of chelation.

Intravenous dosing resulted in greater elimination of radioactivity in urine at the 0 to 4 hour time point, but the percent of dose recovered drastically decreased by the 4 to 8 hour time point, while the level of recovery was relatively steady at both time points following rectal dosing. The slow and consistent movement of CaNa_2EDTA via rectal administration may have lesser toxicity since there are significant blood and tissue levels to chelate metals without a high dose EDTA IV drip over many hours. These data point to the ability of rectal suppositories to deliver a continuous lower dose concentration of EDTA for longer periods of time compared with IV administration, allowing EDTA to bind metals efficiently and effectively.

In tissues, significant amounts of radioactivity were recovered from the prostate following intravenous or rectal dosing, with the highest level of dosed radioactivity (179.6 ppm) recovered 8 hours following rectal dosing. This observation of rectal administration, revealing higher amounts of EDTA in prostate tissue as compared to IV, can have far-reaching implications of a more complete distribution of EDTA into interstitial and intracellular spaces, further leading to more efficient chelation of compartmental-

ized heavy metal content with CaNa_2 suppositories.

EDTA is not bio-transformed in the body. It is excreted in hair, urine, feces, saliva, and perspiration. This study shows that animals excreted 47.3% and 30.3% of dosed radioactivity in urine during the 8 hours following intravenous and rectal dosing, respectively. The 30.3% excretion of EDTA in the urine corresponds closely to the rectal dose bioavailability calculated from the blood levels (36.3%).

Blood samples were taken over an 8-hour period, and during this time, the rectal administration showed high levels of absorbed EDTA with no apparent elimination phase observed. If further blood samples had been taken, the bioavailability calculated for rectally administered EDTA would have undoubtedly been much higher, since the bioavailability calculation presented here only used up to 8-hour blood level data. No extrapolation of the AUC could be done since the levels at 8 hours were actually increasing in two out of three animals. Further research is indicated over a longer time span to quantify the actual half life of the suppository form of administration.

CONCLUSIONS

This proprietary suppository formulation appears to be a viable dosing mechanism for delivery of CaNa_2EDTA to the bloodstream in this rat model, showing substantial circulating levels of EDTA for least 8 hours after administra-

Table 4. Ratio of radioactive residues of EDTA in tissues ($\mu\text{g/g}$) to blood ($\mu\text{g/g}$) following intravenous or rectal administration of ^{14}C -EDTA

| Group A: IV (1 Hour) | Ratio of Radioactive Residues of EDTA | | | |
|----------------------|---------------------------------------|------------------|-------------|----------|
| | Animal No: 55921 | Animal No: 55920 | Mean Values | \pm SD |
| Brain | 0.039 | 0.039 | 0.039 | 0.000 |
| Prostate | 1.80 | 0.357 | 1.08 | 1.02 |

| Group B: IV (8 Hour) | Ratio of Radioactive Residues of EDTA | | | | | |
|----------------------|---------------------------------------|------------------|------------------|------------------|-------------|----------|
| | Animal No: 55918 | Animal No: 55917 | Animal No: 55915 | Animal No: 55914 | Mean Values | \pm SD |
| Brain | 0.345 | 0.318 | 0.487 | 0.351 | 0.375 | 0.076 |
| Prostate | 6.70 | 2.01 | 3.07 | 2.98 | 3.69 | 2.06 |

| Group C: Rectal (8 Hour) | Ratio of Radioactive Residues of EDTA | | | | |
|--------------------------|---------------------------------------|------------------|------------------|-------------|----------|
| | Animal No: 55912 | Animal No: 55911 | Animal No: 55906 | Mean Values | \pm SD |
| Brain | 0.050 | 0.288 | 0.056 | 0.132 | 0.135 |
| Prostate | 8.90 | 14.4 | 17.4 | 13.6 | 4.31 |

Note: Brain was perfused with normal saline prior to collection.

Table 5. Total recovery of radioactivity expressed as percent of administered dose following intravenous or rectal administration of ¹⁴C-EDTA.

| Group | Animal No. | % Administered Dose | | |
|-------|------------|---------------------|--------|-------|
| | | Urine | Tissue | Total |
| A | 55921 | 38.1 | 0.05 | 38.2 |
| | 55920 | 44.7 | 0.02 | 44.7 |
| | Mean: | 41.4 | 0.04 | 41.4 |
| | ± SD: | 4.67 | 0.02 | 4.65 |
| | | | | |
| B | 55918 | 59.7 | 0.02 | 59.8 |
| | 55915 | 54.1 | 0.01 | 54.2 |
| | 55917 | 39.2 | 0.01 | 39.2 |
| | 55914 | 36.0 | 0.02 | 36.0 |
| | Mean: | 47.3 | 0.02 | 47.3 |
| | ± SD: | 11.5 | 0.01 | 11.5 |
| | | | | |
| C | 55912 | 22.5 | 0.01 | 22.5 |
| | 55911 | 36.4 | 0.01 | 36.4 |
| | 55906 | 31.9 | 0.10 | 32.0 |
| | Mean: | 30.3 | 0.04 | 30.3 |
| | ± SD: | 7.08 | 0.05 | 7.09 |
| | | | | |

Table 6. Total recovery of radioactivity expressed as percent of administered dose following intravenous or rectal administration of ¹⁴C- EDTA.

| Sample | % Administered Dose | | |
|--------|---------------------|---------|---------|
| | Group A | Group B | Group C |
| Urine | 41.4 | 47.3 | 30.3 |
| Tissue | 0.04 | 0.02 | 0.04 |
| Total | 41.4 | 47.3 | 30.3 |

tion. EDTA appears to be favorably distributed to the prostate, but not the brain, following both IV and rectal dosing. The excretion of rectal CaNa₂ EDTA administration in urine corresponds well with the rectal dose bioavailability of blood levels. The absolute bioavailability of EDTA in blood following rectal dosing was 36.3% within the 8-hour period. Additional testing is required to confirm and duplicate these results in humans.

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REFERENCES:

- University of the SC/Remington. The science and practice of pharmacy. Columbia, S.C.: Lippincott Williams & Wilkins/Wolters Kluwer, 21:2006.
- Foreman H. *Metal Binding Med*, Proc Symposium, Philadelphia, 1959. 1960;82-94.
- Sbrova J Teisinger J, Arc. *Gewerbepathol*. 1957;15:572.
- Foreman H. *Metal Binding Med*, Proc Symposium, Philadelphia, 1959. 1960;82-94.
- Foreman H, Vier M, Magee M. The metabolism of C¹⁴ labeled ethylenediamine tetraacetic acid in the rat, *J Biol Chem*. 1953;203:1045.
- Yang, SS. Ethylenediaminetetraacetate, Disodium and Calcium Disodium Salts. Seventeenth Report of the Joint FAO/WHO Expert Committee on Food Additives. Wld Hlth Org.techn. rep. ser., 1974, No. 539; FAO Nutrition Meetings Report Series, 1974, No. 53. <http://www.inchem.org/documents/jecfa/jecmono/v05je25.htm>
- Cleton F, Turnbull A, Finch, CA. Synthetic chelating agents in iron metabolism. *J Clin Invest*. 1963 March; 42(3): 327-337.
- Foreman H, Trujillo TT. Synthetic chelating agents in iron metabolism. *J Lab Clin Med*. 1954 Apr;43(4):566-571.
- MacPhail AP, Bothwell TH, Torrance JD, et al. Factors affecting the absorption of iron from Fe(III)EDTA. *Br J Nutr*. 1981 Mar;45(2):215-227.
- Bjarnason I, O'Morain C, Levi AJ, Peters TJ. Absorption of 51 chromium-labeled ethylenediaminetetraacetate in inflammatory bowel disease. *Gastroenterol*. 1983 Aug;85(2):318-322.

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Anti-Microbial plus CaNa₂EDTA Chelation Suppository Therapy for Chronic Prostatitis/Pelvic Pain Syndrome with or without Prostatic Hyperplasia: Preliminary Study

Rita Ellithorpe¹, MD, Larry Clapp², JD, Tony Jimenez³, MD, Brett Jacques⁴, ND, Robert Settineri⁵, MS, PhD, and Garth L. Nicolson⁶, PhD

Tustin Longevity Center, Tustin, CA, ²Prostate Health Resources, Santa Monica, CA, ³Hope 4 Cancer Institute, Baja California, Mexico, ⁴Tustin Longevity Center, Tustin, CA, ⁵Sierra Research, Irvine, CA, and ⁶Institute for Molecular Medicine, Laguna Beach, CA

Abstract

Patients with chronic prostatitis/pelvic pain syndrome are characterized by treatment failures, a high incidence of prostate calcium deposits and poor quality of life. We used proprietary suppositories containing calcium disodium ethylenediaminetetraacetic acid (CaNa₂EDTA) to remove metal and calcium cations. Participants (N=31) with chronic prostatitis, with or without prostate hyperplasia, and prostate calculi (mean age=61) were treated with tetracycline (500 mg/day) and CaNa₂EDTA suppositories (750 mg 4X/week) for 90 days. Using the NIH Chronic Prostatitis Symptom Index significant post-treatment mean reductions in symptoms ($p<0.0106$) and pain ($p<0.0122$) were found along with a significant improvement in mean quality of life ($p<0.0022$). Overall, the mean total scores showed significant post-treatment reduction ($p<0.0006$). In addition, use of the International Prostate Symptom Score indicated significant reductions in 5/7 symptom categories and significant reduction of mean overall scores ($p<0.0001$). Analysis of blood and stool post-treatment indicated significant changes in mobilized, secreted cations (cadmium, copper, boron, lead, molybdenum, magnesium and calcium). In addition, the blood cholesterol/high density lipoprotein ratio was significantly decreased ($p<0.0005$). However, using the Erectile Function Index Questionnaire there was significant mean improvement in only 7/15 questions, resulting in non-significant overall mean improvement, and there were non-significant reductions in prostate calcifications at this suppository dose level. The data suggest that combining CaNa₂EDTA suppositories with tetracycline can significantly reduce symptoms and pain in refractory chronic prostatitis with or without prostate hyperplasia.

Keywords: Prostatitis symptoms, antibiotic, EDTA suppository, bacterial biofilm, prostate calcifications

Corresponding Author: Prof. Garth Nicolson, Dept. of Molecular Pathology, The Institute for Molecular Medicine, P.O. Box 9355, Laguna Beach, CA 92652; Email: gnicolson@immed.org

Introduction

Prostatitis is a common urological condition, and it has been estimated that up to one-half of men will suffer from symptoms of this disease at some time during their lives [1]. At any one time 2-10% of men suffer from prostatitis [2]. Prostatitis is classified within a complex series of syndromes (NIH category I-IV prostatitis) that vary widely in clinical presentation and response to treatment. Acute bacterial prostatitis (category I) and chronic bacterial prostatitis (category II) are characterized by uropathogenic infections of the prostate gland that respond well to antimicrobial treatment, whereas chronic prostatitis/chronic pelvic pain syndrome (category III, accounting for 90%-95% of prostatitis cases) is marked by a mixture of pain, urinary and ejaculatory symptoms and presence of prostate calculi with no uniformly effective therapy [3].

Several different classes of bacterial infections have been found in acute and chronic prostatitis, including Gram-positive and -negative bacteria [1, 2, 4, 5], cell wall-deficient forms (*Mycoplasma*, *Ureaplasma*, *Chlamydia*) [4, 6, 7] and biofilm-forming bacteria [8-10]. The latter forms are likely to be important in chronic prostatitis and have proven to be particularly difficult to effectively treat [10, 11]. Biofilms contain bacterial glycosaminoglycans, salts (especially calcium and other cations) and other molecules [12, 13]. According to Parsek and Singh [14] biofilms support bacterial: (a) adhesion, (b) clustering, (c) localized infection, and (d) increased resistance to antibiotic treatment in the host. They do this by providing a protective structure for bacterial colonies so that they can evade mechanical stresses, host responses and antimicrobial agents.

Calcium and other cations are thought to play a structural role in biofilms as well as in cellular and tissue deposits, such as calculi, a calcium apatite-containing mineral deposit that is often associated with category III chronic prostatitis/chronic pelvic pain syndrome [15]. In some studies the presence of calculi is a feature that is associated with antimicrobial resistance and treatment failure [16, 17]. Although the presence of calculi in chronic prostatitis/chronic pelvic pain syndrome and its association with disease symptoms are controversial [17], Geramoutsos et al. [18] found that larger deposits of calculi were associated with chronic prostatitis symptoms.

Patients with NIH Category III chronic prostatitis/chronic pelvic pain syndrome have a high incidence of calculi, antibiotic treatment failures and poor quality of life [16, 17]. They also show evidence of biofilm-forming bacterial infections [17]. Therefore, Shoskes et al. [17] initiated a preliminary complex treatment study with 16 patients using an antibiotic (500 mg tetracycline per day) plus a calcium chelator (1500 mg ethylenediaminetetraacetic acid, EDTA) and a proprietary vitamin-mineral nutraceutical and found significant decreases in the NIH Chronic Prostatitis Symptom Index (NIH-CPSI) in most patients and decreases in prostatic calcium-containing stones (calculi) in one-half of the patients examined.

Toxic heavy metals, such as mercury, lead, cadmium, nickel and others [19,20], can also be chelated with EDTA, and in some cases these heavy metals are carcinogenic to prostate tissue [21, 22]. Their removal from prostate tissue along with excess calcium can be accomplished with long-term EDTA administration. Thus we initiated a study based on the work of Shoskes et al. [17] to test the hypothesis that combinations of oral tetracycline and a patented, proprietary suppository (Detoxamin®) containing CaNa_2EDTA that is known to produce a long tissue half-life of EDTA in rats [23] would decrease chronic prostatitis symptoms along with cation deposits and increase quality of life.

Patients and methods

Patients

Criteria for inclusion in the study included: men aged 40 years or older (N=31, mean age=61, range 41-73) with a diagnosis of chronic prostatitis and the presence of prostate calcifications on ultrasound, absence of painful pelvic side wall spasm on rectal palpation, and the absence of allergy to tetracycline. Exclusionary criteria included: chronic debilitating condition other than prostatitis, such as chronic inflammatory or irritable bowel disease, chronic diarrhea or constipation, tetracycline allergy, cognitive decline (unable to comprehend instructions and answer questions), renal or hepatic dysfunction, inability to tolerate anal suppositories and absence of prostate calcifications. Of the participants, all but 7 showed evidence of prostate hyperplasia, varying from mild (48%) to moderate or severe (29%) as assessed by prostate sonogram. Participants were entered into an open label treatment trial where laboratory and clinical data were collected before and after treatment. Prior to entry all patients had a complete medical history and urological examination.

Laboratory/Clinical Tests

The following laboratory/clinical tests were performed pre- and post-treatment: non-fasting comprehensive blood chemistry and lipid panel (Southern California Reference Laboratory), Tustin, CA), blood analysis for heavy metals and essential minerals, fecal metals analysis (Doctors Data, Inc., St. Charles, IL), digital rectal exam and prostate sonogram using a Fukuda Denshi Model FF sonic UF-750XT Power Color Doppler (PCD) with 3D imaging capability. Analysis of elements was performed by ICP-Mass Spectroscopy following digestion of the specimen in a closed microwave system. For a given mineral element, these procedures measure the sum of the amounts of surface-adhering and intracellular content, regardless of chemical form. Participants also took the NIH-CPSI [24], International Prostate Symptom Score (IPSS) [25] and Erectile Function Index (EFI) [26] surveys.

Study Design

The study received Institutional Review Board (IRB) approval, and before admission each patient signed an informed consent document. Participants in the study were examined and tested (as above) before and after three months administration of tetracycline (HPN Pharmaceuticals, Torrance, CA; 500 mg PO daily) and use of Detoxamin® (World Health Products, Draper, Utah) time-release suppositories (4-times per week) each containing 750 mg disodium calcium EDTA (CaNa_2EDTA). Each day during the trial participants took two tabs of a multi-vitamin, mineral and trace mineral supplement (Health Genesis, World Health Products) to replace essential minerals and one capsule of a probiotic mixture in an oil matrix capsule (stored refrigerated; Healthy Trinity, Natren, Westlake Village, CA) to replace depleted gut flora [27,28]. At the end of therapy laboratory and clinical tests and clinical surveys were completed as described above and compared to pre-treatment tests and surveys.

Statistical analyses

Post-pre measurements were analyzed by paired t-test comparisons with Bonferroni corrections. Reliability was evaluated by Cronbach's alpha coefficient. In some cases parametric statistical results were checked by nonparametric Wilcoxon tests.

Results

Blood and fecal elements

A panel of mineral element tests were performed on packed red blood cells pre/post treatment and compared among participants. Post-treatment analyses indicated that there were significant increases in mean red blood cell copper, boron, molybdenum and magnesium levels and a borderline significant increase in calcium and decreases in arsenic, cadmium and lead, suggesting that the CaNa₂EDTA was mobilizing some tissue elements during the trial (Table 1). Analysis of fecal metals indicated that mean copper, tungsten and to a lesser degree beryllium levels were significantly increased post-treatment, whereas cadmium was significantly decreased (Table 2).

Blood Serum Chemistries

The non-fasting comprehensive blood chemistry panel showed the cholesterol/HDL ratio was significantly decreased ($p < 0.0005$) post-treatment, but no significant changes in creatinine, BUN/creatinine ratio or calcium levels were observed (Table 3).

Chronic prostatitis symptom index (NIH-CPSI)

Examination of the NIH-CPSI domains indicated that there were significant decreases in mean Urinary Symptom Score, mean Pain Symptom Score and mean Quality of Life Score post-treatment (Table 4). The mean Urinary Symptom Score post-treatment decreased from a pre-treatment mean of 3.709 ± 2.08 to a mean of 2.84 ± 2.25 , a mean decrease of 0.87 ± 1.99 post-treatment ($p < 0.0106$), whereas the pain symptom mean score decreased from 4.35 ± 4.54 to 3.10 ± 3.67 , a mean decrease of 1.25 ± 2.95 ($p < 0.0122$). The mean quality of life index score decreased from 4.68 ± 2.81 to a mean of 3.45 ± 2.53 , a mean decrease of 1.23 ± 2.22 post-treatment ($p < 0.0022$). Overall the NIH-CPSI mean total scores decreased from 12.74 ± 8.31 to 9.39 ± 6.16 , a mean decrease of 3.35 ± 0.93 ($p < 0.0006$).

International prostate symptom score (IPSS)

Using the International Prostate Symptom Score (IPSS) survey form indicated significant reductions in 5 out of 7 categories and significant reduction of overall symptoms post-compared to pre-treatment (Table 5). Significant reductions were found in incomplete

emptying of bladder ($p < 0.0082$), frequency of urination ($P < 0.0044$), urgency of urination ($p < 0.00001$), and intermittency of urination ($P < 0.0741$), weak stream ($p < 0.0003$), quality of life score ($p < 0.0191$) and overall score ($p < 0.0001$) (Table 5). Non-significant reductions were found in urination straining and nocturia (Table 5).

Erectile function index (EFI)

Using the EFI survey scores on 7 of 15 questions showed positive changes, indicating improvement in self-evaluated conditions, whereas 8 of 15 showed negative changes (Table 6). Thus overall there was not a significant overall change in EFI scores. The lack of significant improvement was confirmed by use of nonparametric Wilcoxon tests.

PCD sonography

Transrectal Power Color Doppler (PCD) sonography showed that there were no significant decreases in prostatic calcifications at the treatment levels used in this study (data not shown).

Discussion

Similar to Shoskes et al. [17] we found that patients with NIH Category III chronic prostatitis could be successfully treated in an open label trial with tetracycline (500 mg/day) and Detoxamin® suppositories containing 750 mg of CaNa_2EDTA used 4-times per week for 90 days. In the present study the dose of CaNa_2EDTA was half of the amount used by Shoskes et al. [17]. Using the NIH-CPSI and IPSS survey forms significant post-treatment reductions in mean index of prostatitis symptoms and pain were found along with significant improvements in mean quality of life and mean overall scores. The IPSS is also used as a validated instrument for benign prostatic hyperplasia [29], suggesting the positive influence of symptom reduction for this condition as well as prostatitis. Approximately 77% of the patients in our study had benign prostatic hyperplasia or cancerous prostate lesions.

We did not see a significant increase in mean overall Erectile Function Index post-treatment; however, in 7 of 15 questions there were significant improvements. Since this patient group is generally unresponsive to multiple courses of antibiotics alone and other therapies, we agree with Shoskes et al. [17] that this approach represents a significant improvement in therapy of Category III chronic prostatitis. As in Shoskes et al. [17], this open label study is

limited in terms of the number of patients entered and lack of blinded control arms, and thus no conclusions can be drawn as to mechanism of action and durability of the effect.

In our study, Detoxamin® CaNa₂EDTA suppositories were used to loosen and remove calcium and heavy metals from patients with established prostatic calcifications and chronic prostatitis while on tetracycline. That some metals and calcium were being mobilized by the CaNa₂EDTA suppositories was shown in blood and fecal tests. In particular, the mean levels of copper, boron, molybdenum, magnesium and calcium were significantly increased in packed red blood cells as well as significant decreases in arsenic, cadmium and lead in fecal matter in the post-treatment tests, suggesting that tissue stores of these elements were being affected. The presence of heavy metals and cations like calcium varies widely in different patients, and we found differences among patients in the amounts of these elements present in blood and stool when CaNa₂EDTA suppositories were used. Since approximately 77% of the patients in our study had a diagnosis of benign or cancerous prostate lesions, the removal of potentially carcinogenic heavy metals was a desirable effect of the therapy. Certain heavy metals, such as arsenic, cadmium, chromium and lead, among others, are known have carcinogenic potential, and their long-term exposure is associated with an increased incidence of prostate and other cancers [30,31]. However, the potential role of heavy metals in the pathogenesis and progression of already established cancers and in affecting the symptoms of chronic prostatitis remain uncertain.

We previously studied the pharmacokinetics and tissue concentrations (bioavailability) of Detoxamin® CaNa₂EDTA and compared it to IV administration of CaNa₂EDTA [23]. We found that there were substantially higher tissue-to-blood concentration ratios of EDTA via the suppository route compared to IV administration. Prostate tissues were found to have 8 hr mean tissue-to-blood ratios of 13.6 (rectal suppository) and 3.69 (IV), respectively [23]. This finding supports the mechanism of action of CaNa₂EDTA chelation suppositories on the target tissue in question (prostate) and was an initial justification for this clinical trial.

Another aspect of the use of CaNa₂EDTA suppositories is that bacterial biofilms contain high amounts of divalent cations like calcium that are important in maintaining their structures [32]. The presence of biofilm-producing bacteria has been associated with chronic prostatitis [14,17,32], and the use of calcium supplements is associated with increased risk of urinary

tract infections [33]. Calcium is also important in bacterial adhesion element in the urinary tract either as an adhesion promoting cation or inhibitor of secreted host glycoproteins that prevent bacterial adhesion to the urinary tract [34]. Thus removal of excess calcium ion and calcium complexed to extracellular bacterial proteoglycans and glycosylaminoglycans could be important in reducing bacterial adherence and chronic prostatitis symptoms.

There is a rather long history of the use of EDTA in clinical studies [35-39]. Animal experiments have shown that EDTA is extremely safe and relatively high doses are well tolerated without noticeable side effects [40,41]. However, in a few reports a few subjects receiving IV EDTA had reversible renal damage [42,43], and temporary numbness and tingling in IV administered EDTA are relatively common [44]. None of these adverse effects were found in the current study using Detoxamin® CaNa₂EDTA suppositories. The Detoxamin® CaNa₂EDTA suppositories were found to be well tolerated and safe. Interestingly, although not the focus of this study, we found that mean HDL and LDL lipid blood panel determinations showed significant improvements post-treatment. We and others [17] have concluded that the addition of EDTA suppositories to antibiotic therapy of chronic prostatitis significantly reduced symptoms and improved outcome.

Table 1. Red Blood Cell Elements Pre-Post Treatment Comparisons*

| Element | Pre Treatment | SD | Post Treatment | SD | Mean Change | SD | Min | Max | Test Statistic | p > t | p > t | p < t |
|------------|---------------|--------|----------------|--------|-------------|--------|---------|--------|----------------|--------|--------|--------|
| Lead | 0.0286 | 0.0122 | 0.0214 | 0.0091 | -0.0073 | 0.0056 | -0.023 | 0.004 | -7.23 | 0.0000 | 1.0000 | 0.0000 |
| Copper | 0.5667 | 0.0457 | 0.6035 | 0.0504 | 0.0358 | 0.0387 | -0.05 | 0.1 | 5.15 | 0.0000 | 0.0000 | 1.0000 |
| Boron | 0.0534 | 0.0282 | 0.0676 | 0.0354 | 0.0142 | 0.0223 | -0.03 | 0.064 | 3.56 | 0.0013 | 0.0006 | 0.9994 |
| Molybdenum | 0.0009 | 0.0002 | 0.0012 | 0.0003 | 0.0003 | 0.0004 | -0.0007 | 0.0011 | 3.53 | 0.0014 | 0.0007 | 0.9993 |
| Magnesium | 43.9677 | 3.7281 | 45.7097 | 3.6805 | 1.7419 | 2.8162 | -3 | 7 | 3.44 | 0.0017 | 0.0009 | 0.9991 |
| Cadmium | 0.0011 | 0.0002 | 0.0010 | 0.0001 | -0.0001 | 0.0003 | -0.001 | 0 | -2.42 | 0.0217 | 0.9892 | 0.0108 |
| Arsenic | 0.0051 | 0.0041 | 0.0044 | 0.0029 | -0.0007 | 0.0019 | -0.006 | 0.003 | -2.12 | 0.0425 | 0.9787 | 0.0213 |
| Zinc | 11.6903 | 0.8931 | 11.4742 | 0.9518 | -0.2161 | 0.5734 | -1.6 | 1.2 | -2.10 | 0.0444 | 0.9778 | 0.0222 |
| Calcium | 12.2258 | 1.9615 | 13.1613 | 1.6752 | 0.9355 | 2.5682 | -7 | 6 | 2.03 | 0.0515 | 0.0258 | 0.9742 |
| Potassium† | 78.3871 | 2.5518 | 79.1290 | 2.7418 | 0.7419 | 2.6073 | -4 | 5 | 1.58 | 0.1236 | 0.0618 | 0.9382 |
| Mercury | 0.0051 | 0.0052 | 0.0043 | 0.0040 | -0.0007 | 0.0027 | -0.009 | 0.005 | -1.53 | 0.1358 | 0.9321 | 0.0679 |
| Iron | 934.838 | 38.827 | 943.483 | 32.289 | 8.6452 | 32.701 | -58 | 67 | 1.47 | 0.1515 | 0.0757 | 0.9243 |
| Manganese | 0.0152 | 0.0049 | 0.0148 | 0.0044 | -0.0003 | 0.0019 | -0.006 | 0.002 | -0.96 | 0.3442 | 0.8279 | 0.1721 |
| Vanadium | 0.0002 | 0.0000 | 0.0002 | 0.0001 | 0.0000 | 0.0001 | -0.0002 | 0.0003 | 0.57 | 0.5722 | 0.2861 | 0.7139 |
| Selenium | 0.3632 | 0.2145 | 0.3571 | 0.1650 | -0.0061 | 0.0838 | -0.34 | 0.09 | -0.41 | 0.6868 | 0.6566 | 0.3434 |
| Phosphorus | 567.903 | 31.442 | 569.096 | 23.873 | 1.1935 | 22.983 | -48 | 61 | 0.29 | 0.7745 | 0.3872 | 0.6128 |
| Chromium | 0.0008 | 0.0004 | 0.0008 | 0.0007 | 0.0000 | 0.0008 | -0.0017 | 0.0035 | 0.24 | 0.8107 | 0.4053 | 0.5947 |
| Thallium | 0.0001 | 0.0000 | 0.0001 | 0.0000 | 0.0000 | 0.0000 | 0 | 0 | 1.00 | 1.0000 | 1.0000 | 1.0000 |

*Results reported as mg/g or ppm

†Results reported as mEq/g

| Table 2. Fecal Elements Pre-Post Treatment Comparisons* | | | | | | | | | | | | |
|---|---------------|--------|----------------|--------|-------------|--------|--------|-------|----------------|--------|--------|--------|
| Element | Pre Treatment | SD | Post Treatment | SD | Mean Change | SD | Min | Max | Test Statistic | p > t | p > t | p < t |
| | | | | | | | | | | | | |
| Copper | 43.710 | 14.272 | 59.000 | 17.920 | 15.290 | 20.980 | -25 | 67 | 4.06 | 0.0003 | 0.0002 | 0.9998 |
| Cadmium | 0.557 | 0.354 | 0.371 | 0.179 | -0.186 | 0.301 | -1.03 | 0.19 | -3.45 | 0.0017 | 0.9992 | 0.0008 |
| Tungston | 0.093 | 0.060 | 0.151 | 0.092 | 0.058 | 0.109 | -0.261 | 0.228 | 2.99 | 0.0055 | 0.0028 | 0.9972 |
| Beryllium | 0.006 | 0.005 | 0.008 | 0.007 | 0.003 | 0.008 | -0.022 | 0.025 | 1.82 | 0.0791 | 0.0395 | 0.9605 |
| Antimony | 0.062 | 0.037 | 0.047 | 0.033 | -0.015 | 0.048 | -0.112 | 0.104 | -1.72 | 0.0956 | 0.9522 | 0.0478 |
| Nickel | 5.771 | 2.774 | 6.803 | 2.803 | 1.032 | 3.554 | -6.9 | 10.1 | 1.62 | 0.1164 | 0.0582 | 0.9418 |
| Thallium | 0.018 | 0.014 | 0.015 | 0.005 | -0.003 | 0.011 | -0.049 | 0.013 | -1.61 | 0.1189 | 0.9405 | 0.0595 |
| Arsenic | 0.219 | 0.139 | 0.189 | 0.104 | -0.030 | 0.105 | -0.41 | 0.11 | -1.59 | 0.1234 | 0.9383 | 0.0617 |
| Bismuth | 0.070 | 0.197 | 0.207 | 0.676 | 0.137 | 0.708 | -0.988 | 3.537 | 1.08 | 0.2900 | 0.1450 | 0.8550 |
| Platinum | 0.001 | 0.001 | 0.032 | 0.167 | 0.031 | 0.167 | 0 | 0.93 | 1.03 | 0.3092 | 0.1546 | 0.8454 |
| Mercury | 0.180 | 0.288 | 0.210 | 0.377 | 0.031 | 0.167 | -0.3 | 0.796 | 1.02 | 0.3167 | 0.1583 | 0.8417 |
| Lead | 0.247 | 0.145 | 0.270 | 0.136 | 0.023 | 0.170 | -0.54 | 0.41 | 0.75 | 0.4586 | 0.2293 | 0.7707 |
| Uranium | 0.115 | 0.091 | 0.110 | 0.101 | -0.005 | 0.102 | -0.204 | 0.372 | -0.27 | 0.7861 | 0.6069 | 0.3931 |

*Results reported as mg/Kg dry weight of feces

| Table 3. Comprehensive Blood Chemistry Panels Pre-Post Treatment Comparisons | | | | | | | | |
|--|-------------|--------|-------|--------|----------------|--------|--------|--------|
| Parameter | Mean Change | SD | Min | Max | Test Statistic | p > t | p > t | p < t |
| SCRL SGOT | 4.226 | 4.287 | -8 | 13 | 5.49 | 0.0000 | 0.0000 | 1.0000 |
| SCRL HDL CHOL | 5.903 | 6.764 | -3 | 20 | 4.86 | 0.0000 | 0.0000 | 1.0000 |
| SCRL SGPT | 3.097 | 4.077 | -10 | 10 | 4.23 | 0.0002 | 0.0001 | 0.9999 |
| SCRL URIC ACID | -0.477 | 0.644 | -1.9 | 1.4 | -4.13 | 0.0003 | 0.9999 | 0.0001 |
| SCRL CO2 | 1.645 | 2.259 | -3 | 6 | 4.05 | 0.0003 | 0.0002 | 0.9998 |
| SCRL CHOL/HDL | -0.455 | 0.653 | -3.1 | 0.9 | -3.88 | 0.0005 | 0.9997 | 0.0003 |
| SCRL BILIRUBIN | 0.129 | 0.218 | -0.2 | 0.7 | 3.30 | 0.0025 | 0.0013 | 0.9987 |
| SCRL POTASSIUM | -0.410 | 0.734 | -1.6 | 1.3 | -3.11 | 0.0041 | 0.9979 | 0.0021 |
| SCRL LDL CHOL | -14.290 | 26.768 | -78 | 56 | -2.97 | 0.0058 | 0.9971 | 0.0029 |
| SCRL CHLORIDE | -1.290 | 2.636 | -7 | 3 | -2.73 | 0.0106 | 0.9947 | 0.0053 |
| SCRL ALK PHOSPHATE | 4.032 | 9.318 | -16 | 32 | 2.41 | 0.0223 | 0.0112 | 0.9888 |
| SCRL A/G RATIO | -0.058 | 0.139 | -0.3 | 0.2 | -2.33 | 0.0265 | 0.9868 | 0.0132 |
| SCRL SODIUM | -1.065 | 2.756 | -7 | 5 | -2.15 | 0.0397 | 0.9802 | 0.0198 |
| SCRL GLOBULIN | 0.081 | 0.227 | -0.5 | 0.5 | 1.98 | 0.0574 | 0.0287 | 0.9713 |
| SCRL LPPLA2 | -10.774 | 30.737 | -71 | 60 | -1.95 | 0.0604 | 0.9698 | 0.0302 |
| SCRL CREATININE | 0.029 | 0.086 | -0.1 | 0.2 | 1.87 | 0.0711 | 0.0355 | 0.9645 |
| SCRL PSA TOTAL | 7.079 | 21.103 | -4.34 | 107.22 | 1.87 | 0.0716 | 0.0358 | 0.9642 |
| SCRL BUN/CREAT RATIO | 0.855 | 3.870 | -5.6 | 13.3 | 1.81 | 0.0803 | 0.0402 | 0.9598 |
| SCRL GLUCOSE | 8.000 | 24.596 | -25 | 64 | 1.78 | 0.0853 | 0.0427 | 0.9573 |
| SCRL CHOL | -8.097 | 32.232 | -99 | 65 | -1.40 | 0.1722 | 0.9139 | 0.0861 |
| SCRL GGPT | 0.581 | 2.391 | -4 | 9 | 1.35 | 0.1865 | 0.0932 | 0.9068 |
| SCRL BUN | 1.355 | 4.168 | -6 | 16 | 1.23 | 0.2283 | 0.1141 | 0.8859 |
| SCRL HS CRP | -1.074 | 5.007 | -27.7 | 0.7 | -1.19 | 0.2416 | 0.8792 | 0.1208 |
| SCRL CALCIUM | 0.065 | 0.316 | -0.6 | 1 | 1.14 | 0.2643 | 0.1321 | 0.8679 |
| SCRL VIT D 25 | -2.226 | 15.836 | -31 | 52 | -0.78 | 0.4400 | 0.7800 | 0.2200 |
| SCRL TOTAL PROTEIN | -0.161 | 1.310 | -7 | 0.7 | -0.69 | 0.4982 | 0.7509 | 0.2491 |
| SCRL SCORE ALBUMIN | -0.016 | 0.163 | -0.3 | 0.4 | -0.55 | 0.5869 | 0.7066 | 0.2934 |
| SCRL VLDL CHOL | 1.258 | 14.799 | -32 | 53 | 0.47 | 0.6394 | 0.3197 | 0.6803 |
| SCRL TRIGLYCERIDES | 5.581 | 74.359 | -163 | 269 | 0.42 | 0.6790 | 0.3395 | 0.6605 |

| Table 4. NIH-CPSI (Chronic Pain Symptom Index) Scores Pre-Post Treatment Comparison | | | | | | | | | | |
|--|----------------------|-------|-----------------------|-------|----------------|--------|-------------------|--------|--------|--------|
| Parameter | Pre Treatmen t | SD | Post Treatmen t | SD | Mean Change | SD | Test Statistic | p > t | p > 1 | p < 1 |
| Urinary Symptom Score | 3.7097 | 2.085 | 2.839 | 2.225 | -0.871 | 1.9957 | -2.43 | 0.0213 | 0.9894 | 0.0106 |
| Pain Symptom Score | 4.355 | 4.542 | 3.097 | 3.673 | -1.258 | 2.9549 | -2.37 | 0.0244 | 0.9878 | 0.0122 |
| QOL Score | 4.677 | 2.809 | 3.452 | 2.253 | -1.226 | 2.2167 | -3.08 | 0.0044 | 0.9978 | 0.0022 |
| | | | | | | | | | | |
| Total Score | 12.742 | 8.306 | 9.387 | 6.157 | -3.355 | 0.9312 | -3.60 | 0.0011 | 0.9994 | 0.0006 |

| Table 5. International Prostate Symptom Scores (IPSS) Matched pair t-tests for each question, quality of life and total score | | | | | | | |
|--|----------|--------------|----------------|-------------------|--------|--------|--------|
| Parameter | Pre Mean | Post Mean | Mean Change | Test Statistic | p > t | p > t | p < t |
| Incomplete Emptying | 1.677 | 0.968 | -0.710 | -2.832 | 0.0082 | 0.9959 | 0.0041 |
| Frequency | 2.581 | 1.839 | -0.742 | -3.081 | 0.0044 | 0.9978 | 0.0022 |
| Intermittency | 1.645 | 1.065 | -0.581 | -1.851 | 0.0741 | 0.9630 | 0.0370 |
| Urgency | 2.290 | 0.968 | -1.323 | -5.261 | 0.0000 | 1.0000 | 0.0000 |
| Weak Stream | 2.387 | 1.419 | -0.968 | -4.055 | 0.0003 | 0.9998 | 0.0002 |
| Straining | 0.774 | 0.516 | -0.258 | -1.092 | 0.2835 | 0.8582 | 0.1418 |
| Nocturia | 1.871 | 1.597 | -0.274 | -1.516 | 0.1400 | 0.9300 | 0.0700 |
| | | | | | | | |
| QOL Score | 2.774 | 2.129 | -0.645 | -2.477 | 0.0191 | 0.9904 | 0.0096 |
| Total Score | 13.226 | 8.371 | -4.855 | -4.526 | 0.0001 | 1.0000 | 0.0000 |

| Table 6. Erectile Function Index (EFI) Scores Matched pair t-tests for each question and EFI total score | | | | | | | |
|---|----------|-----------|-------------|----------------|--------|-------|-------|
| Question | Pre Mean | Post Mean | Mean Change | Test Statistic | p > t | p > t | p < t |
| EF1 | 4.35 | 3.85 | -0.50 | -2.125 | 0.042 | 0.979 | 0.021 |
| EF2 | 4.23 | 3.97 | -0.26 | -1.393 | 0.174 | 0.913 | 0.087 |
| EF3 | 3.84 | 3.63 | -0.21 | -0.806 | 0.426 | 0.787 | 0.213 |
| EF4 | 3.56 | 3.47 | -0.10 | -0.390 | 0.699 | 0.650 | 0.350 |
| EF5 | 3.48 | 3.65 | 0.16 | 0.579 | 0.567 | 0.283 | 0.717 |
| EF6 | 2.19 | 3.13 | 0.94 | 2.799 | 0.009 | 0.004 | 0.996 |
| EF7 | 3.55 | 3.52 | -0.03 | -0.133 | 0.895 | 0.552 | 0.448 |
| EF8 | 3.48 | 3.48 | 0.00 | 0.000 | 1.000 | 0.500 | 0.500 |
| EF9 | 4.05 | 3.77 | -0.27 | -0.902 | 0.374 | 0.813 | 0.187 |
| EF10 | 4.31 | 3.87 | -0.44 | -1.510 | 0.142 | 0.929 | 0.071 |
| EF11 | 4.29 | 4.10 | -0.19 | -0.828 | 0.414 | 0.793 | 0.207 |
| EF12 | 3.53 | 3.68 | 0.15 | 1.055 | 0.300 | 0.150 | 0.850 |
| EF13 | 3.65 | 3.94 | 0.29 | 1.103 | 0.279 | 0.139 | 0.861 |
| EF14 | 3.60 | 4.00 | 0.40 | 1.388 | 0.175 | 0.088 | 0.912 |
| EF15 | 3.69 | 3.87 | 0.18 | 1.232 | 0.227 | 0.114 | 0.886 |
| EF Total Score | 55.69 | 55.79 | 0.10 | 0.037 | 0.971 | 0.485 | 0.515 |

References

1. Domingue GR, Hellstrom WJG. Prostatitis. Clin Microbiol Rev 1998; 11:604-613.
2. Habermacher GM, Chason JT, Schaeffer AJ. Prostatitis/chronic pelvic pain syndrome. Annu Rev Med 2006; 57:195-206.
3. Krieger JN, Ross SO, Deutsch L, Riley DE. The NIH Consensus concept of chronic prostatitis/chronic pelvic pain syndrome compared with traditional concepts of nonbacterial prostatitis and prostatodynia. Curr Urol Rep 2002; 3:301-6.
4. Domingue GJ. Cryptic bacterial infection in chronic prostatitis: diagnostic and therapeutic implications. Curr Opin Urol 1998; 8:45-49.
5. Beckman TJ, Edson RS. Methicillin-resistant Staphylococcus aureus prostatitis. Urology 2007; 69:779-783.
6. Skerk V, Krhen I, Schonwald S, Cajic V, Markovinovic L, Roglic S, Zekan S, Andracevic AT, Kruzic V. The role of unusual pathogens in prostatitis syndrome. Int J Antimicrob Agents 2004; 24 (Suppl 1): S53-S56.
7. Mandar R, Raukas E, Turk A, Korrovitis P, Punab M. Mycoplasmas in semen of chronic prostatitis patients. Scand J Urol Nephrol 2005; 39:479-482.
8. Wood HM, Shoskes DA. The role of nanobacteria in urologic disease. World J Urol 2006; 24:51-54.
9. Kanamaru S, Kurazono H, Terai A, Monden K, Kumon K, Kumon H, Mizunoe Y, Ogawa O, Yamamoto S. Increased biofilm formation in Escherichia coli isolated from acute prostatitis. Int J Antimicrob Agents 2006; 28(Suppl 1):S21-S25.
10. Soto SM, Smithson A, Martinez JA, Horcajada JP, Mensa J, Vila J. Biofilm formation in uropathogenic Escherichia coli strains: relationship with prostatitis, urovirulence factors and antimicrobial resistance. J Urol 2007; 177:365-368.
11. Tunuguntla HSGR, Evans CP. Management of prostatitis. Pros Cancer Pros Dis 2002; 5:172-179.
12. van Houdt R, Michiels CW. Role of bacterial cell surface structures in Escherichia coli biofilm formation. Res Microbiol 2005; 156:626-633.
13. Reisner A, Krogfelt KA, Klein BM, Zechner EL, Molin S. In vitro biofilm formation of commensal and pathogenic Escherichia coli strains: impact of environmental and genetic factors. J Bacteriol 2006;

- 188:3572-3581.
14. Parsek MR, Siingh PK. Bacterial biofilms: an emerging link to disease pathogenesis. *Annu Rev Microbiol* 2003; 57:677-701.
15. Shoskes DA, Hakin L, Ghoniem G, Jackson CL. Long-term results of multimodal therapy for chronic prostatitis/ chronic pelvic pain syndrome. *J Urol* 2003; 169:1406-1410.
16. Rugendorff EW, Weidner W, Ebeling L, Buck AC. Results of treatment with pollen extract (Cernilton N) in chronic prostatitis and prostatodynia. *Br J Urol* 1993; 71:433-438.
17. Shoskes DA, Thomas KD, Gomez E. Anti-nonobacterial therapy for men with chronic chronic prostatitis/chronic pelvic pain syndrome and prostatic stones: preliminary experience. *J Urol* 2005; 173:474-477.
18. Geramoutsos I, Gyftopoulos K, Perimenis P, Thanou V, Liagka D, Stamblis D, Barbalias G. Clinical correlation of prostatic lithiasis with chronic pelvic pain syndromes in young adults. *Eur Urol* 2004; 45:333-337.
19. Lippmann M, Thurston GD. Exposure assessment: input into risk assessment. *Arch Environ Health* 1988; 43:113-123.
20. Goyer RA. Nutrition and metal toxicity. *Am J Clin Nutr* 1995; 61(Suppl 3):646S-650S.
21. Magos L. Epidemiological and experimental aspects of metal carcinogenesis: physiochemical properties, kinetics and active species. *Environ Health Persp* 1991; 95:157-189.
22. Piscator M. Role of Cadmium in carcinogenesis with special reference to cancer of the prostate. *Environ Health Persp* 1981; 40:107-120.
23. Ellithorpe R, Mazur P, Gum G, Button G, Pfadenhauer EH, Settineri RA, Nicolson GL. Comparison of the absorption, brain and prostate distribution and elimination of CaNa₂EDTA of rectal chelation suppositories to intravenous administration. *J Am Nutricutical Assoc* 2007; 10: in press.
24. Probert KJ, Litwin MS, Wang Y, Alexander RB, Calhoun E, Nickel JC, O'Leary MP, Pontari M, McNaughton-Collins M. Responsiveness of the National Institutes of Health Chronic Prostatitis Symptom Index (NIH-CPSI). *Qual Life Res*. 2006; 15:299-305.
25. Rosen RC, Riley A, Wagner G, Osterloh IH, Kirkpatrick J, Mishra A. The international index of erectile function (IIEF): a multidimensional scale for assessment of erectile dysfunction. *Urology*. 1997; 49:822-830.
26. Barry MJ, Fowler FJ Jr, O'Leary MP, Bruskewitz RC, Holtgrewe HL, Mebust WK, Cockett AT. The American Urological Association symptom index for benign prostatic hyperplasia. The Measurement Committee of the American Urological Association. *J Urol* 1992; 148:1549-1557.
27. Madden JA, Plummer SF, Tang J, Garaiova I, Plummer NT, Herbison M, Hunter JO, Shimada T, Cheng L, Shirakawa T. Effect of probiotics on preventing disruption of the intestinal microflora following antibiotic therapy: a double-blind, placebo-controlled pilot study. *Int Immunopharmacol*. 2005; 5:1091-1097.
28. Bondarenko VM, Lykova EA, Matsulevich TV. Microecological aspects of small intestinal bacterial overgrowth syndrome. *Zh Mikrobiol Epidemiol Immunobiol*. 2006; 6:57-63.
29. Madsen FA, Bruskewitz RC. Clinical manifestations of benign prostatic hyperplasia. *Urol Clin North Am* 1995; 22:291-298.
30. Doll R, Fishbein L, Infante P, Landrigan P, Lloyd JW, Mason TJ, Mastromatteo E, Norseth T et al. Problems of epidemiological evidence. *Environ Health Perspect* 1981; 40:11-20.
31. Kazantzis G. Role of cobalt, iron, lead, manganese, mercury, platinum, selenium and titanium in carcinogenesis. *Environ Health Perspect* 1981 40:143-161.
32. Jones BV, Mahenthiralingam E, Sabbuba NA, Stickler DJ. Role of swarming in the formation of crystalline *Proteus mirabilis* biofilms on urinary catheters. *J Med Microbiol* 2005; 54:807-813.
33. Apicella LL, Sobota AE. Increased urinary tract infection associated with the use of calcium supplements. *Urol Res* 1990; 18:213-217.
34. Sobota AE, Apicella LL. Reduction in the anti-adherence activity of Tamm-Horsfall protein with increasing concentration of calcium. *Urol Res* 1991; 19:177-180.
35. Belknap EL. EDTA in the treatment of lead poisoning. *Indust Med Surg* 1952; 21:305-306.
36. Mosher LE, McCann DS. Some in vivo effects of chelation-I: Rheumatoid arthritis. *J Chronic Dis* 1963; 16:325-328.
37. Chappell LT, Wilson J. Chelation therapy for vascular disease. *Circulation*. 1999; 99:164-165.
38. Clarke NE Sr. Atherosclerosis, occlusive vascular disease and EDTA. *Am J Cardiol* 1960; 6:233-236.
39. Chisolm JJ Jr: The use of chelating agents in the treatment of acute and chronic lead intoxication in childhood. *J Pediat* 1968; 73:1-38.
40. Castellino N, Aloj S. Effects of calcium sodium ethylenediaminetetra-acetate on the kinetics of the distribution and excretion of lead in the rat. *Brit J Indust Med* 1965; 22:172-178.
41. Aronson AL, Hammond PB, Strauss AC. Studies with calcium ethylenediaminetetraacetate in calves;

- toxicity and use in bovine lead poisoning. *Toxicol Appl Pharmacol* 1968; 12:337-349.
42. Betieman V, Landy E, Moseka JK and Wedsen RP. Contribution of lead hypertension with renal impairment. *N Engl J. Med.* 1983; 309:17-21.
 43. Mc Dragh, EW, Rudolph DJ, Cherskin E. The effect of EDTA chelation therapy plus supportive multi-vitamin trace mineral supplementation upon renal function: as study on serum creatinine. *J. Holistic Med* 1982; 4:146-151.
 44. Chen IW, Park HM, King LR, Bahr GK, Goldsmith RE. Radioimmunoassay of parathyroid hormone: peripheral plasma immunoreactive parathyroid hormone response to ethylenediaminetetraacetate. *J Nucl Med* 1974; 15:763-769.

Effects of CaNa₂ EDTA (Detoxamin[®]) Suppositories on Excretion of Heavy Metals

Living Longer Institute, Cincinnati, Ohio

Pelletier, M., MD, CCN, FACOG, Lavalley, J., ND, RPh, CCN, Ellithorpe, R., MD*, Schmidt, M., PhD, CCN, Pelton, R., RPh, PhD, CCN, Settineri, R., MS

Abstract

A formidable amount of data exists that clearly indicate the insidious toxicity of non-physiological metals such as mercury, lead, nickel, cadmium, arsenic and aluminum where specific mechanisms for the neurotoxic, nephrotoxic and immune-dysregulatory effects of these metals are identified. A major portion of the population is at risk for chronic, low-level exposure to toxic heavy metals from environmental and occupational sources, as well as dental materials. This clinical interim clinical pilot trial has evaluated the safety and efficacy of a recently developed suppository delivery of CaNa₂ EDTA. Suppositories provide direct access to the systemic circulation, efficiently bypassing portal circulation and liver metabolism via the hemorrhoidal veins on the first pass. The study examined the ability of CaNa₂ EDTA suppositories to remove a variety of the most prevalent toxic heavy metals as determined by excretion in urine and feces. Twelve healthy adult subjects, ranging from 25 to 65 years of age, were administered CaNa₂ EDTA suppositories over a course of 90 days. Excretory specimens were evaluated on 0 (pre-treatment), 3 and 90 days post-treatment. An initial provocation with DMSA was given orally several days before the Day 0 samples were taken and again on Day 90 post-treatment. Results showed significant excretion ($P < 0.05$) with cumulative CaNa₂ EDTA therapy of Day 3 and Day 90 compared to Day 0 with arsenic, lead, cadmium and nickel in feces samples as well as significant excretions ($P < 0.05$) with arsenic, lead, mercury, cadmium and nickel in urine samples. There were no differences in the safety profile comparisons within comprehensive metabolic chemistry panels between pre- and post-treatment blood values. Only minor transient complaints of loose stools and gas were reported in several subjects. Calcium disodium EDTA suppositories appear to be a safe and effective means to slowly and consistently remove a variety of toxic heavy metals as evidenced by urine and fecal analysis. There is increased need and demand for more consumer friendly, less invasive, less time-consuming broad specificity metal chelation protocols consistent with the time constraints and health goals of many members of our society today.

*** Presenter**

Introduction

A plethora of published biomedical studies clearly indicate the insidious toxicity of non-physiological metals such as mercury, lead, cadmium and aluminum, and the specific mechanisms for the neurotoxic, nephrotoxic and immune-dysregulatory effects of the metals have been well elucidated. To date, DMPS (iv and po) DMSA (po) and slow-drip $\text{Na}_2\text{-EDTA}$ have been appropriately evaluated for efficacy in both acute and chronic metal toxicity. The USEPA, FDA, CDC and State Health departments recognize the growing global problem of chronic low level exposure to toxic metals; however the prevailing criteria for initiation of treatment fall short of the more rigorous standards of those who practice preventative/comprehensive medicine versus crisis management. $\text{Na}_2\text{-EDTA}$ is FDA approved for lead detoxification (also chelates other metals), but the most common method of administration (slow drip intravenous method) is very expensive, invasive and far too time-consuming for most adults in our fast-paced society.

Adults are at risk for chronic, low-level exposure to toxic metals from environmental and occupational sources, as well as dental materials. Furthermore, aging is associated with increased risk for chronic, low-level *re-exposure* to lead because the vast majority of lead is sequestered in bone and dissolution of the bone matrix is a common problem with aging. Lead, released from the bone, where it is relatively inert, has a far greater adverse effects when it is subsequently taken up by extremely vulnerable cells in the central and peripheral nervous system, heart and kidneys. The bottom line: there is increased need and demand for more consumer friendly, less invasive, less time-consuming broad specificity metal chelation protocols that are consistent with the time constraints and health goals of many members of our society today.

Detoxamin $\text{CaNa}_2\text{-EDTA}$ Suppositories provide a safe and effective alternative to the expensive and invasive traditional slow drip EDTA protocol for metal detoxification. The fact that $\text{CaNa}_2\text{-EDTA}$ administered rectally might be an effective means for Hg detoxification is in sharp contrast to the relative inefficiency of traditional EDTA chelation, which is based upon *urinary excretion*.

Within the era of cost-containment and the risk of AIDS and other communicable blood-borne diseases, time constraints and affordability issues, suppository drug delivery is becoming a more viable option for doctors and patients. Suppositories provide direct access to the systemic circulation, efficiently bypassing the portal circulation and the liver metabolism on the first pass. It is a little known fact that the lower and middle hemorrhoidal veins bypass the liver and do not undergo first-pass metabolism. Therefore, suppositories can deliver the drug rapidly to the lower and middle hemorrhoidal veins for absorption. The rectum is an interesting area for drug absorption because it is not buffered and has a *neutral pH*. It also has very little enzymatic activity, thus enzymatic degradation does not occur. The rectal mucosa is more capable than the gastric mucosa of tolerating various drug-related irritations. This is especially important in patients with gastric disease. The anorectal physiology provides a

large surface area for drug absorption. Another factor that is important in drug delivery is drug solubility. The osmosis process allows the drug to transfer from the vehicle in the suppository, across the membrane of the rectum, and into the hemorrhoidal veins. As we become more aware of the potential complications of infection associated with the use of IVs, suppository administration provides a preferable alternative.

The body sequesters Hg in the liver by binding it to cholesterol and converting it into bile that then flows into the intestines. As bile is used to breakdown dietary lipids, some of the Hg becomes unbound in the intestinal tract. Detoxamin $\text{CaNa}_2\text{-EDTA}$ Suppositories deliver the chelation ability just where the maximum Hg excretion is taking place.

With traditional EDTA chelation, patients are limited to no more than two IV treatments a week because of the renal and hepatic toxicity of EDTA. Detoxamin $\text{CaNa}_2\text{-EDTA}$ Suppositories provide a much slower administering rate, which allows:

- The EDTA has more opportunities to bind to heavy metals. Through galvanic series of metals & alloys, the EDTA has greater opportunities to bind with heavy metals closer to the cathodic end of the series.
- The maximum amount of recommended EDTA in one week to be spaced out into nightly treatment, *greatly enhancing the chelation efficacy of Detoxamin $\text{CaNa}_2\text{-EDTA}$.*

With traditional IV EDTA chelation, physicians and nursing staff need to be constantly vigilant for blood serum calcium deficiency.

- Traditional EDTA IV lowers ionized plasma calcium during infusion. The body attempts to maintain homeostasis by producing an increase in circulating parathormone. The intermittent infusion period increases of parathormone caused by traditional EDTA IV infusion and have a profound negative effect on bone metabolism.
- Hypocalcaemic tetany is an inherent risk of traditional EDTA IV chelation. Medical staff need to be ever vigilant for warning signs of neuromuscular irritability and fasciculation that requires the administration of intravenous calcium, such as calcium gluconate or similar products, to reverse the potentially life threatening symptoms of hypocalcaemia.
- *There have been no reported hypocalcaemia episodes with Detoxamin $\text{CaNa}_2\text{-EDTA}$*

Rectal administration of Detoxamin $\text{CaNa}_2\text{-EDTA}$ suppositories has proven to be a safe and effective alternative to the traditional slow drip EDTA protocol, but currently there is no published data yet to address the efficacy of Hg chelation with Detoxamin $\text{CaNa}_2\text{-EDTA}$ Suppositories.

Objectives

Primary:

- To assess the effect of Detoxamin on mobilization of Pb, Hg, Cd, As, Fe, and other metals in humans as determined by blood, urine, feces, and hair.

Secondary:

- To assess the safety of Detoxamin as measured by symptom inventory, serum chemistry, blood lipids, and selected inflammatory markers in humans.
- To determine whether Detoxamin therapy is associated with alteration in markers of oxidative stress.
- To assess the variability in urinary excretion of selected metals from baseline to 90-day endpoint, following DMPS IV push, in non-treated human controls.

Experimental Design

1) Subjects.

Healthy adult (25-65 y.o.) human subjects will be identified at the Living Longer Institute. Subjects will be excluded from the study if they (a) have undergone placement or removal of amalgams within the past 3 months, (b) have undergone chelation therapy, (c) are exposed through occupation or hobbies to Hg, Pb, Cd, As, or other toxic metals, (d) have been diagnosed with diabetes, cancer, renal disease/insufficiency, biliary obstruction or liver disease (eg. hepatitis), or any other significant medical condition, (e) do not have regular bowel habits, (f) have chronic inflammatory bowel disease, diarrhea or constipation, (g) are pregnant, lactating or use diuretics, (h) are allergic to EDTA. The numbers of small, medium and large mercury amalgams should be recorded for each subject upon initial evaluation and, if possible, the number of mercury-filled root canals.

2) Study Population

The study will be conducted in up to 50 adult male and female subjects in generally good health. A single clinical investigational center will participate.

Subjects will be non-randomly assigned to one of two groups as designated below:

| | | |
|----------------|------|-----------------------|
| A ₀ | n=10 | Non-Treatment Control |
| A ₁ | n=40 | Detoxamin Suppository |

(2) Crucial Dietary Restrictions

After identifying subjects (n=50), informed consent forms will be signed and the subjects will adhere to the following dietary restrictions. The following are to be STRICTLY AVOIDED FOR AT LEAST 7 DAYS PRIOR TO THE BASELINE URINE AND FECAL SPECIMEN COLLECTIONS AND THROUGHOUT THE DURATION OF THE STUDY: any type of fish or seafood (including shellfish, seaweed), bentonite clay, Na-alginate, keratin, aluminum containing antacids, colonics, barium enemas, mineral or castor oil, colloidal minerals, activated charcoal, “chitosan-type” products, chlorella, cilantro, porphra-zyme or other chlorophyll-rich supplements/phytonutrients, products that contain EDTA or any other potential metal mobilization agent. Due to the potential lead contamination of some calcium supplements, Ca supplementation should be withheld unless absolutely necessary.

(3) Experimental Protocol

Before the administration of Detoxamin CaNa_2 -EDTA Suppositories, and after 7 days of adherence to dietary restrictions, the subjects will collect a 24-hr. urine specimen, a bowel movement, blood sample, and hair sample. The urine and fecal and hair samples will serve as the baseline to unprovoked toxic metal data. After completing the tests described above, the Detoxamin CaNa_2 -EDTA Suppositories will be administered. Continued adherence to dietary restrictions is imperative throughout the remainder of the sample collection period. Detoxamin CaNa_2 -750mg EDTA Suppositories will be taken at night by each subject, prior to bedtime for 90 days (total 90 suppositories). Subjects should eat at least 3 hours prior to bedtime to prevent potential discomfort. Patients should be encouraged to consume 2-3 liters of purified water per day. Patients are also taking a multi-vitamin, mineral and trace mineral supplement (Health-Genesis) to replace the essentials that are removed during therapy.

(4) Specimen Collections

All specimens must be submitted to The Living Longer Institute in the provided containers, as they are ascertained to be trace element free.

Summary of testing / specimen requirements

Pre- Detoxamin CaNa_2 -EDTA Suppositories

- a) Hair Analysis
- b) Comprehensive stool analysis with fecal metals analysis.
- c) 24-hr urine collection for baseline urine elements
- d) Blood sample

Post- Detoxamin CaNa_2 -EDTA Suppositories

- a) Hair Analysis
- b) Comprehensive Stool Analysis with Fecal Metals analysis after the 3rd suppository, and one after the 90th suppository.

- c) 24-hr urine collection (toxic & essential) after the 3rd suppository, and one after the 90th suppository.
- d) Blood sample after the 90th suppository.

| Hematology (whole blood) | Chemistry (serum) | Pregnancy (urine) |
|---|---|------------------------------|
| Hemoglobin Hematocrit Total diff. and leukocyte count Red blood cell count Platelet count Platelet aggregation | 24 fasting sample Glucose Total protein Albumin Total bilirubin Alanine aminotransferase Aspartate aminotransferase Alkaline phosphatase LDH Calcium Sodium Potassium BUN Uric acid Creatinine VAP Cholesterol test* (see below) Serum Iron Serum Ferritin Transferrin Saturation Total Iron Binding Capacity PTT (activated partial thromboplastin time) Thrombin time Prothrombin time Hfe Genotype** | Females |

Schematic Outline of Study Procedures

| Assessment Description | Prescreening | Baseline | Follow-up Post- CaEDTA Provocation | Follow-up | Follow-up |
|--|----------------------------|------------------|---|-------------------|--------------------|
| Study timepoint Clinic visit # | Day -14 to -1 Telephone | Day 0 Visit 1 | Day 3 Visit 2 | Day 90 Visit 3 | Day 100 Visit 4 |
| Informed consent | VERBAL | WRITTEN | | | |
| Review of entry criteria | X | X | | | |
| Medical history | | X | | | |
| Dietary questionnaire | X | | | | |
| Vital signs and weight | | X | | X | |
| BMI | | X | | X | |
| Diagnostic exam | | X | | X | |
| Oral DMSA screen for study inclusion/exclusion | X | | | | |
| IV DMPS provocation w/12-hour urine | | X | | | X |
| CBC with differential | | X | | X | |
| Clinical chemistry panel | | X | | X | |
| Phlebotomy for retained specimens | | X | | X | |
| Urine pregnancy test | | X | | | X |
| Dispense study medication | | X | | | |
| Administration of study medication | | X | | X | |
| Drug accountability | | X | | X | |
| Subject global assessment of disease (VAS)-Diary card | X | | | | |
| Inv. global assessment of disease | | X | | | |
| Mini mental status examination | | X | | | |
| Hamilton Rating Scale for Depression | | X | | | |
| Adverse events | | X | | X | X |
| Concomitant medications | X | X | | X | X |

Corporate Sponsorship

This trial was conducted by Living Longer Health in collaboration with World Health Products, LLC, for-profit corporations operating in Cincinnati, OH and Irvine, CA respectively.

Results

Significant excretion was observed ($P < 0.05$) with cumulative CaNa_2EDTA therapy of Day 3 and Day 90 compared to Day 0 with lead, arsenic, cadmium (Figure 1) and nickel (Figure 2) in fecal samples. In addition, statistically significant cumulative excretions ($P < 0.05$) in arsenic, lead, mercury, cadmium and nickel in urine samples (Figure 3) were found. There were no differences in the safety profile comparisons within comprehensive metabolic chemistry panels between pre- and post-treatment blood values. Only minor transient complaints of loose stools and gas were reported in several subjects. Calcium disodium EDTA suppositories (Detoxamin) appear to be a safe and effective means to slowly and consistently remove a variety of toxic heavy metals, as evidenced by urine and fecal analysis.

Figure 1

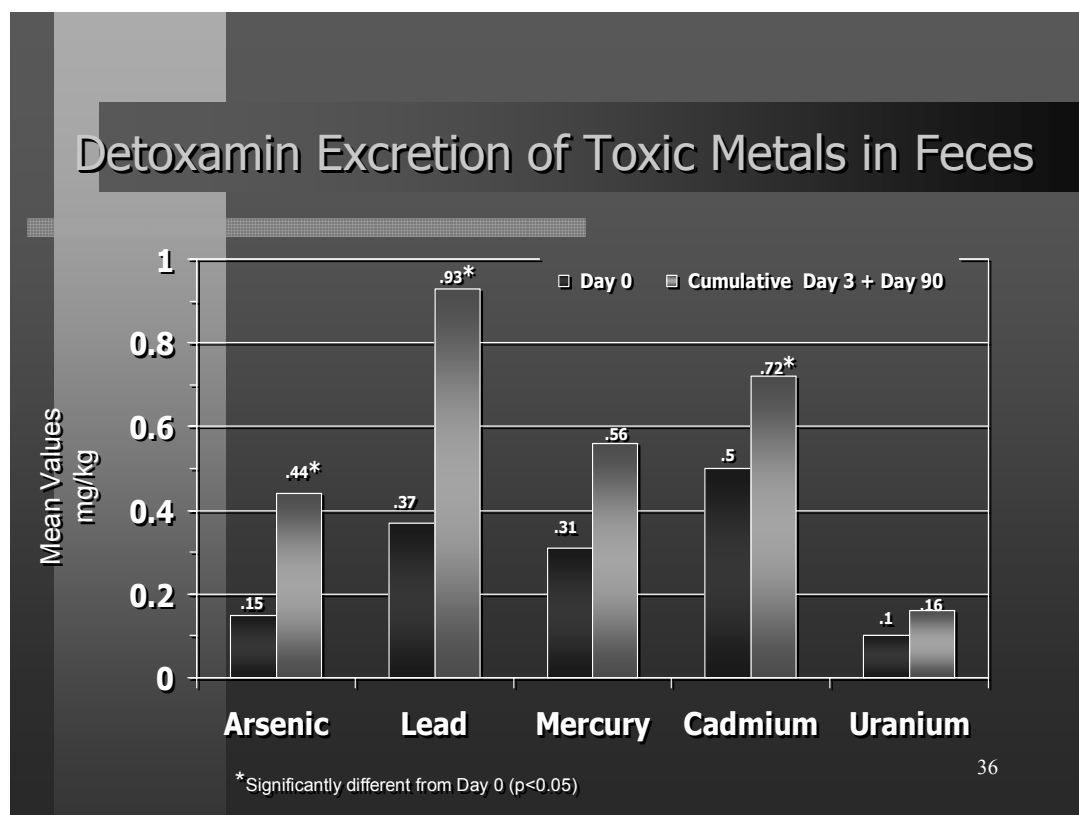


Figure 2

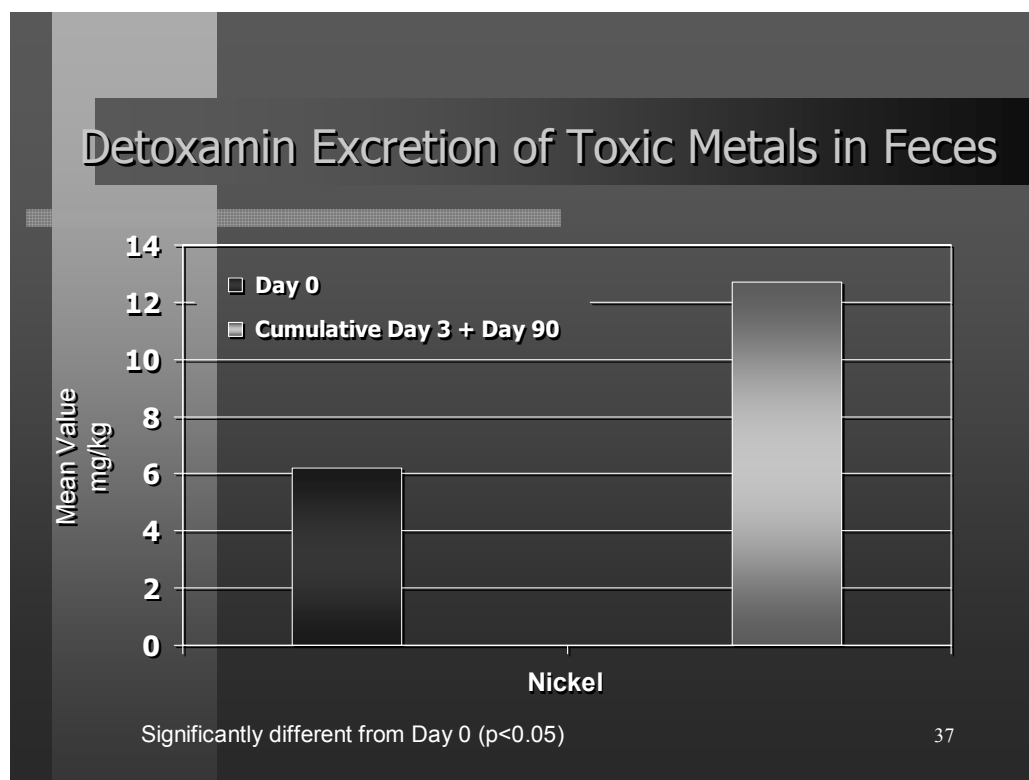
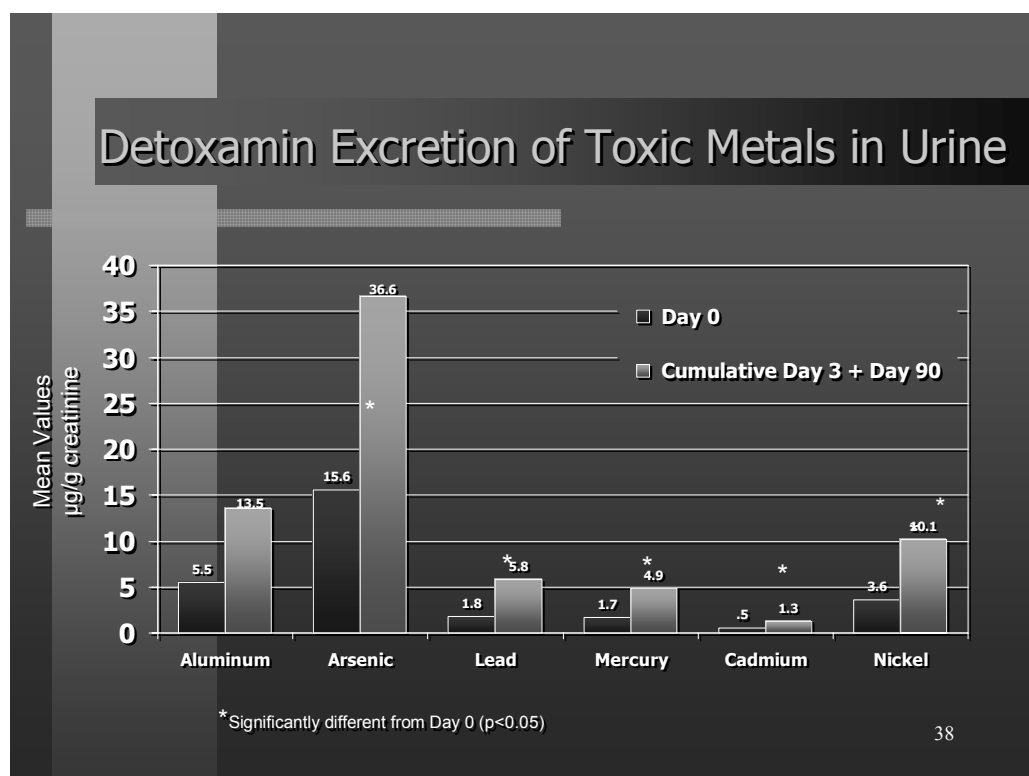


Figure 3



The Effects of Detoxamin CaNa₂ EDTA Suppositories on Elevated Blood Lead Levels in Children

IRB Approved Study in the Dominican Republic

A Clinical Report in Association with Columbia University and Fordham University

Ted Rozema, MD

Abstract: The effect of lead poisoning on high percentages of the pediatric population is cause for concern and it is one of the most common and preventable pediatric health problems today. Currently, the primary form of medical intervention consists of an expensive and painful CaNa₂ EDTA intramuscular injection. The availability of an easily administered effective medical treatment is an important component in controlling worldwide lead poisoning. This study utilizes a new form and route of administration of CaNa₂ EDTA, Detoxamin® suppositories. Rectal CaNa₂ EDTA suppositories (2000mg) were administered once per evening to 20 children for 10 consecutive days, no treatment after the following 10 consecutive days and re-administration of the initial suppository dose for another 10 consecutive days. Lead blood showed continual excretion from the pre-treatment concentration of 66.64µg/dL to 83.67µg/dL post treatment at the end of the 30 trial period. Lead urine levels rose from a baseline of 4.23µg/g creatinine to 325.55µg/g creatinine after only one suppository administration. The lead urine levels then dropped to 61.45µg/g creatinine within the first 10 days of treatment, and decreased to 9.94µg/g creatinine after the next 10 consecutive days of no treatment and rose to 22.71µg/g creatinine after the last 10 days of suppository administration. These data indicate significant and consistent excretion of lead in both blood and urine with the use of CaNa₂ EDTA suppositories in children exposed to high levels of lead. CaNa₂ EDTA suppositories offer a simple, convenient, non-invasive and cost- effective means of effective chelation for lead poisoning.

Introduction: Childhood lead poisoning is one of the most common pediatric health problems in the world today, and it is entirely preventable and reversible. Enough is now known about the sources and pathways of lead exposure, about ways of preventing this exposure, and about ways of reducing the lead content of the body to eradicate this disease permanently. The persistence of lead poisoning, in light of all that is known, presents a singular and direct challenge to public health authorities, clinicians, regulatory agencies, and society.

Lead is ubiquitous in the human environment as a result of industrialization. It has no known physiological value. Children are particularly susceptible to lead's toxic effects. Lead poisoning, for the most part, is silent: most poisoned children have no symptoms. The vast majority of cases, therefore, go undiagnosed and untreated. Lead poisoning is widespread. It is not solely a problem of inner city or minority children. No socioeconomic group, geographic area, or racial or ethnic population is spared.

Previous lead statements issued by the Centers for Disease Control (CDC) have acknowledged the adverse effects of lead at lower and lower levels. In the most recent previous CDC lead statement, published in 1985, the threshold for action was set at a blood lead level of 25 µg/dL, although it was acknowledged that adverse effects occur below that level. In the past several years, however, the scientific evidence showing that some adverse effects occur below levels at least as low as 10 µg/dL in children has become so overwhelming and compelling that it must be a major force in determining how we approach childhood lead exposure.

It is not possible to select a single number to define lead poisoning. Epidemiological studies have identified harmful effects of lead in children at blood lead levels at least as low as 10 µg/dL. Some studies have suggested harmful effects at even lower levels, but the body of information accumulated so far is not adequate for effects below about 10 µg/dL to be evaluated definitively. As yet, no threshold has been identified for the harmful effects of lead.

Because 10 µg/dL is the lower level of range at which effects are now identified, primary prevention activities are typically directed at reducing children's blood lead levels below 10 µg/dL or 14 µg/dL. While the overall goal should be to reduce children's blood lead levels below 10 µg/dL, there are entrenched reasons for not attempting to do interventions directed at individual children to lower blood lead levels of 10-14 µg/dL. First, practical medical interventions for children with blood lead levels in this range have previously

been unavailable. Second, the sheer numbers of children in this range would preclude effective case management in established intravenous therapy. Clearly, a simply and effective therapy such as suppository is needed.

The single, all-purpose definition of childhood lead poisoning has been replaced with a multi-tiered approach, described in the following table:

| CLASS | Blood lead concentration (µg/dL) | COMMENT |
|-------|----------------------------------|---|
| I | <10 | A child in Class I is not considered to be lead poisoned. |
| IIA | 10-14 | Many children (or a large proportion of children) with blood lead levels in this range should trigger community-wide childhood lead poisoning prevention activities. Children in this range may need to be rescreened more frequently. A decrease in blood lead levels would be beneficial. |
| IIB | 15-19 | Child should receive nutritional and educational interventions and more frequent screening. If the blood lead level persists in this range, environmental investigation and intervention should be done. Non-invasive medical intervention should be done. |
| III | 20-44 | Environmental evaluation, remediation and a medical examination should take place. Such a child needs pharmacological treatment of lead poisoning. |
| IV | 45-69 | A child in Class IV will need both medical and environmental interventions, including even I.M. chelation therapy. |
| V | 69> | A child with Class V lead poisoning is a medical emergency. Medical and environmental management must begin immediately. |

Lead is a poison that affects virtually every system in the body. The risks of lead exposure are not based on theoretical calculations. They are well known from studies of children themselves and are not extrapolated from data on laboratory animals or high-dose occupational exposure. Since 1970, our understanding of childhood lead poisoning has changed substantially. As investigators have used more sensitive measures and better study designs, the generally recognized level for lead toxicity has progressively shifted downward. Before the mid-1960s, a level above 60 µg/dL was considered toxic (Chisholm and Harrison, 1956). By 1978, the defined level of toxicity had declined 50% to 30 µg/dL.

Lower blood lead levels cause adverse effects on the central nervous system, kidney and hematopoietic system. Blood lead levels as low as 10 µg/dL, which do not cause distinctive symptoms, are associated with decreased intelligence and impaired neurobehavioral development (Davis and Svendsgaard, 1987; Mushak et al, 1989).

The concern about adverse effects on central nervous system functioning at blood lead levels as low as 10 µg/dL is based on a large number of rigorous epidemiological and experimental studies. Several well-designed and carefully conducted cross-sectional and retrospective cohort studies in many different countries have been conducted (Lansdown et al., 1986; Fulton et al., 1987; Fergusson et al., 1988; Silva et al., 1988; Bergomi et al., 1989; Hansen et al., 1989; Hatzakis et al., 1989; Winneke et al., 1990; Lyngbye et al., 1990; Needleman et al., 1990; Yule et al., 1981; Hawk et al., 1986; Schroeder et al., 1985). Some inconsistencies can be found in the results of these studies, but the weight of the evidence clearly supports the hypothesis that decrements in children's cognition are evident at blood lead levels well below 25 µg/dL. No threshold for the lead-IQ relationship is discernable from these data. Recent evaluation of 24 major cross-sectional studies provides strong support for the hypothesis that children's IQ scores are inversely related to lead burden (Needleman and Gatsonis, 1990).

According to the Natural Resources Defense Council, blood lead levels as low as 10 µg/dL, which do not cause distinctive symptoms, are associated with reading and learning disabilities, reduced attention span and behavioral problems. The ramifications of the proliferation of lead pollution from industrialization, combined with the devastating effects of health, are sobering. A simple and effective therapy, such as EDTA chelation via suppository, is urgently needed.

Materials and Methods: A cluster of previously untreated children with high blood lead levels was desired for the purpose of testing the efficacy of Calcium disodium EDTA rectal suppositories to remove toxic metals from the human body.

1.) Determinization of study area: Friends of Lead Free Children, a non-profit organization connected to Columbia University and Fordham University, assisted in the search. A residential neighborhood in Haina, Dominican Republic was selected. The residential neighborhood was located adjacent to a battery recycling plant. All preliminary testing indicated 100% of residents as markedly toxic with lead.

2.) The selection of subjects into the study: Children who had been identified with blood lead levels over 10 mcg/dL were determined in a twenty-four hour urine collection by Ion Coupled Plasma Emission Spectroscopy. Hg. Analysis was determined by cold vapor mercury analysis.

3.) Individual treatment of lead overload: Cautious removal of lead from body depots was achieved through the use of Calcium disodium EDTA rectal suppositories. The use of suppositories provided for the prevention of local corrosive action of toxic metals on mucous membranes.

4.) Compensation: Compensation was not paid to subjects; however, no charges were incurred by participants for the drug and laboratory testing.

5.) Safety: By determining the concentration of heavy metals in the urine following provocative stimulation, the therapy with EDTA was scientifically determined, providing a safe treatment program. The study simultaneously provided diagnostic information regarding heavy metal burden as well as a defined treatment protocol for lead toxicity in a pediatric population. EDTA is a substance with low systemic and local toxicity and is generally well tolerated. The drug, per se, has been classified GRAS by the FDA, no cases of anaphylaxis have been reported through the oral administration of EDTA or through its use as a food additive.

6.) Alternative therapies: Alternative therapies were available for the treatment of metal intoxications, including (R,S)-2,3-dimercapto propane-1-sulfonic acid (DMPS,) as well as its close standing analog, DMSA. A significant advantage of using EDTA suppositories in a pediatric population include:

—a) Cooperative binding constant for lead.

—b) The suppository route of administration at bedtime was (is) an easy and acceptable delivery system.

—c) The antioxidant/free radical quenching role of EDTA made it superior over the other agents available due to the fact that neurological dysfunction was (is) recognized as a result of free radical mediated damage.

—d) EDTA was already approved for oral administration by the FDA and is on the GRAS list.

—e) EDTA is an ANTIDOTE to counteract the TOXIC action of lead from the environment.

7.) Medical care: Medical care was provided by Universidad de Autonomia de Santa Domingo. In the event of a medical emergency connected with the study, subjects were to contact the appropriate center, but this was never necessary. In addition, all participants could receive product and clinical information by calling: Ted Rozema, M.D., the principal investigator.

8.) Data coordination: Data was coordinated and maintained by the principal investigator, all data was statistically analyzed. Information was made available to all appropriate authorities, including IRB of the GLCCM and the FDA.

9.) Clinical laboratory: Clinical laboratory facilities and medical support was provided by AmScot Medical Laboratories, Inc. To ensure the safety and integrity of the study, the following analyses were assessed:

a) Baseline:

— Smac 18 with CBC - manual differential Blood lead determination

— Urine (24-hour collection) —heavy metals to include: Pb, Cd, Hg, As, Ni, Al

— B2 - micro globulin (serum) Anti - TPO Total Ca/Ca²⁺ Mg/Mg²⁺ Pt/APTT PTH

b) Provocative EDTA challenge:

— Blood lead determination

— Urine (9-hour) - heavy metals - Pb, Cd, Hg, As, Ni, Al

— B2 - micro globulin (serum) Total Ca/Ca²⁺ Mg/Mg²⁺ Pt/APTT PTH

c) Mid-study laboratory evaluation:

- Blood lead determination
- CBC - manual differential Urine (9-hour) - heavy metals - Pb, Cd, Hg, As, Ni, Al
- B2 - micro globulin (serum)
- Total Ca/Ca²⁺ Mg/Mg²⁺
- Pt/APTT PTH

d) Post study (6 weeks):

- Blood lead determination
- SMAC 18 with CBC - manual differential
- Urine (9-hour) - heavy metals - Pb, Cd, Hg, As, Ni, Al
- B2 - micro globulin (serum)
- Total Ca/Ca²⁺ Mg/Mg²⁺
- Anti-TPO
- PTH

Research Protocol: A study to determine the efficacy of Calcium disodium EDTA (Detoxamin, supplied from World Health Products, Draper, Utah) used as a rectal suppository in removing toxic metals from the human body. Subjects: children with proven lead toxicity (blood lead levels $>10\mu\text{g/dL}$). Study design:

1.) Enrollment.

2.) Blood lead levels drawn to enter into study with simultaneous determination of urine lead excretion (total urine minerals - if possible with parental assistance).

3.) Treatment phase.

| | PRE | AFTER 10 DAYS | AFTER 10 DAYS | AFTER 10 DAYS |
|-------|---------------------|---------------------|---------------------|---------------------|
| BLOOD | XX — Sup p | XX — No supp | XX — Supp | XX |
| URINE | XX — Sup p | XX — No supp | XX — Supp | XX |

4.) Placement of a rectal suppository containing 2 grams of Calcium Disodium EDTA nightly for 10 days, then 10 days without EDTA, then placement of the EDTA suppository for 10 days, continue this program for two courses of treatment.

5.) Laboratory determinations:

Purpose is to demonstrate gradual reduction of both blood and urine lead levels over time with a simple and cost-effective method. It was anticipated that

methods to reduce lead intake would be in place during and after this study. Unfortunately, no environmental mitigation was ever enacted.

Specimen Collection Regimen: Pre-study:

- 1.) Collection of 3 to 5 ml of whole blood in heparinized, lead-free cures.
- 2.) Collection of 9 hours of urine. This was measured from the time the child went to bed until 9 hours later. It was anticipated that the children were not getting up at night to urinate and mother would need to watch to catch the first morning specimen, then determine the 9-hour point and collect the additional urine to make the complete collection. This provided a base line for both blood levels and excretion on a daily basis.

Just before the first suppository:

- 1.) Collection of 1 to 5 ml of whole blood in heparinized, lead-free cures.
- 2.) Insertion of the first suppository in the child's rectum, high as possible, just before the child goes to sleep, preferable with the child already in the bed.

The morning after the first suppository:

- 1.) Collection of 9 hours of urine. This was measured from the time the child went to bed until 9 hours later. It was anticipated that the children were not getting up at night to urinate and mother would need to watch to catch the first morning specimen, then determine the 9 hour point and collect the additional urine to make the complete collection.

The morning before the 10th suppository:

- 1.) Collection of 3 to 5 ml of whole blood in heparinized, lead-free cures.

The morning after the 10th suppository:

- 1.) Collection of 9 hours of urine. This was measured from the time the child went to bed until 9 hours later. It was anticipated that the children were not getting up at night to urinate and mother would need to watch to catch the first morning specimen then determine when is the 9 hour point and collect the additional urine to make the complete collection.

The morning of the 19th day: This is the last day without a suppository before the next ten days of suppository administration.

- 1.) Collection of 3 to 5 ml of whole blood in heparinized, lead-free cures.

2.) Collection of 9 hours of urine. This was measured from the time the child went to bed until 9 hours later. It was anticipated that the children were not getting up at night to urinate and mother would need to watch to catch the first morning specimen, then determine the 9 hour point and collect the additional urine to make the complete collection. This gave us a determination of equilibration after no treatment for 10 days.

The morning of the 30th day:

1.) Collection of 3 to 5 ml of whole blood in heparinized, lead-free cures.

The morning after the 30th suppository:

1.) Collection of 9 hours of urine. This was measured from the time the child went to bed until 9 hours later. It was anticipated that the children were not getting up at night to urinate and mother would need to watch to catch the first morning specimen, then determine the 9 hour point and collect the additional urine to make the complete collection. All specimens were taken to the laboratory of Dr. Conrado Depratt at the Instituto De Quimica of the Universidad Autonoma de Santo Domingo.

Results: Average 20 children test data:

BLOOD LEAD
LEVELS

| | | |
|--|-------|------------------|
| Pre-study | 66.64 | $\mu\text{d/gL}$ |
| After 10 days of suppositories | 39.09 | $\mu\text{d/gL}$ |
| After 10 days without suppositories | 61.45 | $\mu\text{d/gL}$ |
| After 10 more days on suppositories | 83.67 | $\mu\text{d/gL}$ |

URINE LEAD
EXCRETION LEVELS

| | | |
|--|---------|------------|
| Pre-study | 004.23 | μd/g creat |
| After 1st suppository | 325.55 | μd/g creat |
| After 10 days of suppositories | 061.445 | μd/g creat |
| After 10 days without suppositories | 009.04 | μd/g creat |
| After 10 more days on suppositories | 022.71 | μd/g creat |

The data clearly demonstrates that Detoxamin, (EDTA delivered in rectal suppository form), effectively removes lead from children with lead poisoning. The continued high excretion level, after 10 days without Detoxamin is of special interest. Also of special interest is the rebound effect in the blood lead levels. Its degree reflects the high amount of stored lead in the tissue and bones and the attendant mobilization effect. Each time the blood lead level was diminished, additional lead was mobilized from the tissues and bones.

It was anticipated that methods to reduce lead intake would be in place during and after this study. Unfortunately, no environmental mitigation was ever enacted. Ideally, environmental intervention would have been enforced and the Detoxamin Calcium disodium EDTA rectal suppository therapy would have continued for a 6-month duration. This circumstance was not possible.

Clinical Case Studies of Patients on CaNa_2EDTA (Detoxamin[©])

Rita Ellithorpe, MD

Tustin Longevity Center, Tustin California

Summary

Dr. Rita Ellithorpe performed random packed red blood toxic heavy metal analysis to approximately 279 patients. She found a 98% presence of multiple toxic metals in the test sampling. Dr. Ellithorpe has over seven years experience with practical clinical use of Detoxamin and has administered the supplement to over 1800 patients totaling over 100,000 doses. The overall clinical outcomes, in general, with patients on Detoxamin are improved mental clarity, increased energy, increased endurance with physical activity, reduced blood pressure, improved lipid profiles, enhanced cardiovascular performance, improved libido, improvement in symptoms of prostatitis and prostate conditions and an overall improvement in quality of life.

Figures 1, 2 and 3 are summaries three clinical case studies of patients on Detoxamin.

Figure 1

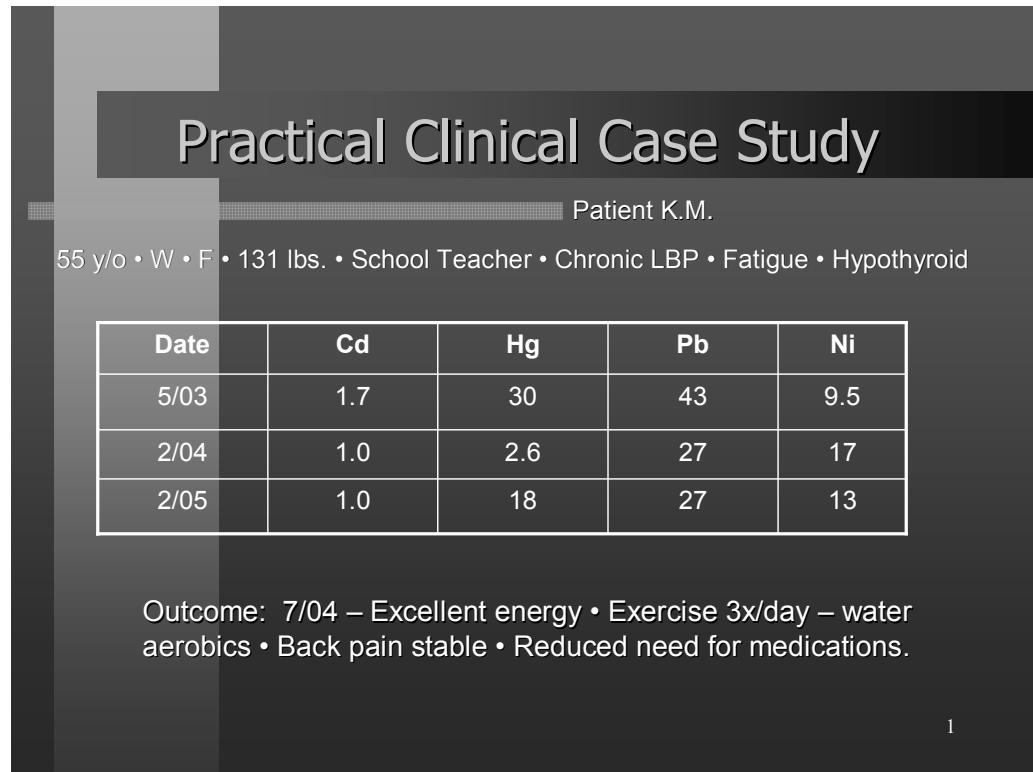


Figure 2

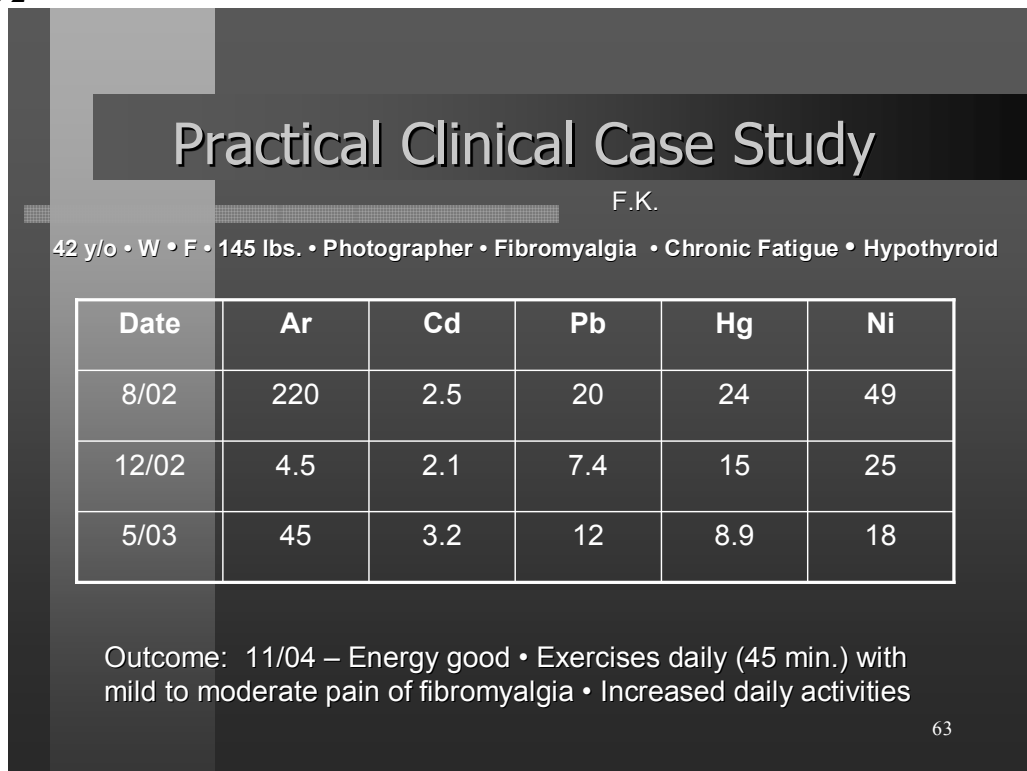
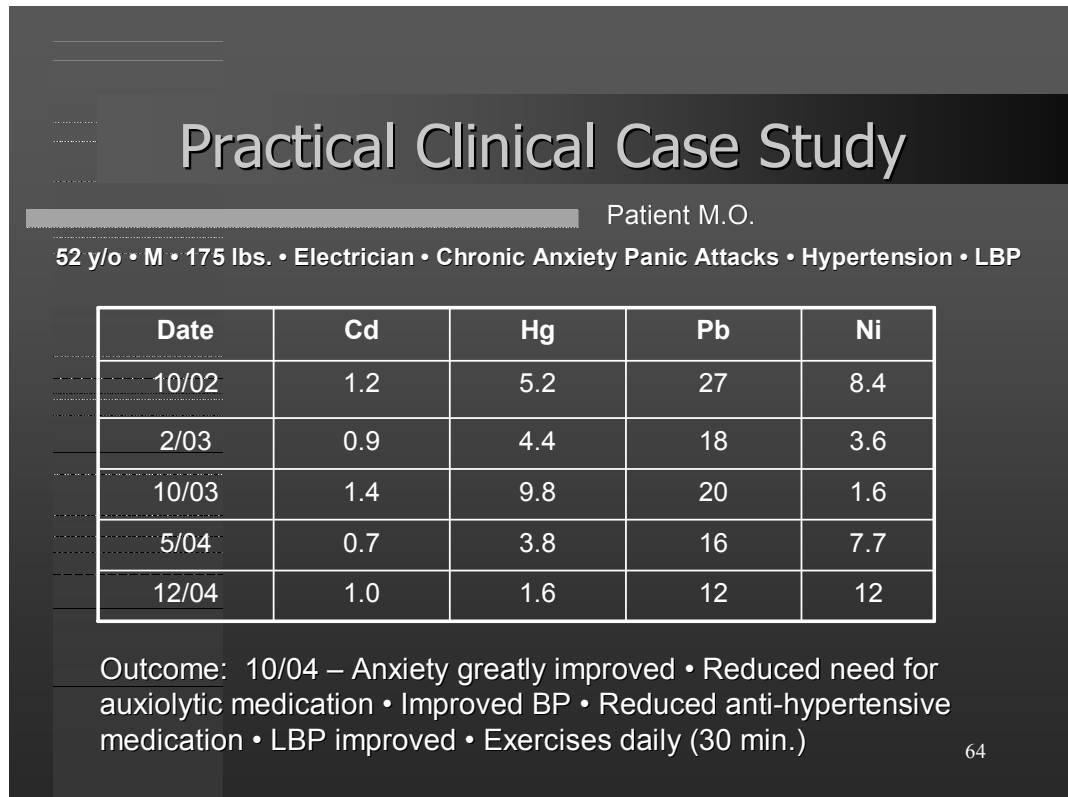
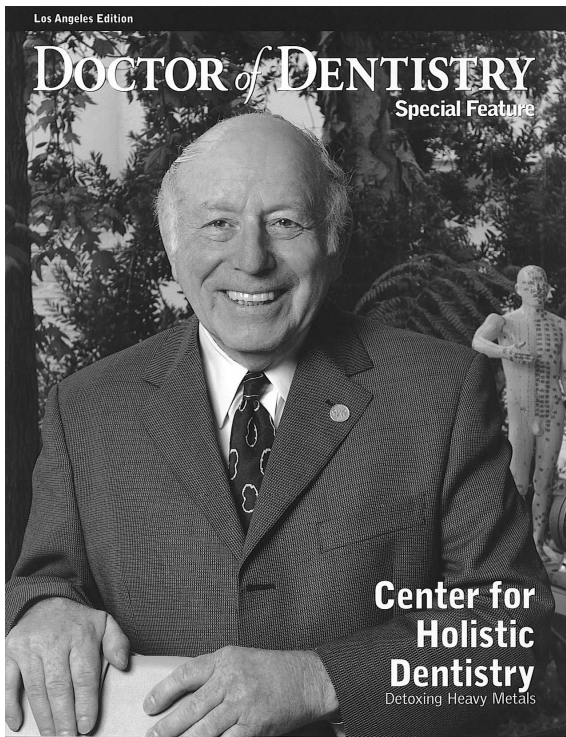


Figure 3





Dr. Harold Ravins

Detoxing Heavy Metals

By Debbie L. Sklar

There are a lot of exciting new discoveries in dental medicine to provide better outcomes for patients. One of the emerging fields is what has been called complementary, integrative or holistic medicine, combining traditional folk remedies from cultures worldwide with the most cutting-edge technologies and techniques of the mainstream.

Harold Ravins, D.D.S., a general dentist and the founder of Raveco Holistic Dental Center in Beverly Hills, has been in private practice since 1963. "I've always thought that the mouth is a mirror into the overall body," said Dr. Ravins. He said he hopes that in the near future holistic dentistry will become the norm.

"As heavy metal toxicity and acupuncture were eventually accepted by mainstream medicine, dentists should keep an open mind about other elements of alternative medicine," he commented.

With his drive and determination, Dr. Ravins continues to break new ground in the area of holistic dentistry by offering patients what he believes to be a healthier way to take care of their teeth - as part of their whole body.

THE PATH TO INTEGRATIVE DENTISTRY

Born and raised in Buffalo, NY, Dr. Ravins is the son of third-generation kosher meat packers. He said he knew early on that he wanted to become a dentist and was always intrigued by biology.

"I learned how to stitch the chickens as a kid, and this helped me become an excellent surgeon," he laughed, "I was interested in dentistry at the age of 9 or 10, as well as nature because my own dentist took a lot of time explaining dentistry to me."

After graduating from Fairleigh Dickenson School of Dentistry in New Jersey in 1960, he attended a graduate program in oral bacteriology at Ohio State University. He went back to his native Buffalo for a year to work with his mentor, but quickly tired of shoveling snow. Instead, he longed for the sunny weather of California and found himself on the road headed west.

"I always had the aspiration of becoming a successful dentist in California," he recalled. "Dentistry there, as now, was the gold standard for the country and I was very into cosmetic work that was being done in Beverly Hills. I came, got a job here, but always wanted to have my own practice."



Harold Ravins, D.D.S., practices a high level of conventional dentistry within the context of a holistic approach to health.

However, the formative years were not always smooth sailing for Dr. Ravins.

"It was about 1968 when I was diagnosed with incurable sciatica by my orthopedist," he said. "It was a long weekend in Vegas and after walking the strip with my cane and wife, a taxi cab pulled up to us and it turned out to be a classmate of my wife's who was going for his chiropractic boards."

The classmate invited Dr. Ravins and his wife up to his apartment where he treated the dentist, who suddenly felt much better.

"On Monday, I went back to the tennis courts, and when my orthopedist, also a tennis player, heard my story he said a few unpublishable words and I realized most other doctors would refuse to give evidence of the efficacy of alternative modalities a fair hearing," he said. "I later studied with chiropractors, and introduced cranial therapy to several of my study groups at that time and I published an article called 'Car Crash, Jaw Lash.'"

Dr. Ravins, who still attends many seminars and conferences to keep abreast of the fields, always seems to be in the right place at the right time. He has been fortunate to study with some of his most prominent dental colleagues, among them Dr. Harry Tepper, an orthodontist and innovator in his own right. He taught Dr. Ravins to use removable braces to prevent and correct crooked jaws and teeth for children and adults - a revolutionary idea then. Dr. Ravins did orthodontics for the first 25 years of his career.

"I was learning about more treatment procedures that most dentists did not accept," he said. "But there was always a minority of us who were more open-minded. I was always interested in nutrition and, amazingly, our dental school in New Jersey had the first organic cafeteria in the country in 1956. I joined the International Academy of Applied Nutrition in the 1960s."

Taking a leap of faith, Dr. Ravins decided to create the Raveco Holistic Dental Center after three years of building up his general practice in Beverly Hills.

"I became concerned about the harmful effects of the mercury in amalgam fillings, which are 50% mercury, around this time," he explained. "I haven't done an amalgam filling in 20 years, and I use porcelain or white composites instead. I used to use gold, but not anymore because of the new CAD-CAM system and I have been following the European dentists by staying metal free. I had my own amalgams removed 38 years ago."

He was intrigued by the broader field of holistic dentistry, he said. Holistic, Dr. Ravins explains, indicates that what is going on in the mouth affects the entire body.

"Back in the early 1900s, a very famous Canadian physician, Sir William Osler, wrote that the mouth is a mirror of the body. He was celebrated as the 'Father' of modern clinical medical practice, I, too, take the position that whatever dentistry is performed in the mouth affects the general health of the patient. It's no surprise that holistic dentistry is becoming more popular in the public's eye and it is going to become a much bigger part of general dentistry in the coming decade."



Dr. Ravins tests his patients' cellular health using a device NASA uses to determine the health of astronauts.

A FIRST VISIT

When a patient first visits Dr. Ravin's office, they receive a standard dental examination, including checking for periodontal infection.

But the holistic objective is to also find hidden infections in the mouth that are not considered the usual dental problems. In many situations, pain is not the primary indicator of a health problem. Detection is accomplished with good quality X-rays and a comprehensive clinical examination, which includes various levels of dental acupuncture energistic testing (James Oschman, Ph.D., correlates traditional Asian medicine's concept of the body's energy with Western science in Energy medicine: The Scientific Basis).

Another part of this first visit includes a bite analysis involving the T-scan test. The patient bites down on paper with a computer chip that transmits a picture of how the teeth contact each other and how well the jaw is functioning.

Included in this comprehensive examination, according to the principles of whole body dentistry, is the BIA, or bioelectrical impedance analysis, which was developed for NASA in the 1970s to measure body mass and detects cellular health in astronauts. It is a painless way to measure the electrical activity of the cell membranes on the foot to determine whether cells are absorbing nutrients and resisting toxins. It is a good indicator as to whether the immune system is functioning normally and provides an excellent marker for overall wellness.

The safest and easiest way to diagnose the presence of heavy metals in the body is a fecal test, which Dr. Ravins stated provides incontrovertible scientific evidence that many patients suffer from poisoning. The metals measured are mercury (with amalgams and without), lead, nickel, uranium, antimony, tungsten, thallium, platinum, beryllium, arsenic, bismuth, cadmium and copper. The fee is \$185; the total evaluation including a written report is under \$1,000.

"The center has been looking for years for an accurate, safe way to determine mercury toxicity," he said. "We discovered the heavy metal fecal test that was used in pediatric medicine for years, but that was not very popular with the U.S. medical community." The center also found the best lab in the U.S. for processing the test accurately.

At present, Dr. Ravins has more than 200 patients who have been tested for heavy metals in their system before they had amalgam removal. (The center also found a lab that met its standards.) The result was that 99% showed toxic levels of mercury. He invites other dentists to take the test for themselves and to let him know if they might have an interest in attending a workshop about detoxification he plans to conduct.

"I don't provoke or tell my patients that they should have their amalgams taken out because of the mercury," he said. "I give them literature that talks about the harmful effects of mercury. They are asking me to remove them, and I, in turn, ask them to take the fecal test; most come back with high levels. Yet, according to the American Dental Association, there is no scientific proof that the mercury in amalgam is leaking into the body."

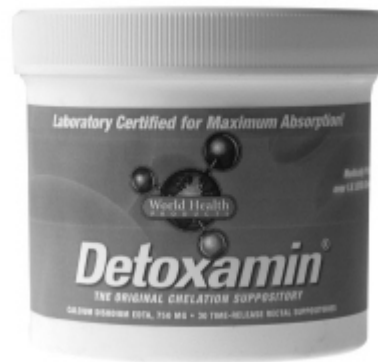
It's an illogical position, argues Dr. Ravins. "The government's own scientists agree that mercury is very toxic, we now have tests that show a large number of people have high levels of mercury, so are we going to say that putting mercury in the mouth for a lifetime is not related?" In fact, although amalgam fillings saved millions of teeth in the past, they have been banned in some European countries for many years and we now have healthier materials to use instead of amalgam, he points out.

The ADA does say it is okay to remove amalgams at the patient's request and most dentists will do so cynically, Dr. Ravins commented. But his colleagues are starting to come around. The State Dental Licensing Board no longer requires using mercury amalgams to practice and is the first dental board in the U.S. to take that pioneering position. "Ten years ago, I would have been considered nuts with all of this, but now the evidence is hard to deny."

DETOXIFYING MERCURY

Once the levels of mercury have been uncovered, it is not as easy as just going in and chiseling out the old amalgams.

"It requires a great deal of experience and understanding, because each patient is so different," he said. "The periodontal evaluation is extremely important because if you have an infection in the mouth, it can contribute to other infections in the body. But if we do a BIA and the cells aren't working right, it means that something is wrong with the overall cellular health."



Dr. Ravins is so adamant about the dangers of mercury that he even has negative ion machines in his operatories to take out mercury vapors in his office.

"The mercury vapors are most horrendous to our health. Since the vapors are positive(ly) charged, they attach to the negative ions generated in the room and are collected on a positive cathode reservoir on the wall," he said. "I have to clean the reservoirs every three weeks. Most other toxic gases are also positively electrically charged."

So, why not just simply open a window? Because he would be fined \$2,500, since mercury is recognized as so poisonous by the Environmental Protection Agency.

Dr. Ravins said that recently, a young woman contemplating pregnancy came in for a dental examination regarding the mercury issue.

"She said she wanted to be toxic-free on conception and then referred her husband to me for the same reason," he recalled.

One of the ways of ridding the body of mercury toxins is by using a new detoxification product called Detoxamin, a CaN2 or calcium disodium EDTA time-released suppository, manufactured by World Health Products in Irvine, CA.

(EDTA is certified by the FDA for doctors to remove excess lead from the body and the FDA recently approved research of Can2 EDTA indicating this was effective for removing mercury, as well, and a safer chemical to use.)

"Through all our studies, we've discovered that it could take months, or even years, to rid the body of such toxins. This suppository is a convenient way to do it and I'm so impressed by this product that I decided to start using the product on myself," Dr. Ravins said.


World Health Products is the originator and developer of the patented formula and "Detoxamin is the only product of its kind that actually has the proof to back up its claims," Dr. Ravins noted, "Many companies promote better price and make product claims that have absolutely no scientific or clinical reference of any kinds."

"Scientific evidence is important because all EDTA suppositories are not alike. Formulas, ingredients, consistency, time-release and other important aspects that create high absorption of EDTA are all different. If you have used IV EDTA chelation before, you know absorption and bioavailability are the keys to performance. Detoxamin is the only viable alternative to IV EDTA chelation in the world." according to CEO Edward Salmon.

IV EDTA chelation although very efficacious, is very expensive, invasive, time-consuming and inconvenient, he points out, putting this needed therapy out of reach for most people. Oral EDTA chelation products are poorly absorbed and Detoxamin fits today's lifestyle with its simple, convenient, gently and efficient toxin removal with proven pharmacokinetic absorption and clinical efficiency, he added.

"Our patented chelation is becoming the preferred choice due to the obvious convenience and affordability factor, but more importantly, to the extraordinary efficacy, safety and scientific validation through third party (research) "Salmon pointed out. "Our current research is focused on the diseases of the prostate, with significant success assessed by the University of California at Irvine biostatisticians."

A typical fecal test shows a patient's heavy metal toxicity.



LAB#: P051205-0423-1

PATIENT: John Doe

SEX: Male

AGE: 38

CLIENT#: 1000

DOCTOR: Harold E. Ravins, DDS

10001 Wilshire Blvd.

Los Angeles, CA 90024

POTENTIALLY TOXIC METALS

| METALS | RESULT mg/kg | REFERENCE RANGE | PERCENTILE | |
|-----------|-----------------|-----------------------|------------------|------------------|
| | | | 99 th | 50 th |
| Mercury | 0.310 | <0.05 (no amalgams)* | | |
| Mercury | 0.310 | <0.5 (with amalgams)* | | |
| Antimony | 0.075 | < 0.100 | | |
| Arsenic | 0.10 | < 0.30 | | |
| Beryllium | 0.004 | < 0.003 | | |
| Bismuth | 0.050 | < 0.100 | | |
| Cadmium | 0.04 | < 0.10 | | |
| Chromium | 0.0 | < 0.1 | | |
| Copper | 0.05 | < 0.50 | | |
| Lead | 0.05 | < 0.50 | | |
| Nickel | 0.1 | < 0.1 | | |
| Platinum | <0.01 | < 0.003 | | |
| Thallium | 0.010 | < 0.020 | | |
| Vanadium | 0.007 | < 0.003 | | |
| Uranium | 0.101 | < 0.100 | | |

% WATER CONTENT

| | RESULT %H ₂ O | EXPECTED RANGE | 2SD LOW | 2SD HIGH |
|-----------------|-----------------------------|-------------------|---------|----------|
| % WATER CONTENT | 74 | 60-85% | 72.5% | 72.5% |

THE EVIDENCE ON MERCURY

As for mercury, World Health Products said it is one of the most toxic elements on the planet, probably second only to plutonium, yet people worldwide have it in all tissues of their bodies. It continues to be dumped into our waterways and soil, placed into our teeth and injected into our bodies through vaccinations.

Toxicity cause by excessive mercury exposure is now becoming recognized as a widespread environmental problem and is continuing to attract a great deal of public attention. A Nationals Academy of Science

study published in July 2001 estimates that up to 60,000 children born in the U.S. each year may be affected by mercury toxicity. In March of 2002, an environmental group had charged the FDA of failing to warn the public of the dangers of mercury contamination from eating tuna, which contains high levels of mercury. Texas researchers have found a possible link between autism and mercury in the air and water. In fact, the incidence of autism has grown in the past 20 years, from one in every 2,000 children to as high as one in every 166. Researchers have been hard pressed to explain the increase, but many believe mercury to be the culprit, World Health Products points out.

The United Nations' World Health Organization also reports that the amount of mercury absorbed daily by the average human body is 0.3 micrograms (mcg) from water and air, 2.61 mcg from fish and 17 mcg from dental amalgams. Research points out that 80% of mercury vapor is absorbed into the blood, going directly from the nose to the brain, following nasal nerve pathways. Dentists have four times as much of a body burden of mercury than

average non-dentist people. Dental workers show 50 to 300% more mercury in hair and fingernails than the average population. Before public awareness campaigns started, it is a notable fact that the preservative thimerosal (usually added to vaccines) contained mercury. In 1999, the Centers for Disease Control called for the removal of mercury from vaccines. Paradoxically, the CDC still continues to recommend the measles, mumps and rubella vaccine, World Health said.

According to World Health's website: "If you are one of the millions of Americans who has received silver dental fillings, take notice. Mercury makes up about 50% of every amalgam dental filling, also known as 'silver' fillings, Amalgam fillings can release mercury for up to 70 years. Someone with eight amalgams, for example, could have 120 mcg released into the saliva per day. The maximum allowable by the EPA is less than 0.1 mcg per kilogram of body weight per day to be absorbed into the human body. We now know that the dental mercury/silver amalgam filling is 'chemically and electrically active.' Science has proven that every time we eat, drink or breathe, we may be absorbing disturbing releases of decomposing, toxic particles - mercury included. This chronic, toxic accumulation is being shown to have serious, long-term consequences on our immune system, resulting in a variety of diseases and conditions. Consider that while 78% of Americans have dental fillings, 95% of people with disorders of the central nervous system such as MS, epilepsy, paralysis and migraines also have silver dental fillings. This begs the question, 'Would you want mercury - one of the most powerful neurotoxins on the planet - embedded in your mouth, only inches from your brain?' The answer is obvious. This is the same reason why you can no longer buy an oral or rectal mercury thermometer.

Salmon challenges dentists to take the heavy metal toxicity fecal test, which World Health can also supply, as a starting point. It is simple, using a prepaid package, with results in a week.

ACUPUNCTURE AND INTERCONNECTION

"Every medical problem is interconnected throughout the body, which is why, in acupuncture, teeth are tied to different organs," Dr. Ravins said. "Since the science is there regarding acupuncture, the insurance industry has recognized this several years ago. As a certified dental acupuncturist, I use it on almost every patient. This is part of the new dentistry involving quantum mechanics and energy, which (Raveco Holistic Dental Center) has documented over the years. By crossing the midline of the mouth - the conception meridian - with a metal wire or bridge, this blocks the energy flow. The symptoms that are cleared include headaches and neck aches' advantages then include improved vision, equilibrium restored, and behavior modifications in children with orthodontic wires. Many prostate patients have been able to reverse their prostate disease diagnosis by addressing the mouth as part of the overall cause of the problem." Heavy metals have been shown to inflame the prostate, too.

But everything Dr. Ravins does is not "alternative." He tries to stay abreast of the latest dental technology and is an avid user of the CEREC system for making dental crowns. He is fully equipped with digital X-rays, which reduce radiation by 90% compared to film. He also used Diagnodent, a laser instrument which measure decay where the X-ray cannot.

"The center as also been using laser dentistry for more than 10 years in the treating of gum disease, and more recently, we are able to treat teeth for decay, nerve infection and tooth sensitivity without using injections, as well as for bleaching," he said. "It's excellent for children - no more needles for drilling."



Raveco Holistic Dental Center staff (left to right): Elisa Goller (front office), Sandy Urbino-Mallette (office manager), Nicole Mirilla (front office), Jolia Navarro (dental assistant), Yesenia Ramirez (dental assistant), Leanh Ly, RDH, Martha Moreno (dental assistant), Harold Ravins, D.D.S., CDA, Giri Palani, D.D.S.

Dr. Ravins is hoping to add an associate to the Raveco Holistic Dental Center to help him expand the practice to meet demand as the public becomes more interested in these things.

"I plan to take my 45 years of study and teaching of holistic dentistry to the forefront of the dental profession," he said. "The plan is to share all of this with a new partner who can continue the importance of this work and possibly buy the practice at some point. In five years, I see myself doing clinical research to show how the mouth can help keep a person healthier and prevent disease."

At 73, Dr. Rains is just getting started with what may be the most ambitious project in the history of dentistry.

Harold Ravins, D.D.S.
12381 Wilshire Blvd. #103
Los Angeles, CA 90025
(310)207-4617



hravins@earthlink.net
www.ravecoholistic.com

Edward Salmon, CEO
World Health Products
1582 Deere Ave #B
Irvine, CA 92606
(949) 474-0309
jsalmon@detoxamin.com
www.detoxamin.com
(can provide doctors with both the heavy
metal toxicity fecal test and Detoxamin)



Safety Profile

**World Health Products, LLC
12685 South 125 East
Draper, Utah USA 84020
877-656-4553**

Safety Profile

Detoxamin Safety Overview

DETOXAMIN®

U.S. Patent 5,602,180
Calcium Disodium EDTA
Calcium Disodium EDTA Time-Release Suppositories
For the removal of toxic heavy metals.

Description

Detoxamin (CaNa₂EDTA) is a rectal, time-release suppository that binds and removes or decreases toxic heavy metal contamination in the body and comes in four strengths 375mg, 750mg and 1000mg and 1500mg. Detoxamin uses calcium disodium EDTA, a synthetic amino acid, in a suppository form as a chelating agent in a cocoa-butter base with methocel E4M premium USP for a time-release effect. The suppositories are a soft solid, bullet-shaped preparation designed for easy self insertion into the rectum through the anus. Each suppository dissolves at body temperature and gradually spreads over the lining of the lower rectum where it is absorbed into the bloodstream.

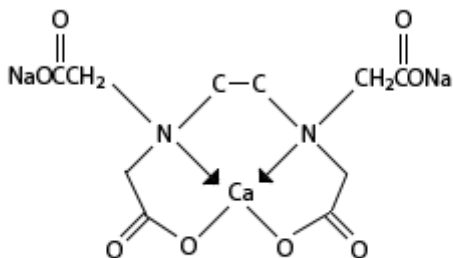
Recent Pharmacokinetic Data

Pre-clinical animal studies suggest that Detoxamin's absolute bioavailability following rectal administration is 36.3 % of the IV dose; this was ascertained by C14 labeled radio-labeled calcium disodium EDTA (Detoxamin). The ratio of radioactive residues of EDTA in prostate tissues showed a mean value of 13.6 via the rectal route compared to 3.69 via the IV route. The total recovery of C14 EDTA expressed as percent of administered dose was 30.3 % rectally and 47.3% IV after eight hours. The study reflected a standard pharmacokinetic model for IV dosing, whereas a unique bi-phasic curve was observed in rectal administration over the same eight hour period. The suppository Detoxamin formulation appears to be well absorbed delivering high levels of EDTA in prostate tissue and is efficiently excreted.

Structural Formula

Edetate Calcium Disodium, USP, CAS 23411-34-9;
Calcium Disodium Ethylenediaminetetraacetate; Calcium
Disodium (Ethylenedinitrilo) Tetraacetate; Calcium
Disodium Edetate; Ethylenediaminetetraacetic Acid,
Calcium Disodium Salt) USP

C₁₀H₁₂CaN₂Na₂O₈ · 2H₂O



Clinical Pharmacology

The pharmacologic effects of CaNa₂EDTA are due to the formation of chelates that bind with any metal that has the ability to displace calcium from the molecule, a feature shared by lead, zinc, cadmium, manganese, iron and mercury and other metals. Intravenous studies have shown that EDTA is distributed primarily in the extracellular fluid with only about 5% of the plasma concentration found in spinal fluid. Almost none of the compound is metabolized. The primary source of lead chelated by EDTA is from bone, subsequently, soft-tissue lead is redistributed to the bone when chelation is stopped. (3, 4) It has been shown in animals that following a single dose EDTA, urinary lead output increases and the blood lead concentration decreases with subsequent decreases in internal redistribution of lead. (5)

Detoxamin, CaNa₂EDTA may have an altered pharmacological profile from IV EDTA due to the rectal route of administration.

Pre-Clinical Toxicology Studies

The acute oral LD₅₀ of calcium EDTA for the rat was found to be 10,000 ± 740 mg per kilogram body weight and for the rabbit and dog, approximately 7 and 11 g. respectively. The acute toxicity in rats was not altered by prior feeding of a diet suboptimal in respect to calcium, iron, copper and manganese.

In 2-year feeding studies with rats receiving diets containing calcium EDTA at levels to provide 50, 125 or 250 mg per kilogram body weight no adverse effects on growth or food efficiency were observed. Hematological examinations conducted periodically and determination of prothrombin time, blood sugar, NPN and serum calcium were likewise normal throughout the test period. Responses similar to those seen in the parent generation were observed in the rats of the three succeeding generations maintained on the same diet. Under the stresses of repeated pregnancies and lactation, no adverse effect of calcium EDTA was observed as measured by any of the usual indexes of reproduction or lactation efficiency. At autopsy neither gross examination nor the weights of the major organs disclosed any significant differences between the test and control groups. The histopathologic findings likewise revealed no consistent or dose-related effects.

Observations on the incidence and severity of dental caries, and the “line tests” of the tibias failed to suggest any evidence of an adverse effect on the calcification processes. Determinations of the xanthine oxidase content in liver tissue at autopsy, and on carbonic

anhydrase content of the blood, revealed no significant changes resulting from chronic ingestion of calcium EDTA. The normal physiologic responses and behavior of the rats, even under the stresses of reproduction through successive generations, are consistent with the lack of effect of the chelating agent on these or other important metallo-enzyme systems.

Groups of dogs were fed diets furnishing 0, 50, 100 and 250 mg of calcium EDTA per kilogram body weight for 1 year. Every dog gained weight during the test period, regardless of dietary treatment. The hematologic findings suggest that the dogs at all dosage levels were in an even better state of health after 1 year of test feeding than they were initially. No significant deviations from normal or control values were noted throughout the test period in urine or in blood chemistry, including the values for prothrombin time. Roentgenographic examinations of the rib cages and leg bones of the dogs in the highest dosage group and of the femurs of all dogs at or near the termination of the test period showed no evidence of osseous change.

All dogs survived the 1-year test period. No gross aberrations were seen at autopsy, and the weights of the liver, kidneys, spleen, heart, adrenals and gonads were all within normal limits. Histopathologic findings of the liver, kidneys, pituitary, adrenals and 12 additional organs in the dogs of the highest dosage group were negative (6)

Ca-disodium EDTA LD₅₀

| <u>Animal</u> | <u>Route</u> | <u>(mg/kg bw)</u> | <u>References</u> |
|---------------|--------------|-------------------|--------------------|
| Rat | oral | 10,000 ± 740 | Oser et al., 1963 |
| Rabbit | oral | 7,000 approx. | Oser et al., 1963 |
| Rabbit | i.p. | 500 approx. | Bauer et al., 1952 |
| Dog | oral | 12,000 approx. | Oser et al., 1963 |

CaNa₂EDTA produced minimal focal hydropic kidney changes in 58% of animals, disappearing almost two weeks after stopping the injections (7).

Short-term studies

Rat

Groups of five male rats received 250 or 500 mg/kg bw CaNa₂EDTA i.p. daily for three to 21 days and some were observed for an additional two weeks. Weight gain was satisfactory and histology of lung, thymus, kidney, liver, spleen, adrenal, small gut and heart was normal except for mild to moderate renal hydropic change with focal subcapsular swelling and proliferation in glomerular loops at the 500 mg level. There was very slight involvement with complete recovery at the 250 mg level. Lesions were not more severe with simultaneous cortisone administration (7).

Groups of three male and three female rats were fed for four months on a low mineral diet containing one-half the usual portion of

salt mixture (i.e. 1.25% instead of 2.50%) with the addition of 0% and 1.5% CaNa_2EDTA . The test group showed a reduced weight gain, but there was no distinct difference in general condition of the animals (8).

In another experiment three groups of eight to 13 male and female rats were fed a low-mineral diet containing 0, 0.5 and 1% of CaNa_2EDTA for 205 days. No significant differences from the controls were shown regarding weight gain, mortality, gross pathology of the organs and histopathology of liver, kidney and spleen except a very slight dilatation of hepatic sinusoids. Blood coagulation time, total bone ash and blood calcium level were unaffected. No significant erosion of molars was noted. Basal metabolism was in the normal range (9).

Dog

Four groups of one male and three female mongrels were fed diets containing 0, 50, 100 and 250 mg/kg bw CaNa_2EDTA daily for 12 months. All appeared in good health, without significant change in blood cells, hemoglobin and urine (pH, albumin, sugar, sediment). Blood sugar, non-protein nitrogen and prothrombin time remained normal. Radiographs of ribs and of long bones showed no adverse changes at the 250 mg level. All dogs survived for one year. Gross and microscopic findings were normal (6).

Long-term studies

Rat

Four groups of 25 male and 25 female rats were fed diets containing 0, 50, 125 and 250 mg/kg bw CaNa_2EDTA for two years. Feeding was carried on through four successive generations. Rats were mated after 12 weeks' feeding and allowed to lactate for three weeks with one week's rest before producing a second litter. Ten male and 10 female rats of each group (F_1 generation) and similar F_2 and F_3 generation groups were allowed to produce two litters. Of the second litters of the F_1 , F_2 and F_3 generations only the control and the 250 mg/kg bw groups were kept until the end of two-years' study on the F_0 generation. This scheme permitted terminal observation to be made on rats receiving test diets for 0, 0.5, 1, 1.5 or 2 years in the F_3 , F_2 , F_1 and F_0 generations, respectively. No significant abnormalities in appearance and behavior were noted during the 12 weeks of the post weaning period in all generations. The feeding experiment showed no statistically significant differences in weight gain, food efficiency, haemopoiesis, blood sugar, non-protein nitrogen, serum calcium, urine, organ weights and histopathology of liver, kidney, spleen, heart, adrenals, thyroid and gonads. Fertility, lactation and weaning were not adversely affected for each mating. Mortality and tumor incidence were unrelated to dosage level. The prothrombin time was normal. There was no evidence of any chelate effect on calcification of bone and teeth. Liver xanthine oxidase and

blood carbonic anhydrase activities were unchanged (6).

Comments:

CaNa₂EDTA is metabolically inert and no accumulation in the body has been found. A vast clinical experience in its use in the treatment of metal poisoning has demonstrated its safety in man. Long-term feeding studies in rats and dogs gave no evidence of interference with mineral metabolism in either species. Adverse effects on mineral metabolism and nephrotoxicity were only seen after parenteral administration of high doses.

Clinical Safety

Two carefully monitored clinical trials revealed no significant adverse effects, either patient reported or within blood chemistry panels of those on the studies. The researchers at the Living Longer Institute, Cincinnati, OH state that the dosage form is gentle and appears to create little biological burden and is well tolerated. Only minor transient complaints of loose stools, rectal gas, headache, lethargy and minor joint pain were reported.

Within the Living Longer clinical the following lab parameters were compared between pre and post treatment with Detoxamin in all subjects:

Comprehensive Metabolic Panel

- | | |
|------------------------|----------------------------------|
| – Albumin | - Total Protein |
| – Total Bilirubin | - Sodium |
| – Calcium | - AST (SGOT) |
| – Chloride | - Urea Nitrogen (BUN) |
| – Creatinine, Serum | - Bicarbonate (CO ₂) |
| – Glucose | - ALT (SGPT) |
| – Alkaline Phosphatase | - C-Reactive Protein |
| – Potassium | |

No statistical difference in above lab parameters between pre and post treatment were observed with Detoxamin.

Dr. Ted Rozema performed a study with 20 children with high lead blood levels utilizing 1000 mg Detoxamin suppositories over a thirty-day period. No adverse effects were reported.

Safety in Clinical Practice

Dr. Rita Ellithorpe, MD has treated over 1800 patients with Detoxamin and administered over 100,000 doses of the product over a six-year period; other than minor discomfort complaints that were dismissed due to other meds or lifestyle events occurring within a very small percentage of those on the product. Dr. Ellithorpe observed no significant adverse effects and no aberrant blood chemistry values in her practice with the use of Detoxamin.

Post Marketing Safety

There have been over 2,500,000 doses of Detoxamin administered to over 50,000 patients worldwide, with over 2500 health care professionals recommending Detoxamin to their patients within the last eight years. The most common minor complaints were loose stools and gas experienced in the first few applications. Detoxamin is safe for children.

Side Effects

Detoxamin in the suppository form appears to have few minor side effects. Renal excretory function should be within normal limits prior to treatment. Daily urinalysis is not required when using Detoxamin suppositories. With Detoxamin the following side effects may occur rarely: loose stools, headache, rectal discomfort, nausea, and loss of appetite. These symptoms are usually rare and transient. The most common complaints are loose stools and gas experienced in the first few applications.

Usage in Pregnancy

Safe use of CaNa₂EDTA suppositories has not been established with respect to adverse effects on human fetal development. However, animal studies on fetal development utilizing high doses of EDTA have shown no ill effects for several generations. It is recommended that Detoxamin should not be used in women of childbearing potential and particularly during pregnancy only when, in the judgment of the physician, the potential benefits outweigh the possible hazards.

Precautions

Treatment with Detoxamin has been shown to cause a lowering of blood sugar and insulin requirements in patients with diabetes who are treated with insulin. Diabetic patients should check their insulin and glucose levels. Adverse side effects are extremely rare. Detoxamin exhibits no known adverse renal, hepatic, cardiovascular, gastrointestinal or central nervous system effects.

References

1. Thomas DJ Chisolm JJ Lead, zinc and copper decorpration during calcium disodium ethylenediamine tetraacetate treatment of lead-poison children. J Pharmacol Exp. Therapeu 1986: 239: 829-835.
2. The Pharmacological Basis of Therapeutics 7th edition, Goodman and Oilman, editors. Macmillan Publishing Company: New York, 1985, pp. 1619-1622
3. Hammond PB, Aronson AL, Olsen We, The mechanism of mobilization of lead by ethylenediaminetetraacetate, J Pharmocol Exp. therapeu 1967: 157: 196-206
4. Van deByer FL, D'Haese PC, Visser WJ et al. Bone lead in dialysis patients. Kidney Int; 1988: 33: 601-60.
5. Cory-SlectaDA, WeissBCoxC. Mobilization and redistribution of lead over the course of calcium disodium elhylenediamineetraacetate acetate dictation therapy. J Pharmacol Exp. Therapereu 1987: 243: 804-813
6. Oser, B. L., Oser, M. & Spencer, H. C. (1963) Toxicol. appl. Pharmacol., 5, 142
7. Reuber, M. D. & Schmieller, G. C. (1962) Arch. environ. Health, 5, 430

8. Yang, S. S. (1964) Fd. Cosmet. Toxicol., 2, 763
9. Chan, M. S. (1964) Fd. Cosmet. Toxicol., 2, 763-765
10. Chisolm JJ Mobilization of lead by calcium disodium edetate AM J Dis Child 1987.141: 1256-1257

Other Resources

11. Drug Evaluations, 6th Edition, American Medical Association, Saunder, Philadelphia, 1986, pp 1637-1639
12. Centers for Disease Control: Preventing lead poisoning in young children. Atlanta, GA. Department of Health and Human Services, 1985 Jan.
13. Finberg L. Rajagopal V. Diagnosis and treatment of lead poisoning in children. J. Family Med 1985 April 3-12
14. Schardein JL. Sakowski R Petere J. et al. Teralogenesis studies with EDTA and its salts in rats, Toxicol Appl Pharmacol 1981: 61: 423-428
15. Swenerton H. Hurly LS Teratogenic effects of chelating agent and their prevention by zinc. Science 1971: 173:62-64
15. American Hospital Formulary Service. Drug Information. 1988. Pp. 1695-1698

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**Tustin
Longevity
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Rita Ellithorpe, M.D.

13422 Newport Avenue Suite L
Tustin, CA 92780
(714) 544-1521 ph www.tlcmd.net
(714) 544-1904 fax rellithorpe@tlcmd.net

STATEMENT OF CLINICAL EXPERIENCE WITH
DETOXAMIN (CaNa₂ EDTA)

Rita R. Ellithorpe, M.D.

I have been familiar with E.D.T.A. chelation therapy for the treatment of potentially toxic heavy metals for most of my life, having received this therapy myself as early as the age of sixteen. I have seen several of my own family members receive this therapy on multiple occasions for primarily its cardiovascular enhancing effects. Each of these personal experiences was positive and was completed without any adverse side effect. Having now reached the age of fifty and obtaining some distance from the completion of Medical School (North Chicago Medical School/1978-82) and an internship in family practice (Womack Army Medical Center, Fort Bragg, NC/1982-83), I have now administered this FDA approved chelating agent to countless patients of mine as well as to myself and to my family members. Most of this therapy was made available through the intravenous form of chelation until the convenience and vastly more affordable form of suppositories (Detoxamin - CaNa₂ EDTA) which has become available to me approximately five years ago. I am truly pleased with the effectiveness in this form for the reduction of potentially toxic heavy metals in appropriately screened, eligible patients. I can state without reservation the safety and health-enhancing potential of this product for this industrialized, toxic environment we live in is enormous. I have treated over 1800 patients with Detoxamin and administered over 100,000 doses of the product and observed no significant adverse effects and no aberrant blood chemistry values. Our results at my clinic indicate this has been a great help in restoring and preserving patient health.

Toxic Heavy Metal Diagnosis

Fecal Metal Analysis

We recommend an initial stool analysis to detect the presence and quantity of toxic heavy metals in your patients. This is a simple, relatively easy procedure that is done at home and mailed to the diagnostic laboratory. Our clinical trials have shown significant excretions of heavy metals with Detoxamin treatment over time.

It is suggested to perform a fecal test before Detoxamin is administered and follow up after month of treatment (one suppository 4 evenings per week for six months).

The fecal metal analysis offers the following advantages to health practitioners:

1. Assessment of exposure to, and excretion of, toxic metals and elements
2. Monitor natural route of metal detoxification
3. Convenient specimen collection procedure
4. Analysis by ICP-MS
5. Result specific commentary provided
6. Correlates to significant findings increased of metal excretion with Detoxamin

Description

Analysis of elements in feces provides important information about the potential for toxic metal burden. For many toxic metals, fecal (biliary) excretion is the primary natural route of elimination from the body. Fecal elemental analysis also provides a direct indication of dietary exposure to toxic metals. Specimen collection is convenient for the patient and only requires a single-step procedure.

Analysis of elements in feces provides a comprehensive evaluation of environmental exposure, potential for accumulation in the body, and possibly endogenous detoxification of potentially toxic metals. For many toxic elements such as mercury, cadmium, lead, antimony and uranium, biliary excretion into the feces is the primary natural route of elimination from the body. The primary process by which the body eliminates the insidious sulfhydryl reactive metals is through the formation of metal-glutathione complexes, of which greater than 90% are excreted into the bile. Evidence for the extent of exposure to mercury from dental amalgams is provided by the fact that fecal mercury levels are highly correlated with the number of amalgams in the mouth.* It also clear that fecal mercury levels for people with dental amalgams are remarkably similar from day to day, and approximately ten times higher than in people who do not have mercury amalgams.

Administration of pharmaceutical metal binding agents results in excretion of toxic metals primarily through the kidneys into the urine. In contrast, support of natural detoxification processes enhances the rate of excretion of toxic metals into the feces. Elemental analysis of fecal specimens can provide a valuable tool to monitor the

efficacy of natural detoxification of metals in infants or patients who are on very limited and defined diets that do not contain contaminated solid foods. A preliminary study performed at Doctor's Data indicates that biliary/fecal excretion of mercury and lead may be markedly enhanced following high dose intravenous administration of ascorbic acid. Other orthomolecular or nutraceutical protocols may also enhance the fecal excretion of metals and hence potentially decrease burden on the kidneys. Further research to identify and validate such therapies is warranted.

A primary objective of preventive medicine is avoidance or removal of exposure to toxic substances. The rate of oral absorption of toxic metals varies considerably among elements, and among subspecies of a particular element. Fecal elemental analysis can provide a direct indication of dietary exposure. Orally, the percent absorption of nickel, cadmium and lead is usually quite low, but varies significantly in part due to the relative abundance of antagonistic essential elements in the diet. That is particularly evident for lead and calcium, and cadmium and zinc. Chronic, low-level assimilation of the toxic metals can result in significant accumulation in the body. The results of fecal elemental analysis can help identify and eliminate dietary exposure to toxic metals.

The fecal metals test was not developed to replace the pre and post urinary toxic metals provocation test, but rather provides an alternative for infants, children or adults for whom urine collection is problematic, or for individuals who do not tolerate the available pharmaceutical metal detoxification agents. Elements are measured by ICP-MS and expressed on a dry weight basis to eliminate variability related to water content of the specimen.

Fecal metal tests can be ordered directly from Doctor's Data, Inc:

Doctor's Data, Inc.
3755 Illinois Avenue
St. Charles, IL 60174-2420
800.323.2784

Optional Heavy Metal Analysis

Red Blood Cell Elements

1. Measurement of toxic and functional intracellular elements
2. Analysis by ICP-MS
3. Result specific commentary provided
4. Requires unwashed packed red blood cells

Analysis of red blood cells provides the best diagnostic tool for assessing the status of elements that have important functions inside cells or on blood cell membranes. Blood cell element levels are useful for assessing cardiac influences, anti-inflammatory processes, anemia, immunological function, glucose tolerance and other disorders that are associated specifically with zinc deficiency.

Urine Toxic Metals

1. Assessment of toxic element burden and essential element wasting
2. Monitors detoxification therapy
3. Analysis by ICP-MS
4. Interpretive report provided
5. Variable urine collection periods

Analysis of elements in urine provides diagnostic information on potentially toxic elements such as lead, mercury, cadmium, nickel, beryllium, arsenic and aluminum, and assessment of the efficiency of renal resorption of essential elements such as magnesium, calcium, sodium and potassium.

Hair Toxic Element Exposure Profile

1. Measurement of toxic and essential elements
2. Inexpensive, noninvasive
3. Analysis by ICP-MS
4. Result specific commentary provided
5. Requires 0.25 g hair

Extensive research established that scalp hair element levels are related to human systemic levels. The strength of this relationship varies for specific elements, and many researchers consider hair as the tissue of choice for toxic and several nutrient elements. Unlike blood, hair element levels are not regulated by homeostatic mechanisms. Thus, deviations in hair element levels often appear prior to overt symptoms and can thereby be a valuable preliminary tool for predicting the development of physiological abnormalities.

EDTA and Heavy Metal Toxicity References

1. Ahrens FA and Aronson AL: A comparative study of the toxic effects of calcium and chromium chelates of ethylenediaminetetraacetate in the dog. *Toxic Appl Pharmac* 18:10, 1971.
2. Ahrens FA and Aronson AL: Toxicity of calcium ethylenediaminetetraacetate in dogs. *Fed Proc Fedn Am Soc Exp Biol* 27:1401, 1968.
3. Adams WJ and McGee CT: Chelation therapy: a survey of treatment outcomes and selected socio-medical factors. *J Adv Med* 5(3):189, 1992.
4. Agerty HA: Lead poisoning in children. *Medical Clinics of North America*. (36):1587, 1952.
5. Ahrens FA and Aronson AL: Comparative study of the toxic effects of calcium chelates of ethylenediaminetetraacetate in the dog. *Tox Appl Pharmacol* (18)1:10 1971.
6. Alfthan G, Pikkarainen J, Huttunen JK, Puska P: Association between cardiovascular death and myocardial infarction and serum selenium in a matched- pair longitudinal study. *Lancet* 2(8291):175, 1982.
7. Allen AC: *The Kidney* Grune and Stratton, N.Y., p. 329, 1962.
8. Altmann J, Wakim KG and Winkelmann RK: Effects of edathamil disodium on the kidney. *J Invest Dermatol* (38): 215, 1962.
9. Angle CR and McIntire MS: Lead poisoning during pregnancy. Fetal tolerance of calcium disodium edetate . *Am J Dis Child* 108:436, 1964.
10. Ames BN: Dietary carcinogens and anticarcinogens. *Science* 221:1256, 1983.
11. Aronov DM: First experience with the treatment of atherosclerosis patients with calcinosis of the arteries with trilon-B (disodium salt of EDTA). (Russ, Moscow) *Klin Med* 41:19, 1963.
12. Aronson AL and Ahrens FA: The mechanism of renal transport and excretion of ethylenediaminetetraacetate with interspecies comparison. *Toxic Appl Pharmac* 18:1, 1971.
13. Aronson AL and Hammond PB: Effect of two chelating agents on the distribution and excretion of lead. *J Pharmacol Exp Ther* 146:241-251, 1964.
14. Aronson AL, Hammond PB and Straffuss AC: Studies with calcium ethylenediaminetetraacetate in calves; toxicity and use in bovine lead poisoning. *Toxicol Appl Pharmacol* 12:337-349, 1968.
15. Atherosclerosis and auto oxidation of cholesterol, editorial. *Lancet* ii:964, 1980.
16. Batchelor TM, McCall M and Mosher RM: Potassium dieresis induced by edathamil disodium. *JAMA* 187:305, 1964.
17. Bates GW, Billups C and Saltman P: The kinetics and mechanism of iron (III) exchange between chelates and transferrin. II. The presentation and removal with ethylenediaminetetraacetate. *J Biol Chem* 242:2816, 1967.

18. Batuman V, Landy E, Maesaka JK, Wedeen RP: Contribution of lead to hypertension with renal impairment. *N Engl J Med* 309(1):17, 1983.
19. Batuman V, Maesaka JK, Haddad B, et al: The role of lead in gout nephropathy. *N.E.J.M.* (304)9:520, 1981.
20. Bauer RO, Rullo FR, Spooner C, et al: Acute and subacute toxicity of ethylene diamine tetraacetic acid (EDTA) salts. *Federation Proc* (11): 321, 1952.
21. Bechtel JT, White JE and Estes EH Jr: The electrocardiographic effects of hypocalcemia induced in normal subjects with edathamil disodium. *Circulation* 13:837, 1956.
22. Belknap EL: EDTA in the treatment of lead poisoning. *Indust Med & Surg* (21):305, 1952.
23. Berkman N, Michaeli Y, Or R and Eldor A: EDTA dependent pseudothrombocytopenia: a clinical study of 18 patients and a review of the literature. *Am J Hematol* 36:195, 1991.
24. Bessman SP, Ried H and Rubin M: Treatment of lead encephalopathy with calcium disodium versenate. *Ann Med Soc DC* 21:312, 1952.
25. Bhat RK, et al: Trace elements in hair and environmental exposure. *Sci Total Environ* 22(2):169, 1982.
26. Birk RE and Rupe CE: The treatment of systemic sclerosis with disodium EDTA, pyridoxine and reserpine. *Henry Ford Hosp Med Bull* 14:109, 1966.
27. Birk RE and Rupe CE: Systemic sclerosis. Fourteen cases treated with chelation (disodium EDTA) and/or pyridoxine, with comments on the possible role of altered tryptophan metabolism in pathogenesis. *Henry Ford Hosp Med Bull* 10:523, 1962.
28. Bjorksten J: The cross-linkage theory of aging as a predictive indicator. *Rejuvenation* 8:59, 1980.
29. Bjorksten J: Possibilities and limitations of chelation as a means for life extension. *Rejuvenation* 8:67, 1980.
30. Blake DR, Hall ND, Bacon PA, Dieppe PA, Halliwell B, Gutteridge JM: The importance of iron in rheumatoid disease. *Lancet* :1142, 1981.
31. Blake DR, Hall ND, Bacon PA, Dieppe PA, Halliwell B, Gutteridge JMC: Effect of a specific iron chelating agent on animal models of inflammation. *Ann Rheum Dis* 42:89, 1983.
32. Bland JS: The utility of hair analysis: A reevaluation. *J Holistic Medicine* 5(1):16, 1983.
33. Blumer W, Reich T: Leaded gasoline - cause of cancer. *Environmental International* 3:465, 1980.
34. Blumer W and Cranton EM: Ninety percent reduction in cancer mortality after chelation therapy with EDTA. *J Adv Med* 2:183, 1989.
35. Bolick LE and Blankenhorn DH: A quantitative study of coronary arterial calcification. *Am J Pathol* 39:511, 1961.
36. Boyle AJ, Jasper JJ, McCormick H, et al: Studies in human and induced atherosclerosis employing - (EDTA). *Bull Swiss Acad Med Sci* 13:408, 1957.

37. Boyle M, Wegria R, Cathcart RT, et al: Effects of intravenous injections of nicotine on the circulation. *Am Heart J* (34): 65, 1947.
38. Boyle AJ, Clarke NE, Mosher RE, McCann DS: Chelation therapy in circulatory and sclerosing diseases. *Fed Proc* 20(3) (Part II) (suppl 10):243-257, 1961.
39. Boyle AS, Mosher LE, McCann DS: Some in vivo effects of chelation-I: Rheumatoid arthritis. *J Chronic Dis* 16:325, 1963.
40. Brachet P and Klein C: Cell response to CAMP during aggregation phase of Dictyostelium discoideum. Comparison of the inhibitory effects of progesterone and the stimulatory action of EDTA and ionophore A23187. *Differentiation* (8)1:1, 1977.
41. Brecher A: Bye-Bye Bypass: The Truth About Chelation Therapy. Troup, Texas, Health Savers Press, 1989.
42. Brien TG and Fay J: [51Cr] EDTA biological half life as an index of renal function. *J Nuc Med.* (13) 5:339, 1972.
43. Brecher A: Bye-Bye Bypass: The Truth About Chelation Therapy. Troup, Texas, Health Savers Press, 1989.
44. Brucknerova O, Tulacek J and Krojzl O: Chelates in the treatment of obliterating arteriopathies. *Vnitřní Lek* 14:841, 1968.
45. Brucknerova O and Malinovska V: First clinical experience with combined treatment with chelation III and glucagon in ischemic disease of the lower extremities, *Cas Lek Cas* 119:814, 1980.
46. Brucknerova O, Tulacek J: Chelates in the treatment of occlusive atherosclerosis. (Czech, Praha) *Vnitřní Lek* 18:729, 1972.
47. Burckhardt P, Boillat AM, Reudi B, et al: Effect of parathyroid hormone immunoheterogeneity. *Schweiz Med Wochenschr* (105) 50:1692, 1975.
48. Butler AM: Use of calcium ethylenediaminetetraacetate in treating heavy-metal poisoning. *Arch Indust Hyg Occupat Med* 7:136, 1952.
49. Butterfield JD, McGraw CP: Free radical pathology. *Stroke* 9(5):443, 1978.
50. Calcium disodium edetate and disodium edetate. Drugs for human use; drug efficacy study implementation. Department of Health, Education, and Welfare. Food and Drug Administration. Federal Registrar 35 F.R. 437, 1970.
51. Casdorff HR: Chelation therapy: a reappraisal. *N Z Med J* 66, 1983.
52. Casdorff HR: EDTA chelation therapy, efficacy in arteriosclerotic heart disease. *J Holistic Medicine* 3(1):53, 1981.
53. Casdorff HR: EDTA chelation therapy II, efficacy in brain disorders. *J Holistic Medicine* 3(2):101, 1981.

54. Casdorph HR, Farr CH: EDTA chelation therapy III: Treatment of peripheral arterial occlusion, an alternative to amputation. *J Holistic Medicine* 3(1):3, 1983.
55. Cashion WR Jr: What about chelation? *Texas Medicine* 80:6, 1984.
56. Castellino N and Aloj S: Effects of calcium sodium ethylenediaminetetra-acetate on the kinetics of the distribution and excretion of lead in the rat. *Brit J Indust Med* 22:172, 1965.
57. Catsch A: Radioactive metal mobilization. *Fed Proc* 20 (Supl 10):206, 1961.
58. Catsch A and Harmuth-Hoene AE: Pharmacology and therapeutic applications of agents used in heavy metal poisoning. *Pharmac Ther. A*, (1):1, 1976.
59. Chaitlow L: *Chelation Therapy: The Revolutionary Alternative to Heart Surgery*. San Francisco, Thorsons SF, 1991.
60. Chantler C and Barratt TM: Estimation of glomerular filtration rate from plasma clearance of 51-chromium edetic acid. *Arch Dis Child* 47:613, 1972.
61. Chantler C, Garnett ES, Parsons V and Veall N: Glomerular filtration rate measurement in man by the single injection method using 51Cr-EDTA. *Clin Sci* 37:169, 1969.
62. Chappell LT and Stahl JP: The correlation between EDTA chelation therapy and improvement in cardiovascular function: a meta-analysis. *J Adv Med* 6(3):139, 1993.
63. Chappell LT, Stahl JP and Evans R: EDTA chelation treatment for vascular disease: a meta-analysis using unpublished data. *J Adv Med* 7(3):131, 1994.
64. Chappell LT, Janson M, Whitaker J: A challenge to cardiovascular surgeons. *J Adv Med* (In Press)
65. Chappell LT: Chelation therapy, smoking and health care costs (Letter). *J Adv Med* 7:107, 1994.
66. Chappell LT: EDTA chelation therapy should be more commonly used in the treatment of vascular disease. *Alternative Therapies Health Med* 1:53, 1995.
67. Chappell LT, Miranda R, Rubin M, Carter JP and Trowbridge J: Chelation therapy (Letter). *Circulation* 92:1350, 1995.
68. Chelation therapy (Diagnostic and therapeutic technology assessment). *JAMA* 250:672, 1983.
69. Chelation therapy: a second look. *The Harvard Medical School Health Letter*, IX:1, 1984.
70. Chelation Therapy Clinic: Chelation therapy - the treatment of choice for relief from and prevention of, cardiovascular and age-related diseases. Auckland, New Zealand, The Chelation Therapy Clinic, 1987.
71. Chelation therapy. Resolution: 66 (I-84). AMA House of Delegates. 1984.
72. Chelation therapy. Report of the Council on Scientific Affairs. Report F (I-84). AMA House of Delegates. 1984.
73. Chelation therapy - An informal summary. Department of Health & Human Services. National Institutes of Health. National Heart, Lung, and Blood Institute. Bethesda, Maryland. June 1992.

74. Chen IW, Park HM, King LR, Bahr GK and Goldsmith RE: Radioimmunoassay of parathyroid hormone: peripheral plasma immunoreactive parathyroid hormone response to ethylenediaminetetraacetate. *J Nucl Med* 15:763, 1974.
75. Cheraskin E, Wussow DG, McDonagh EW and Rudolph CJ: Effect of EDTA chelation and supportive multivitamin/trace mineral supplementation with and without physical activity on the heart rate. *J Intern Acad of Prev Medi* 8(6):5, 1984.
76. Chisolm JJ Jr: Chelation therapy in children with subclinical plumbism. *Pediatrics* 53:441, 1974.
77. Chisolm JJ Jr: The use of chelating agents in the treatment of acute and chronic lead intoxication in childhood. *J Pediat* 73:1, 1968.
78. Clarke NE Sr, Clarke NE Jr, Mosher RE: Treatment of occlusive vascular disease with disodium ethylene diamine tetraacetic acid (EDTA). *The Amer J of the Med Sci* June:732, 1960.
79. Clarke NE, Clarke CN, and Mosher RE: Treatment of angina pectoris with disodium ethylene diamine tetra-acetic acid. *Am J of Med Sci* 232:654, 1956.
80. Clarke NE Sr: Atherosclerosis, occlusive vascular disease and EDTA. *Am J Cardiol* 6(2):233, 1960.
81. Clarke NE, Clarke CN, Mosher RE: The "in vivo" dissolution of metastatic calcium: An approach to atherosclerosis. *Am J Med Sci* 229:142, 1955.
82. Cockcroft DW, Gault MH: Prediction of creatinine clearance from serum creatinine. *Nephron* 16:31, 1976.
83. Cohen SH, Gong JK and Fishler MC: Ethylene diamine tetraacetic acid (EDTA) treatment of internal radioactive contamination. *Nucleonics* (11)1: 56, 1953.
84. Cohen S, Weissler AM and Schoenfeld CD: Antagonism of the contractile effect of digitalis by EDTA in the normal human ventricle. *Am Heart J* 69:502, 1965.
85. Cook JD, Finch CA, Smith NJ: Evaluation of the iron status of a population. *Blood* 48(3):449, 1976.
86. Couric JM: Chelation therapy overdone. *FDA Consumer* 16:28. 1982.
87. Cranton EM and Frackelton JP: Negative Danish Study of EDTA chelation biased. *Townsend Letter for Doctors* July, 604-605, Letter, 1992.
88. Cranton EM: Critique of the American Medical Association's published position on hair analysis. *J Holistic Medicine* 8(1):47, 1986.
89. Cranton EM, Bland JS, Chatt A, Krakovitz R, Wright JV: Standardization and interpretation of human hair for elemental concentrations. *J Holistic Medicine* 1982;4:10, 1982.
90. Cranton EM: Update on hair analysis in clinical medicine. *J Holistic Medicine* 7(2):120, 1986.
91. Cranton EM and Frackelton JP: The current status of EDTA chelation therapy in the treatment of occlusive arterial disease. *Journal of Holistic Medicine* 4: 24, 1982.

92. Cranton EM: Protocol of the American College of Advancement in Medicine for the safe and effective administration of EDTA chelation therapy. *J Adv Med* 2:269, 1989.
93. Cranton EM: A textbook on EDTA chelation therapy. *J Adv Med* 2:1-416, 1989.
94. Cranton EM and Frackelton JP: Free radical pathology in age associated diseases; treatment with EDTA chelation, nutrition and antioxidants. *J Holistic Med* 6: 1, 1984.
95. Cranton EM: Kidney effects of ethylene diamine tetraacetic acid (EDTA): A literature review. *J Holistic Medicine* 4:152, 1982.
96. Cranton EM: The current status of EDTA chelation therapy, editorial. *J Holistic Med* 7(1):3, 1985.
97. Cranton EM, Brecher A: *Bypassing Bypass: The New Technique of Chelation Therapy* ed 2 ,Medex Publishers Inc, P.O. Box 44, Trout Dale, VA 24378, 1989.
98. Craven, PC and Morelli HF: Chelation therapy. *West J Med*, 122:277, 1975.
99. Curran CL: Metal chelating agents and hepatic cholesterol synthesis. *Proc Soc Exper Biol & Med* 88:101, 1955.
100. Darwish NM and Kratzer FH: Metabolism of ethylenediaminetetraacetic acid (EDTA) by chickens. *J Nutr* 86:187, 1965.
101. David O, Hoffman SP, Sverd J, Clark J, Voeller K: Lead and hyperactivity, behavioral response to chelation: A pilot study. *Am J Psychiatry* 133:1155, 1976.
102. Davis FA, Becker FO, Michael JA and Sorensen E: Effect of intravenous sodium bicarbonate disodium EDTA and hyperventilation on visual and oculomotor signs in multiple sclerosis. *J Neurol Neuro-surg Psychiat* 33:723, 1970.
103. Davis PS and Deller DJ: Effect of orally administered chelating agents EDTA, DTPA and fructose on radioiron absorption in man. *Aust Ann Med* 16:70, 1967.
104. Davis H and Moe PJ: Favorable response of calcinosis universalis to edathamil disodium. *Pediatrics* 24:780, 1959.
105. Deftos LJ, Goodman AD, Engleman K, et al: Suppression and stimulation of calcitonin secretion in medullary thyroid carcinoma. *Metab Clin Med.* (20)4: 428, 1971.
106. Del Maestro RF: An approach to free radicals in medicine and biology. *Acta Physiol Scand* 492(suppl):153, 1980.
107. Demopoulos HB: The basis of free radical pathology. *Fed Proc* 32:1859, 1973.
108. Demopoulos, HB: Control of free radicals in the biologic systems. *Fed Proc* 32: 1903, 1973.
109. Demopoulos HB, Pietronigro DD, Flamm ES, Seligman ML: The possible role of free radical reactions in carcinogenesis. *Journal of Environmental Pathology and Toxicology* 3:273, 1980.
110. Demopoulos HB, Flamm ES, Pietronigro DD, Seligman ML: The free radical pathology and the microcirculation in the major central nervous system disorders. *Acta Physiol Scand* 492(suppl):91, 1980.

111. Demopoulos HB: Molecular oxygen in health and disease. Presented AAMP Tenth Annual Spring Meeting May 21, 1983, Los Angeles, California.
112. Demopoulos HB, Peitronigro DD, Seligman ML: The development of secondary pathology with free radical reactions as a threshold mechanism. *J Am Coll Tox* 2(3):173, 1983.
113. Deucher DP: EDTA chelation therapy: an antioxidant strategy. *J Adv Med* 1:182, 1988.
114. Deucher GP: Heavy metals, chelation therapy, free radicals and human disease. *Angiologie*, Octobre 1987.
115. Diagnostic and therapeutic technology assessment: chelation therapy. *JAMA* 250:672, 1983.
116. Dix T: Metabolism of polycyclic aromatic hydrocarbon derivatives to ultimate carcinogens during lipid peroxidation. *Science* 221:77, 1983.
117. Donald GF, Hunter GA, Roman W and Taylor AEJ: Current concepts of cutaneous porphyria and its treatment with particular reference to the use of sodium calcium-edetate. *Brit J Derm* 82:70, 1970.
118. Donsbach KW: Chelation Therapy; Has the Plug Been Pulled on Our Biggest Killer? Huntington Beach, CA, International Institute of Health, 1981.
119. Doolan PD, Schwartz SL, Hayes JR, Mullen JC, Cummings NB: An evaluation of the nephrotoxicity of ethylene diamine tetraacetate and diethylene trisacetate in the rat. *Toxicol Appl Pharmacol* 10:481, 1967.
120. Dormandy TI: Free-radical reaction in biological systems. *Ann R Coll Surg Engl* 62:188, 1980.
121. Dormandy TL: Free-radical oxidation and antioxidants, *Lancet* i:647, 1978.
122. Dormandy TL: An approach to free radicals. *Lancet* 1983; ii:1010, 1983.
123. Dudley HR, Ritchie A, Schilling A, et al: Pathologic changes associated with the use of sodium ethylene diamine tetraacetate in the treatment of hypercalcemia. *NEJM* (252): 331, 1955.
124. EDTA chelation therapy for arteriosclerotic heart disease. *Medical Letter* 23, 1981.
125. EDTA chelation: a rebuttal. *J Adv Med* 5:3, 1992.
126. EDTA chelation therapy for atherosclerotic cardiovascular disease. *Medical Letter* 36:48 1994.
127. Eliot RS and Blout SG Jr: Calcium, chelates and digitalis. A clinical study. *Am Heart J* 62:7, 1961.
128. Elwood PC, Benjamin IT, Waters WE, Sweetnam PM: Mortality and anemia in women. *Lancet* :891, 1974.
129. Emmerson BT: Chronic lead nephropathy. The diagnostic use of calcium EDTA and the association with gout. *Australian Ann Med* 12: 310, 1963.
130. Emmerson BT and Thiele BR: Calcium versenate in the diagnosis of chronic lead nephropathy. *Med J Aust* (1): 243, 1960.

131. Ericson JE, Shirahata H, Patterson CC: Skeletal concentrations of lead in ancient Peruvians. *N Engl J Med* 300:946, 1979.
132. Estep HL, Gardner CT Jr, Taylor JP, Minott A and Tucker HStG Jr: Phosphate excretion patterns following intravenous injection of ethylenediaminetetraacetate ((EDTA). *J Clin Endocr* 25:1385, 1965.
133. Favre HR and Wing AJ: Simultaneous ⁵¹Cr edetic acid, inulin, and endogenous creatinine clearances in 20 patients with renal disease. *Brit Med J* 1:84, 1968.
134. Feigen RD, Shannon DC, Reynolds SL, Shapiro LW and Connelly JP: Lead poisoning in children. *Clin Pediat* 4:38, 1965.
135. Fine SD: Calcium disodium edetate and disodium edetate. Drugs for human use; Drug efficacy study implementation. *Fed Reg* (35)8: Jan. 13, 1970.
136. Fink CW and Baum J: Treatment of calcinosis universalis with chelating agents. *Am J Dis Child* 105:390, 1963.
137. Flamm ES, Demopoulos HB, Seligman ML, Poser RG, Ransohoff J: Free radicals in cerebral ischemia. *Stroke* 9(5):445, 1978.
138. Flynn DM: 5-year controlled trial of chelating agents in treatment of thalassaemia major. *Arch Dis Child* 48:829, 1973.
139. Flytlie K and Hancke C: Letter to the editor: EDTA manipulated. *J Adv Med* 6(1):53, 1993.
140. Foote CS: Chemistry of singlet oxygen VII. Quenching by beta carotene. *J Am Chem Soc* 90:6233, 1968.
141. Foreman H: Pharmacology of some useful chelating agents. In Seven MJ and Johnson LA (eds): *Metal-Binding in Medicine*. Philadelphia, JB Lippincott, 1960.
142. Foreman H: The use of chelating agents for accelerating excretion of radioelements. *J Am Pharm Assoc* (42): 629, 1953.
143. Foreman H: Toxic side effects of ethylenediaminetetraacetic acid. *J Chron Dis* (16):319, 1963.
144. Foreman H: Summary remarks by the chairman. [Proceedings of a conference on biological aspects of metal-binding.] *Fed Proc* 20 (Suppl 10):257, 1961.
145. Foreman H: Use of chelating agents in treatment of metal poisoning with special emphasis on lead). *Fed Proc* 20 (Suppl 10):191, 1961.
146. Foreman H, Finnegan C and Lushbough C: Nephrotoxic hazard from uncontrolled edathamil calcium disodium therapy. *JAMA* (160): 1042, 1956.
147. Foreman H, Vier M and Magee M: The metabolism of C¹⁴ labeled ethylenediaminetetraacetic acid in the rat. *J Biol Chem* 203:1045, 1953.
148. Foreman H, Finnegan C and Lushbaugh CC: Nephrotoxic hazard from uncontrolled edathamil calcium-disodium therapy. *JAMA* 160:1042, 1956.

149. Foreman H, Hardy H, Shipman T, et al: Use of calcium ethylenediaminetetraacetate in cases of lead intoxication. *Ama Archiv Indust Hyg Occup Med* (7): 148, 1953.
150. Foreman H and Trujillo T: Metabolism of carbon 14 labeled ethylenediaminetetraacetic acid in human beings. *J Lab Clin Med* (43): 566, 1954.
151. Forssman O and Nordqvist P: The action in vitro and in vivo of sodium versenate on the phagocytic activity of neutrophile leukocytes. *Acta Haemat* 31:289, 1964.
152. Frackelton JP, Cranton EM: Iron and copper supplementation with EDTA chelation therapy. *J Holistic Med* 8(1):63, 1986.
153. Frackelton JP: Monitoring renal function during EDTA chelation therapy. *J Holistic Med* 8(1):33, 1986.
154. Friedel W, Schulz FH Schroder L: Therapy of atherosclerosis through mucopolysaccharides and EDTA (ethylene diamine tetraacetic acid). (German) *Deutsch Gesundh* 20:1566, 1965.
155. Fromke VL, Lee MY and Watson CJ: Porphyrin metabolism during Versenate® therapy in lead poisoning. Intoxication from an unusual source. *Ann Intern Med* 70:1007, 1969.
156. Fuleihan FJD, Kurban AK, Abboud RT, Beidas-jubran N and Farah FS: An objective evaluation of the treatment of systemic scleroderma with disodium EDTA, pyridoxine and reserpine. *Brit J Derm* 80:184, 1968.
157. Gaby AR: Nutritional factors in cardiovascular disease. *J Holistic Medicine* 5(2):107, 1983.
158. Gerstenblith G, Ouyang P, Achuff S, et al: Nifedipine in unstable angina: A double-blind, randomized trial. *NEJM* (306)15: 885, 1982.
159. Ghiringhelli L and Respighi E: The tubular reabsorption of phosphorus during the test of parathyroid stimulation by versenate (test of Kaiser & Ponsold). *Atti Acad Med Lombarda* 18:191, 1963.
160. Ghiringhelli L, Marinoni E and Respighi E: Study of the parathyroid function by means of the Kaiser & Ponsold test in a group of thyroidectomized patients. *Minerva Med* 54:2829, 1963.
161. Gibbons HL: Edetate disodium calcium, calcium-channel blockers, and heart disease. *JAMA* 246:1728, 1981.
162. Godfrey ME: EDTA chelation as a treatment of arteriosclerosis. *N Z Med J* 93:199, 1990.
163. Godfrey ME, Frackleton JP and Chappell LT: Chelation therapy for intermittent claudication (Letter). *Z Med J* 107:495, 1994.
164. Godfrey ME, Agnihotri R and Strauss A: Chelation and arteriosclerosis. *N Z Med J* 101(838):21, 1988.
165. Goetzel EJ: Oxygenation products of arachidonic acid as mediators of hypersensitivity and inflammation. *Med Clin North Am* 65(4):809, 1981.
166. Godfrey ME, Frackleton JP and Chappell LT: Chelation therapy for intermittent claudication (Letter). *Z Med J* 107:495, 1994.

167. Goodman and Gillman's Pharmacological Basis of Therapeutics: Heavy Metals and Heavy Metal Antagonists. 6:1615, 1980
168. Gordon T, Castelli WP, Hjortland MC, et al: High density lipoprotein as a protective factor against coronary heart disease. The Framington Study. *Am J Med* (62): 707, 1977.
169. Gordon GB, Vance RB: EDTA chelation therapy for atherosclerosis: History and mechanisms of action. *Osteopathic Annals* 4:38, 1976.
170. Gordon GF: Oral chelation with EDTA. *J Holistic Medicine* 8(1):79, 1986.
171. Gordon T, Kannel WB, Hjortland MC, McNamara PM: Menopause and coronary heart disease. The Framingham Study. *Am Intern Med* 89:157, 1978.
172. Gotto AM Jr: Chelation therapy in 1984. *Texas Medicine*, 80:36, 1984.
173. Gould RG: Metals and chelating agents in relation to atherosclerosis. *Fed Proc* 20 (Suppl 10):252, 1961.
174. Grier MT and Meyers DG: So much writing, so little science: A review of 37 years of literature on edetate sodium chelation therapy. *Ann Pharmacother* 27:1504, 1993.
175. Grumbles LA: Radionuclide studies of cerebral and cardiac arteriography before and after chelation therapy. *New Horizons in Holistic Health II (Symposium)*. Chicago, May 27, 1979.
176. Guldager B, Jelnes R, Jorgenson SJ, et al: EDTA treatment of intermittent claudication - a double-blind placebo-controlled study. *J Intern Med* 231:261, 1992.
177. Gutteridge JMC, Rowley DA, Halliwell B: Superoxide-dependent formation of hydroxyl radicals and lipid peroxidation in the presence of iron salts. *Biochem J* 206:605, 1982.
178. Gutteridge JMC, Rowley DA, Halliwell B and Westermarck T: Increased non-protein-bound iron and decreased protection against superoxide radical damage in cerebrospinal fluid from patients with neuronal ceroid lipofuscinoses. *Lancet* ii:459, 1982.
179. Halstead BW: *The Scientific Basis of EDTA Chelation Therapy*. Colton, CA, Golden Quill Publishers, Box 1278, Colton, CA 92324, 1979.
180. Hancke C and Flytlie K: Benefits of EDTA chelation therapy in arteriosclerosis: a retrospective study of 470 patients. *J Adv Med* 6(3):161, 1993.
181. Hancke C and Flytlie K: Manipulation with EDTA. *Ugeskr Laeger* 154:2213, 1992.
182. Hansotia P, Peters H, Bennett M and Brown R: Chelation therapy in Wegener's granulomatosis. Treatment with EDTA. *Ann Otol* 78:388, 1969.
183. Hardy HL: Clinical experience with the use of calcium disodium ethylenediaminetetraacetate in the therapy of lead poisoning. *Fed Proc* 20 (Suppl 10):252, 1961.
184. Harman D: The aging process. *Proc Natl Acad Sci USA* 78:7124, 1981.
185. Harper JW and Gordon GF: *Reprints of Medical Literature on Chelation Therapy*. Los Angeles, American Academy of Medical Preventives, 1975.

186. Hardy HL: Clinical experience with the use of calcium disodium ethylenediaminetetraacetate in the therapy of lead poisoning. *Fed Proc* 20 (Suppl 10):252, 1961.
187. Hatano S, Nishi Y, Usui T: Copper levels in plasma and erythrocytes in healthy Japanese children and adults. *Am J Clin Nutr* 35:120, 1982.
188. Haumont S and Vincent J: Action of calcium versenate on lead fixed in vivo in compact bone. *Exp Cell Res* 18:404, 1959.
189. Hausmann E: Changes in plasma phosphate concentration on infusion of calcium gluconate or Na₂EDTA. *Proc Soc Exp Biol Med* 134:182, 1970.
190. Hay DR: Chelation therapy. *N Z Med J* 101(841):122, 1988.
191. Hay DR: Chelation therapy. *N Z Med J* 101(845):246, 1988.
192. Heath DA, Knapp MS and Walker WHC: Comparison between inulin and ⁵¹Cr-labelled edetic acid for the measurement of glomerular filtration-rate. *Lancet* 2:1110, 1968.
193. Hegsted DM: Major minerals. Section A. Calcium and phosphorus. In: RS Goodheart and ME Shields (Eds.). *Modern Nutrition in Health and Disease*. Lea and Febiger, Phila., p. 266, 1974.
194. Heller J and Vostal J: Renal excretion of calcium-disodium-ethylenediamine-tetraacetic acid - a new tubular secretory mechanism? *Experientia* 20:99, 1964.
195. Herd JK and Vaughan JH: Calcinosis universalis complicating dermatomyositis - its treatment with Na₂EDTA. Report of two cases in children. *Arthritis Rheum* 7:259, 1964.
196. Hess ML, Manson NH, Okabe E: Involvement of free radicals in the pathophysiology of ischemic heart disease. *Can J Physiol Pharmacol* 60(11):1382, 1982.
197. Hjortland MC, McNamara PM, Kannel WB: Some atherogenic concomitants of menopause: The Framingham Study. *Am J Epidemiol* 103:304, 1976.
198. Holland JF, Danielson E and Sahagian-Edwards A: Use of EDTA in hypercalcemic patients. *Proc Soc Exp Biol Med* 84:359, 1953.
199. Holm LW: The use of calcium disodium salt of Versene in heavy-metal poisoning of livestock. *Proc Am Vet Med Assoc* (48): 33, 1954.
200. Hösli P: Therapy of scleroderma with the disodium salt of ethylenediaminetetraacetic acid; a contribution to the toxicology of versenate. Part I. *Arzneimittelforschung* 10:65, 1960.
201. Hösli P: Therapy of scleroderma with the disodium salt of ethylenediaminetetraacetic acid; a contribution to the toxicology of versenate. Part II. *Arzneimittelforschung* 10:177, 1960.
202. Jenkins DW: Toxic Trace Metals in Mammalian Hair and Nails. US Environmental Protection Agency publication No. (EPA)-600/4-79-049. Environmental Monitoring Systems Laboratory, 1979. (Available from National Technical Information Service, U.S. Department of Commerce, Springfield, Virginia 22161.)
203. Jick S and Karsh R: The effect of calcium chelation on cardiac arrhythmias and conduction disturbances. *Am J Cardiol* 4:287, 1959.

204. Johnson SAM: Use of the chelating agent edathamil disodium in atherosclerosis, sarcoidosis and other skin conditions with comments on tryptophan metabolism in sarcoidosis. *Wisconsin Med J* 59:651, 1960.
205. Jonas WB: Meta-analysis of EDTA chelation: math that doesn't matter. *J Adv Med* 7:109, 1994.
206. Jonas WB: Effectiveness of EDTA chelation therapy. *Circulation* 5:1352, 1995.
207. Jones KH and Fourman P: Edetic-acid test of parathyroid insufficiency. *Lancet* 2:119, 1963.
208. Jones RJ: Chelation therapy (Letter). *JAMA* 250:672, 1983.
209. Julian JJ: *Pass or Bypass? Chelation Extends Life*. Hollywood, Wellness Press, 1981.
210. Kalliomaki JL, Markkanen TK and Mustonen VA: Serum calcium and phosphorous homeostasis in man studied by means of the sodium EDTA test. *Acta Med Scan* 170:211, 1961.
211. Kaman RL, Rudolph CJ, McDonagh EW and Walker FM: Effect of EDTA chelation therapy on aortic calcium in rabbits on atherogenic diets: quantitative and histochemical studies. *J Adv Med* 3(1):13, 1990.
212. Kannel WB, Hjortland MC, McNamara PM, Gordon T: Menopause and the risk of cardiovascular disease. The Framingham Study. *Ann Intern Med* 85:447, 1976.
213. Kebe SR: ACTH and a chelating agent for schizophrenia. *West Med* 4:46, 1963.
214. Kimmel CA: Effect of route of administration on the toxicity and teratogenicity of EDTA in the rat. *Toxicol Appl Pharmacol* (40)2:299, 1977.
215. Kindness G and Frackelton JP: Effect of ethylene diamine tetraacetic acid (EDTA) on platelet aggregation in human blood. *J Adv Med* 2(4):519, 1989.
216. Kitchell JR and Meltzer LE: The potential uses of EDTA chelation therapy in the treatment of cardiovascular diseases. *Prog of Cardiovascular Dis* 3:338, 1961.
217. Kitchell JR, Palmon F, Aytan N, Meltzer LE: The treatment of coronary artery disease with disodium EDTA, a reappraisal. *Am J Cardiol* 11:501, 1963.
218. Klein R and Harris SB: Treatment of scleroderma, sclerodactylia and calcinosis by chelation (EDTA). *Am J Med* 19:798, 1955.
219. Klevay L: Hair as a biopsy material-assessment of copper nutriture. *Am J Clin Nutr* 23(8):1194, 1970.
220. Klevay L: Interactions of copper and zinc in cardiovascular disease. In: OA Levander and L. Cheng. *Micro-Nutrient Interactions :Vitamins, Minerals and Hazardous Elements*. *Ann NY Acad Sci* (355); 18, 1980.
221. Kneller LA, Uhl HSM and Brem J: Successful calcium disodium ethylene diamine tetraacetate treatment of lead poisoning in an infant. *N Engl J Med* 252:338, 1955.
222. Koen A, McCann DS and Boyle AJ: Some in vivo effects of chelation. II. Animal experimentation. *J Chron Dis* 16:329, 1963.

223. Konradi MG: Use of the disodium ethylene diaminetetraacetate for diagnosing latent forms of hypoparathyroidism. *Prob Endokrinol (Russ.)* (23)3: 46, 1977.
224. Kozlov VA and Novikova VM: Calcium ion-dependent immunosuppressive effect of ethylenediaminetetraacetic acid (EDTA). *Zh Mikrobiol, Epidemiol Immunobiol (Russ.)* (1): 69, 1978.
225. Lamar CP: Chelation therapy of occlusive arteriosclerosis in diabetic patients. *Angiology* 15:379, 1964.
226. Lamar CP: Chelation endarterectomy for occlusive atherosclerosis. *J Am Geri Soc* 14(3):272, 1966.
227. Lamar CP: Calcium chelation of atherosclerosis, nine years' clinical experience. Read before the Fourteenth Annual Meeting of the American College of Angiology, San Juan, PR, Dec 8, 1968. (Transcript available from ACAM, 23121 Verdugo Dr., Suite 204, Laguna Hills, CA 92653.)
228. Lamb DJ and Leake DS: The effect of EDTA on the oxidation of low density lipoprotein. *Atherosclerosis* 94:35, 1992.
229. Leipzig LS, Boyle AJ, McCann DS: Case histories of rheumatoid arthritis treated with sodium or magnesium EDTA. *J Chronic Dis* 22:553, 1970.
230. Leitner SA: Last chance to live. Wade Allen Publishing, Chesterfield, MO, 1980.
231. Lelievre P and Betz EH: Effect of versene on oxygen consumption of tissues in vitro. *C R Soc Biol* 155:199, 1961.
232. Levine SA, Kidd PM: Antioxidant Adaptation: Its Role in Free Radical Pathology. San Leandro, California, Biocurrents, 1985.
233. Liberman UA, Barzel U, De Vries A and Ellis H: Myositis ossificans traumatica with unusual course. Effect of EDTA on calcium, phosphorus and manganese excretion. *Am J Med Sci* 254:35, 1967.
234. Lochte HL Jr, Ferrebee JW and Thomas ED: The effect of heparin and EDTA on DNA synthesis by marrow in vitro. *J Lab Clin Med* 55:435, 1960.
235. Lockefeer JH, Hackeng WHL and Birkenhager JC: Parathyroid hormone secretion in disorders of calcium metabolism studied by means of EDTA. *Acta Endocr* 75:286, 1974.
236. Lonsdale D: EDTA chelation therapy. *Am J Surg* 166:316, 1993.
237. Luchi RJ, Helwig J Jr and Conn HL Jr: Quinidine toxicity and its treatment. An experimental study. *Am Heart J* 65:340, 1963.
238. Lyons RD: Chelation: miracle cure or false hope? *Med J Aust* 142: 519, 1985.
239. Magee HR: Chelation therapy for atherosclerosis. *Med J Aust* 143:127, 1985.
240. Magee HR: Chelation treatment of atherosclerosis. *Med J Aust* 142:514, 1985.

241. Maunsbach AB, Madden S and Latta H: Light and electron microscope-changes in proximal tubules of rats after administration of glucose, mannitol, sucrose or dextran. *Lab Invest* (11): 421, 1962.
242. Malinovska V, Zechmeister A, Brucknerova E et al: Ultrahistochemical study of the effect of glucagon and Chelation III on arterial wall structure after experimental calcification. *Folia Morphologica* 28:20, 1978.
243. Marcelle R and Lecomte J: On the cardiovascular activities of the sodium salt of ethylenediaminetetraacetic acid. *C R Soc Biol* 153:1483, 1959.
244. Margolis S: Chelation therapy is ineffective for the treatment of peripheral vascular disease. *Alternative Therapies Health Med* 1:53, 1995.
245. Marlow CG and Sheppard G: [51Cr] EDTA, [hydroxymethyl-14C] inulin-T for the determination of glomerular filtration rate. *Clin Chim Acta* 28:479, 1970.
246. Marney SR Jr and Des Prez RM: Rabbit platelet injury by soluble antigen and antibody. III. Effects of the sodium and magnesium salts of EDTA and EGTA. *J Immun* 106:1446, 1971.
247. Marsh L and Fraser FC: Chelating agents and teratogenesis. (Letter) *Lancet* 1:846, 1973.
248. Maxwell GM, Elliot RB and Robertson E: The effect of Na₃EDTA-induced hypocalcemia upon the general and coronary hemodynamics of the intact animal. *Am Heart J* 66:82, 1963.
249. Mayan H, Salomon O, Pauzner R, et al: EDTA induced pseudothrombocytopenia. *South Med J* 85:213, 1992.
250. Meltzer L, Kitchell J and Palmon F: The long term use, side effects, and toxicity of disodium ethylenediaminetetraacetic acid (EDTA). *Am J Med Sci* (242): 11, 1961.
251. Meltzer LE, Ural, ME and Kitchell JR: The treatment of coronary artery disease with disodium EDTA. in :Seven JJ and Johnson LA (eds): *Metal Binding in Medicine*. JB Lipincott Co Philadelphia pp 132, 1960.
252. McCann DS, Koen Z, Zdybek G and Boyle AJ: Effect of chelation on phosphorus metabolism in experimental atherosclerosis. *Circ Res* 11:880, 1962.
253. McCoy JE, Carre IJ and Freeman M: A controlled trial of edathamil calcium disodium in acrodynia. *Pediatrics* 25:304, 1960.
254. McDonagh E: Circulation changes in extremities with chelation therapy. Paper presented at American Academy of Medical Preventics Fall Conference, Las Vegas, Nevada, November, 1982.
255. McDonagh EW, Rudolph CJ and Cheraskin E: The Influence of EDTA salts plus multivitamin-trace mineral therapy upon total serum cholesterol/high-density lipoprotein cholesterol. *Med Hypotheses* 9:643, 1982.
256. McDonagh EW, Rudolph CJ and Cheraskin E: The homeostatic effect of EDTA with supportive multivitamin trace mineral supplementation upon high-density lipoproteins (HDL). *J Osteopath Phys Surg Calf* 8: #2, Spring 1982.

257. McDonagh EW, Rudolph CJ and Cheraskin E: The effect of intra-venous disodium ethylenediaminetetraacetic acid (EDTA) plus supportive multivitamin/trace mineral supplementation upon fasting serum calcium. *Med Hypotheses* 11:431, 1982.
258. McDonagh EW, Rudolph CJ and Cheraskin E: The glycohemoglobin (HbA1c) distribution in EDTA chelation eligible patients. *J Orthomolecular Psych* 12:72,1983.
259. McDonagh EW and Rudolph CJ: A collection of published papers showing the efficacy of EDTA chelation Therapy. Gladstone, MO, McDonagh Medical Center, 1991.
260. McDonagh EW: Chelation can cure: How to reverse heart disease, diabetes, stroke, high blood pressure or surgery. Kansas City, Platinum Pen Publications, 1983.
261. McDonagh EW, Rudolph CJ and Cheraskin E: The effect of intravenous disodium ethylene diamine tetraacetic acid (EDTA) upon blood cholesterol levels in a private practice environment. *J Intern Acad of Prev Med* 7:5, 1982.
262. McDonagh E.W, Rudolph CJ and Cheraskin E: The effect of EDTA chelation therapy with multivitamin/trace mineral supplementation upon reported fatigue. *J of Orthomol Psy* 13(4):1, 1984.
263. McDonagh EW ,Rudolph CJ, Cheraskin E and Wussow DG: The effect of EDTA chelation and supportive multivitamin/trace mineral supplementation with and without physical activity upon systolic blood pressure. *J Orthomol Psy* 13;1, 1984.
264. McDonagh EW,, Rudolph CJ and Cheraskin E: The psychotherapeutic potential of EDTA chelation. *The J OrthomolPsy* 14(3):214, 1985.
265. McDonagh EW, Rudolph CJ and Cheraskin E: The clinical changes in patients treated with EDTA chelation plus multivitamin/trace mineral supplementation. *The J Orthomol Psy* 14(1):61, 1985.
266. McDonagh EW, Rudolph CJ and Cheraskin E: An oculocerebrovasculometric analysis of the improvement in vascular stenosis following edta chelation therapy. *J Holistic Med* 4(1): 21, 1982.
267. McDonagh EW, Rudolph CJ, and Cheraskin E: The effect of EDTA chelation therapy plus multivitamin/trace mineral supplementation upon vascular dynamics: ankle/brachial doppler systolic blood pressure ratio. *J Holistic Med* 7(1):16, 1985.
268. McDonagh EW, Rudolph CJ ,Cheraskin E: The effect of EDTA chelation therapy plus supportive multivitamin-trace mineral supplementation upon renal function: A study in serum creatinine. *J Holistic Medicine* 4:146, 1982.
269. McDonagh EW, Rudolph CS, Cheraskin E: The effect of EDTA chelation therapy plus supportive multivitamin-trace mineral supplementation upon renal function: A study in blood urea nitrogen (BUN). *J Holistic Medicine* 5(2):163, 1983.
270. McGill WF: The effects of EDTA chelation therapy on plaque calcium and mineral metabolism in atherosclerotic rabbits. *Dissertation Abstracts International* Vol. 41, No. 04, October, 1980.
271. McGillem MJ and Mancini GB: Inefficacy of EDTA chelation therapy for coronary atherosclerosis. *N Eng j Med* 318:1618, 1988.

272. Mehbod H: Treatment of lead intoxication. Combined use of peritoneal dialysis and edetate calcium disodium. *JAMA* 201:972, 1967.
273. Meltzer LE: Chelation Therapy. In Seven MJ and Johnson LA (eds): *Metal Binding in Medicine*. Philadelphia, JB Lippincott, 1960.
274. Meltzer LE, Palmon FJ, and Kitchell JR: Hypoglycemia induced by disodium ethylenediamine tetra-acetic acid. *Lancet* 2:637, 1961.
275. Meltzer LE, Kitchell JR, and Palmon FJ: The long term use, side effects, and toxicity of disodium ethylenediamine tetraacetic acid (EDTA). *Am J Med Sci* 242:11, 1961.
276. Meltzer LE, Ural ME, Kitchell JR: The treatment of coronary artery disease with disodium EDTA, in Seven MJ, Johnson, LA (eds): *Metal Binding in Medicine*. Philadelphia, J.B. Lippencott Co, 1960, pp 132.
277. Mischol HR and Wildbolz R: Instrumental chemolysis of renal calculi: Indications and dangers. *J Urol* 105:607, 1971.
278. Moel DI and Kunar K: Reversible nephrotoxic reactions to a combined 2,3-dimercapto-1-propanol and calcium disodium ethylenediaminetetraacetic acid regimen to asymptomatic children with elevated blood lead levels. *Pediatrics* 70:259, 1982.
279. Monaco GP and Green S: Recognizing deception in the promotion of untested and unproven medical treatments. *NY State J of Med* 2:88, 1993.
280. Moncrieff AA, Koumides OP, Clayton BE, Patrick AD, Renwick AGC and Roberts GE: Lead poisoning in children. *Arch Dis Child* 39:1, 1964.
281. Mongan ES: The treatment of progressive systemic sclerosis with disodium edetate. *Arthritis Rheum* 8:1145, 1965.
282. Montgomery MR: Advances in medical fraud: chelation therapy replaces Laetrile. *J Florida Med Assoc* 73:681, 1986.
283. Morgan JM: Chelation therapy in lead nephropathy. *South Med J* 68:1001, 1973.
284. Morgan GT and Drew HDK: Researches on residual affinity and coordination. Pt II. acetylacetones of selenium and tellurium. *J Chem Soc (London)* 117:1456, 1920.
285. Müller SA, Brunstig LA and Winkelmann RK: The treatment of scleroderma with the new chelating agent edathamil. *Arch Dermatol* 3:305, 1962.
286. yers HL: Topical chelation therapy for varicose pigmentation. *Angiology* 17:66, 1966.
287. Myslak Z and Buczkowski M: Effect of calcium Versenate upon kidneys during saturnism therapy. *Pol Arch Med Wewnietrznej* (31); 853, 1961.
288. Nakano J, Cole B and Ishii T: Effects of disodium EDTA on the cardiovascular responses to prostaglandin E1. *Experientia* 24:808, 1968.
289. Nayler WG: Ventricular arrhythmias following the administration of Na₂EDTA. *J Pharmacol Exp Ther* 137:5, 1962.

290. Nedergaard OA and Vagne A: Effect of EDTA and other chelating agents on norepinephrine uptake by rabbit aorta in vitro. *Proc West Pharmacol Soc* 11:87, 1968.
291. Neldner KH, Winklemann RK and Perry HO: Scleroderma. An evaluation of treatment with disodium edetate. *Arch Derm* 86:305, 1962.
292. Nguyen-The-Minh: Treatment of digitalis intoxication with EDTA Na₂. *Presse Med* 71:2385, 1963.
293. Nikitina EL, Abramova MA: Treatment of atherosclerosis patients with Trilon-B (EDTA). (Russ, Moscow) *Kardiologiya* 12:137, 1972.
294. Nodine JH: Edetic acid therapy. (Letter) *JAMA* 212:628, 1970.
295. Nolan KR: Copper toxicity syndrome. *J Orthomol Psychiatr* 12(4):270, 1983.
296. Offer GW: The antagonistic action of magnesium ions and ethylenediaminetetraacetate on myosin A ATPase (potassium activated). *Biochim Biophys Acta* 89:566, 1964.
297. Oliver LD, Mehta R, Sarle HE: Acute renal failure following administration of ethylenediaminetetraacetic acid (EDTA). *Texas Medicine* February 80:40, 1984.
298. Olszewer E, Carter JP: EDTA chelation therapy in chronic degenerative disease. *Medical Hypothesis* 27:41, 1988.
299. Olszewer E and Carter JP: Chelation therapy: a retrospective study of 2,870 patients. *J Adv Med* 27:197, 1989.
300. Olszewer E, Sabbag FC and Carter JP: A pilot double-blind study of sodium magnesium EDTA in peripheral vascular disease. *J Natl Med Assoc* 82:173, 1989.
301. Olwin JH and Koppel JL: Reduction of elevated plasma lipid levels in atherosclerosis following EDTA therapy. *Proc Soc Exp Biol Med* 128:1137, 1968.
302. Ontka JA: Physical and chemical changes in isolated chylomicrons: prevention by EDTA. *J Lipid Res* 11:367, 1970.
303. Oser BL: Observations on the chronic ingestion of a chelating agent by rats and dogs. *Fed Proc* 20(Suppl 10):158, 1961.
304. Oser BL, Oser M and Spencer HC: Safety evaluation studies of calcium EDTA. *Toxicol Appl Pharmacol* 5:142, 1963.
305. Painter JT and Morrow EJ: Porphyria. Its manifestations and treatment with chelating agents. *Texas State J Med* 55:811, 1959.
306. Parfitt AM: Study of parathyroid function in man by EDTA. *J Clin Endocr* 29:569, 1969.
307. Passwater R A, Cranton EM: Trace Elements, Hair Analysis and Nutrition. New Canaan, CT, Keats Publishing Inc, 1983.
308. Patterson R: Chelation therapy and Uncle John. *CMAJ* 140(7):829, 1989.

309. Pavsek K, Drinal J and Selecky FV: Circulatory effects of disodium edetate in digoxin-induced ventricular tachycardia. *Cardiologia* 50:297, 1967.
310. Payne BA and Pierre RV: Pseudothrombocytopenia: a laboratory artifact with potentially serious consequences. *Mayo Clinic Proc* 59:123, 1984.
311. Payne RB: Creatinine clearance: A redundant clinical investigation. *Ann Clin Biochem* 23:243, 1986.
312. Payne JM and Sansom BF: The relative toxicity in rats of disodium ethylene diamine tetra-acetate, sodium oxalate and sodium citrate. *J Physiol* 170:613, 1964.
313. Peart WS, Quesada, T and Tenyi T: The effects of EDTA and EGTA on renin secretion. *Br J Pharmacol* (59) 2: 247, 1977.
314. Peng CF, Kane JJ, Murphy ML, Straub KD: Abnormal mitochondrial oxidative phosphorylation of ischemic myocardium reversed by calcium chelating agents. *J Mol Cell Cardiol* 9:897, 1977.
315. Pentel P, Jorgensen C and Somerville J: Chelation therapy for the treatment of atherosclerosis, an appraisal. *Minn Med* Feb:101, 1984.
316. Perry HM Jr: Chelation therapy in circulatory and sclerosing diseases. *Fed Proc* 20 (Suppl 10):254, 1961.
317. Perry HM Jr and Schroeder HA: Depression of cholesterol levels in human plasma following ethylenediamine tetraacetate and hydralazine. *J Chron Dis* 2:520, 1955.
318. Perry HM and Perry EF: Normal concentrations of some trace minerals in human urine, changes produced by EDTA. *J Clin Invest* (38):1452, 1959.
319. Perry H and Schroeder HA: Lesions resembling Vitamin B complex deficiency and urinary loss of zinc produced by EDTA. *Am J Med* (22): 168, 1957.
320. Perry HM Jr, and Camel GH: Some effects of CaNa₂ EDTA on plasma cholesterol and urinary zinc in man. In: MJ Seven and LA Johnson (Eds). *Metal Binding In Medicine*. JP Lippincott, Phila., 1960.
321. Peters H, Bichinan P and Reese H: Therapy of acute, chronic and mixed hepatic porphyria patients with chelating agents. *Neurology* (8): 621, 1958.
322. Peters RA, Stocken LA, and Thompson RHS: British Anti-Lewisite (BAL). *Nature* 156(3969):616, 1945.
323. Peters HA, Johnson SAM, Cam S, Oral S, Müftü Y and Ergene T: Hexachlorobenzene-induced porphyria: Effect of chelation on the disease, porphyrin and metal metabolism. *Am J Med Sci* 251:314, 1966.
324. Peters HA: Trace minerals, chelating agents and the porphyrias. *Fed Proc* 20 (Suppl 10):227, 1961.
325. Peterson CR: Adverse effects of chelation therapy. *JAMA* 250:2926, 1983.
326. Peto R, Doll R, Buckley JD, Sporn MB: Can dietary beta-carotene materially reduce human cancer rates? *Nature* 290:201, 1981.

327. Petrovic L, Stanovic M, Savicevic M and Poleti D: Aerosol inhalation of CaNa₂E.D.T.A. (mosatil) by workers constantly exposed to lead poisoning. *Brit J Indust Med* 17:201, 1960.
328. Pietronigro DD, Demopoulos HB, Hovsepian M, Flamm ES: Brain ascorbic acid (AA) depletion during cerebral ischemia. *Stroke* 13(1):117, 1982.
329. Platts-Mills A: EDTA chelation therapy. *NZ Med J*, 101(849):465, 1988.
330. Popovici A, Geschechter C, Reinovsky A, et al: Experimental control of calcium levels in vivo. *Proc Soc Exp Biol Med* (74): 415, 1950.
331. Popovici A, and Rubin M: EDTA as an anticoagulant in clinical laboratory studies. *Soc Exper and Biol Med* 74:415, 1950.
332. Pova H, Dantas LS, Faine R, Cranton EM: Antioxidant stimulation of phagocytosis using mannitol as a free radical scavenger. *J Advancement Med* 1(3):143, 1988.
333. Price JM: Some effects of chelating agents on tryptophan metabolism in man. *Fed Proc* 20 (Suppl 10):223, 1961.
334. Proceedings: Hearing on EDTA Chelation Therapy of the Ad Hoc Scientific Advisory Panel on Internal Medicine of the Scientific Board of the California Medical Society, March 26, 1976, San Francisco, California (Transcript available ACAM, 23121 Verdugo Dr., Suite 204, Laguna Hills, CA 92653.)
335. Pullman TN, Lavender AR and Forland M: Synthetic chelating agents in clinical medicine. *Ann Rev Med* 14:175, 1963.
336. Rassmussen H and Bordier P: The Physiological and Cellular Basis of Metabolic Bone Disease. Williams and Wilkins, Baltimore, 1974.
337. Rathmann KL and Golightly LK: Chelation therapy of atherosclerosis. *Drug Intell Clin Pharm* 18:1000, 1984.
338. Raymond JZ and Gross PR: EDTA: preservative dermatitis. *Arch Derm* 100:436, 1969.
339. Rees EL: Aluminum poisoning of Papua New Guinea natives as shown by hair testing. *J Orthomol Psychiatry* 12(4):312, 1983.
340. Reidenberg MM: Kidney function and drug action. *N Eng J Med* 313:816, 1985.
341. Remagen W, Hiller FK and Sanz CM: Metabolic changes in experimental poisoning with ethylenediamine-tetraacetic acid. *Arzneimittelforschung* 11:1097, 1961.
342. Renoux M and Mikol C: Chelation. *Presse Med* 72:3117, 1964.
343. Reuber MD: Accentuation of Ca edetate nephrosis by cortisone. *Arch Path* 76:382, 1963.
344. Reuber MD and Bradley JE: Acute versenate nephrosis occurring as the result of treatment for lead intoxication. *JAMA* (174): 263, 1960.
345. Reuber MD and Lee CW: Calcium disodium edetate nephrosis in inbred rats. *Arch Environ Health* (13): 544, 1966.

346. Reuber MD: Severe nephrosis in older male rats given calcium disodium edetate. *Arch Environ Health* 15:141, 1967.
347. Reuber MD and Schmieler GC: Edetate kidneys lesions in rats. *Arch Environ Health* 5:430, 1962.
348. Reuber MD: Hepatic lesions in young rats given calcium disodium edetate. *Toxicol Appl Pharmacol* 11:321, 1967.
349. Reuter H, Niemeyer G and Gross R: The distribution of EDTA and citrate in blood and plasma. *Throm Diath Haemorrh* 19:213, 1968.
350. Rieders F, Dunnington WG and Brieger H: The efficacy of edathamil calcium disodium in the treatment of occupational lead poisoning. *Indust Med* 24:195, 1955.
351. Riordan HD, Cheraskin E, Dirks M, et al: EDTA chelation/hypertension study: clinical patterns as judged by the Cornell Medical Index Questionnaire. *J Orthomolel Med* 4:91, 1989.
352. Riordan HD, Cheraskin E, Dirks M, et at: Another look at renal function and the EDTA treatment proces. *J Othomolecular Medicine* 2(3):185, 1987.
353. Riordon HD, Cheraskin E, Dirks M, et al: Electrocardiographic changes associated with EDTA chelation therapy. *J Adv Med* 1:191, 1988.
354. Robinson DM: Chelation therapy. *N Z Med J* 74:750, 1982.
355. Rosenbaum JL, Mason D and Seven, MJ: Effect of disodium ethylenediaminetetraacetic acid on digitalis intoxication. *All J ed Sci* (240): 77, 1960.
356. Rossi EC: Effects of ethylenediaminetetraacetic acid (EDTA) and citrate upon platelet glycolysis. *J Lab Clin Med* 69:204, 1967.
357. Rotota MC and Sanowski R: Acute intermittent porphyria. *J Med Soc New Jersey* 61:101, 1964.
358. Rubin M, Gignac S, Bessman SP and Belknap EL: Enhancement of Lead Excretion in Humans by Disodium Calcium Ethylenediamine Tetraacetate. *Science* (117): 659, (June 12), 1953.
359. Rudolph CJ, McDonagh EW and Barber RK: Effect of EDTA chelation and supportive multivitamin/trace mineral supplementation on chronic lung disorders: a study of FVC and FEV1. *J Adv Med* 2(4): 553,1989.
360. Rudolph CJ, McDonagh EW and Barber RK: The effect of EDTA chelation on serum iron. *J Adv Med* 4:39, 1991.
361. Rudolph CJ, Samuels RT and McDonagh EW: Visual field evidence of macular degeneration reversal using a combination of EDTA and multiple vitamin and trace mineral therapy. *J Adv Med* 7:203, 1994.
362. Rudolph CJ and McDonagh EW: Effect of EDTA chelation and supportive multivitamin trace mineral supplementation on carotid circulation: case report. *J Adv Med* 3,(1):5, 1990.
363. Rudolph CJ, Mcdonagh EW And Barber RK: A nonsurgical approach to obstructive carotid stenosis using EDTA Chelation. *J Adv Med* 4(3):157, 1991.

364. Rudolph CJ, McDonagh EW and Barber RK: An observation of the effect of EDTA chelation and supportive multivitamin/trace mineral supplementation on blood platelet volume: a brief communication. *J Adv Med* 3(3):179, 1990.
365. Rudolph CJ, McDonagh EW, Wusson, DG: The effect of intravenous ethylene diamine tetraacetic acid (EDTA) upon bone density levels. *J Advancement Med* 1(2):79, 1988.
366. Rudolph CJ, McDonagh EW: The chelation carrier solution: An analysis of osmolarity and sodium content. *Journal of the International Academy of Preventive Medicine* 8(1):26, 1983.
367. Rutman JZ, Meltzer LE, Kitchell JR, et al: Effect of metal ions on in vitro gluconeogenesis in rat kidney cortex slices. *Am J Physiol* 208:841, 1965.
368. Sachs HK, Blanksma LA, Murray EF and O'Connell MJ: Ambulatory treatment of lead poisoning: report of 1,155 cases. *Pediatrics* 46:389, 1970.
369. Schachter MB Letter to the editor: chelation therapy. *Circulation* 91(4):2291, 1995.
370. Schachter MB: Health fraud versus innovation-a letter to the editor. *J Adv in Med* 6(3):198, 1993.
371. Schoenthaler SJ : Commercial hair analysis: Lack of reference norms and high reliability within and between seven selected laboratories for seventeen trace minerals. *International Journal of Biosocial Research* 8(1):84, 1986.
372. Schroeder HA: *The Poisons Around Us*. Bloomington, IN, Indiana University Press, 1974.
373. Schroeder HA: A practical method for the reduction of plasma cholesterol in man. *J Chron Dis* 4:461, 1956.
374. Schroeder HA, Menhard EM and Perry HM Jr: Antihypertensive effects of metal binding agents. *J Lab & Clin Med* 23:416, 1955.
375. Schubert J: Chelation in medicine. *Sci Am* 214:40, 1966.
376. Schwartz SL, Hayes JR, Ide RS, Johnson CB and Doolan PD: The nephrotoxicity of ethylenediaminetetraacetic acid. *Biochem Pharmacol* (15): 377, 1966.
377. Schwartz SL, Johnson CB and Coolan PD: Study of the mechanism of renal vacuologenesis induced in the rat by ethylenediaminetetraacetate. *Mol Pharm* 6:54, 1970.
378. Schwartz SL, Johnson CB, Hayes JR and Doolan PD: Subcellular localization of ethylenediaminetetraacetate in the proximal tubular cell by the rat kidney. *Biochem Pharmacol* 16:2413, 1967.
379. Scott PJ: Chelation therapy for degenerative vascular disease. *N Z Med J* 95:538, 1982.
380. Scott PJ. Chelation therapy: evolution or devolution of a nostrum? *N Z Med J* 101:109, 1988.
381. Seelig M: *Magnesium Defficiency in the Pathogenesis of Disease* Plenum Press, NY, 1980.
382. Sehnert KW, Clague AF, Cheraskin E: The improvement in renal function following EDTA chelation and multivitamin-trace mineral therapy: A study in creatinine clearance. *Medical Hypothesis* 15(3):307, 1984.

383. Selander S: Treatment of lead poisoning. A comparison between the effects of sodium calcium edetate and penicillamine administered orally and intravenously. *Brit J Indust Med* 24:272, 1967.
384. Seto DSY and Freeman JM: Lead nephropathy in childhood. *Am J Dis Child* 107:337, 1964.
385. Seven MJ: Observations on the dosage of intravenous chelating agents. *Antibiotic Med* 5:251, 1958.
386. Seven MJ and Johnson LA: *Metal Binding in Medicine*, JB Lippincott, Phila, 1960.
387. Seven MJ: Observations on the toxicity of intravenous chelating agents. In: MJ Seven and LA Johnson (Eds). *Metal Binding in Medicine* JP Lippincot, Phila, 1960.
388. Shibata S: Toxicological studies of EDTA salt (sodium ethylenediamine tetra-acetate. *Folia Pharmacol Japonica* 52:113, 1956, and *Chem Abst* 51:9918, 1957.
389. Shrand H: Treatment of lead poisoning with intramuscular edathamil calcium-disodium. *Lancet* 1:310, 1961.
390. Sidbury JB Jr, Bynum JC, and Fetz LL: Effect of Chelating Agent on Urinary Lead Excretion. Comparison of Oral and Intravenous Administration. *Proceedings of the Society of Experimental Biology and Medicine*, 82: 266, 1953.
391. Silverglade A: Chelation clinics. *Chest* 87:274 1985.
392. Sincock A: Life extension in the rotifer by application of chelating agents. *J Gerontol* 30:289, 1975.
393. Singal PK, Kapur N, Dhillon KS, Beamish RE, Dhalla NS: Role of free radicals in catecholamine-induced cardiomyopathy. *Can J Physiol Pharmacol* 60(11):1340, 1982.
394. Sloth-Nielsen J, Guldager B, Mouritzen C, et al: Arteriographic findings in EDTA chelation therapy on peripheral arteriosclerosis. *Am J Surg* 162:122, 1991.
395. Soffer A: Chelation therapy for arteriosclerosis. *JAMA* 233:1206, 1975.
396. Soffer A: Chihuahuas and laetrile, chelation therapy, and honey from Boulder, Colo. *Arch Intern Med* 136:865, 1976.
397. Soffer A: Chelation clinics, an abuse of the physician's freedom of choice. *Chest* 86:157, 1984.
398. Soffer A and Toribara T: Changes in serum and spinal fluid calcium effected by disodium ethylenediaminetetraacetate. *J Lab Clin Med* (58): 542, 1961.
399. Soffer A: *Chelation Therapy*. Charles C. Thomas, Springfield, IL, 1964.
400. Soffer A, Toribara T and Sayman A: Myocardial responses to chelation. *Brit Heart J* 23:690, 1961.
401. Soffer A, Toribara T, Moore-Jones D and Weber D: Clinical applications and untoward reactions and untoward reactions of chelation in cardiac arrhythmias *Arch Inten Med* 106:824, 1960.
402. Smith LL: *Cholesterol Autooxidation*. New York Plenum Press, 1981.

403. Spence JD: Chelation therapy for intermittent claudication. ACP Journal Club, July/August page 8, 1992.
404. Spencer H: Studies of the effect of chelating agents in man. Ann NY Acad Sci (88):435, 1960.
405. Spencer H, Greenberg J, Berger E, Perrone M and Lazlo D: Studies on the effect of ethylenediaminetetraacetic acid in hypercalcemia. J Lab Clin Med (47): 29, 1956.
406. Spencer H, Vankinscott V, Lewin I and Lazlo D: Removal of calcium in man by ethylenediaminetetraacetic acid. (EDTA). J Clin Invest (31):1023, 1952.
407. Stacey BD and Thorborn GD: Chromium-51 ethylenediaminetetraacetate for estimation of glomerular filtration rate. Science 152:1076, 1966.
408. Stamp TCB, Stacey TE and Rose GA: Comparison of glomerular filtration rate measurements using inulin, 51Cr-EDTA, and a phosphate infusion technique. Clin Chim Acta 30:351, 1970.
409. Stamp TCB: 51Cr-edetic-acid clearance and G.F.R. (Letter) Lancet 2:1348, 1968.
410. Stankovic D and Keser-Stankovi, M: Effects of EDTA on liver and kidneys and protective effects of EDTA on these organs in animals treated with lead. Folia Med (14):101, 1979.
411. Stecker RH and Bennett M: Chelation in clinical otosclerosis. Arch Otolaryng 70:627, 1959.
412. Steven FS: The effect of chelating agents on collagen interfibrillar matrix interactions in connective tissue. Biochim Biophys Acta 140:522, 1967.
413. Stevenson JG and Covington TR: Chelation therapy in atherosclerosis. Ann Intern Med 97:789, 1982.
414. Stimson WH: Vitamin A intoxication in adults. Report of a case with summary of the literature. New Eng J Med (265): 369, 1961.
415. Sullivan JL : Iron and the sex difference in heart disease risk. Lancet 1(8233):1293, 1981.
416. Sullivan JL :Iron, aspirin and heart disease risk, editorial. JAMA 247(6):751, 1982.
417. Sullivan JL: Vegetarianism, ischemic heart disease, and iron, editorial, Burr ML: Reply to letter by Sullivan, editorial. Am J Clin Nutr 37:882, 1983.
418. Surawicz B: Use of chelating agent, ethylene diamine tetraacetic acid, in digitalis intoxication and cardiac arrhythmias. Prog in CardiovascularDis p. 432, 1960.
419. Surawicz B, MacDonald M, Kaljot V, et al: Treatment of cardiac arrhythmias with salts of ethylenediaminetetraacetic acid (EDTA). Am Heart J(58): 493, 1959.
420. Suvorov A and Markosyan RA: Some mechanisms of EDTA on platelet aggregation. (All Union Cardiol. Res. Cent. Moscow, Russia.) Byall Eksp Biol Med (1)5: 587, 1981.
421. Swenerton H and Hurley LS: Teratogenic effects of chelating agents and their prevention by zinc. Science Vol 173-3991:62, 1971.

422. Szekely P and Wynne NA: Effects of calcium chelation on digitalis-induced cardiac arrhythmias. *Brit Heart J* 25:589, 1963.
423. Tamburino G, Fiore CE and Petralito A: Comparison of effects of EDTA (versenate) infusion on plasma calcitonin levels in senile osteoporotic and age matched healthy subjects. *Ircs Med Sci: Libr Compend* 4(8): 362, 1976.
424. Tappel AL: Will antioxidant nutrients slow aging processes? *Geriatrics* 23:97, 1968.
425. Tappel AL: Lipid peroxidation damage to cell components. *Fed Proc* 32:1870, 1973.
426. Tappel L: Vitamin E and Selenium Protection from In Vivo lipid peroxidation. *Micronutrient Interactions: Vitamins, Minerals, and Hazardous Elements*. OA L Cheng I Acad Sci (355): 18, 1981.
427. Tate SS: Effect of metal ions and EDTA on the activity of rabbit liver fructose 1,6-diphosphatase *Biochim Biophys Res Commun* 24:662, 1966.
428. Taylor CB, Peng SK, Werthessen NT, Tham P, Lee KT: Spontaneously occurring angiotoxic derivatives of cholesterol. *Am J Clin Nutr* 32:40, 1979.
429. Teisinger J: Biochemical responses to provocative chelation by edetate disodium calcium. *Arch Environ Health* 23:280, 1971.
430. Thatcher RW, Lester ML, McAlester R, Horst R: Effects of low levels of cadmium and lead on cognitive functioning in children. *Arch Environ Health* 37(3):159, 1982.
431. Thompson LJ: Chelation therapy. *N Z Med J* 103:326, 1990.
432. Timmerman A, Kallistratos G and Genner O: In situ lysis of kidney stones. Urologic technique and drugs. *Congr Int Ther* 161-75.
433. Timmermann A and Kallistratos G: Chemotherapy in nephrolithiasis. *Isr J Med Sci* 7:689, 1971.
434. Timmermann A and Kallistratos G: Modern aspects of chemical dissolution of human renal calculi by irrigation. *J Urol* 95:469, 1966.
435. Traub YM, Samuel R, Lubin E, Lewitus Z and Rosenfeld JB: A comparison between the clearances of inulin, endogenous creatinine, and ⁵¹Cr-EDTA. *Isr J Med Sci* 9:487, 1973.
436. Trowbridge W and Walker M: Chelation Therapy: The Key to Unclogging Your Arteries, Improving Oxygenation, Treating Vision Problems. 3rd Ed. Marble Falls, Texas, Better Health Booklet Service, 1990.
437. Truss F: Clinical significance of kidney stone chemolysis. Kidney tolerance to ethylenediaminetetraacetic acid (EDTA). *Med Welt* (7): 238, 1971.
438. Tsang RC, Chen I-W and McEnergh P: Parathyroid function tests with EDTA infusions in infancy and childhood. *J Pediatr* 88:250, 1976.
439. Uhl HSM, Brown HH, Zlatkis A, Zak B, Myers GB and Boyle AJ: Effect of ethylenediamine tetraacetic acid on cholesterol metabolism in rabbits. *Am J Clin Path* 23:1226, 1950.

440. van der Schaar P: Brief Communication: Exercise Tolerance Tests in Chelation Therapy. *J Adv Med* 2(4): 563, 1989.
441. van Rij AM, Solomon C, Packer SGK, et al: Chelation therapy for intermittent claudication. A double-blind, randomized, controlled trial. *Circulation* 90, 1194, 1994.
442. van Rij AM, Solomon C, Packer SGK and Hopkins WG: [Chelation therapy] Response (Letter). *Circulation* 5:1350, 1995.
443. van Rij AM, Solomon C, Packer SGK and Hopkins WG: [Effectiveness of EDTA chelation therapy] Response (Letter). *Circulation* 5:1352, 1995.
444. Vesselinovitch D, Wissler RW, Fiseher-Dzoga K, Hughes R, DuBien L: Regression of atherosclerosis in rabbits. I. Treatment with low fat diet, hyperoxia and hypolipidemic agents. *Atherosclerosis* 19:259, 1974.
445. Vincent GM, Anderson JL, Marshall HW: Coronary spasm producing coronary thrombosis and myocardial infarction. *N Engl J Med* 309(14):220, 1983.
446. Vogt W and Cottier II: Necrotizing nephrosis after treatment of a case of subacute-chronic lead poisoning with CaEDTA in high doses. *Schweizerische Medizinische Wochenschrift* 87(22): 665 1957.
447. Vora NM, Williams GA, Hargis GK, BowserEN, Kawahara W, Jackson BL, Henderson WJ and Kukreja SC: Comparative effect of calcium and of the adrenergic system on calcitonin secretion in man. *J Clin Endocrinol Metab* 46(4): 567, 1978.
448. Walker FMcG: The effects of EDTA chelation therapy on plaque calcium and mineral metabolism in atherosclerotic rabbits. *Physiology (Dissertation Abstracts International)* Vol. 41, No. 04, 1980.
449. Walker M: How to Prevent or Reverse Hardening of the Arteries in: Chelation Therapy. M Evans Co, Inc., New York, 1981.
450. Walker M: The Miracle Healing Power of Chelation Therapy. Canfield, Ohio, Fischer Publishing, 1984.
451. Walker M: The Chelation Way: The Complete Book of Chelation Therapy. Garden City Park, NY, Avery Publishing, 1990.
452. Walker M: Chelation Therapy: How to Prevent or Reverse Hardening of the Arteries. Atlanta, '76 Press, 1987.
453. Walker M and Gordon G: The chelation Answer. Atlanta, '76 Press, 1983.
454. Wartman A: Lampe TL, McCann DS, Boyle AJ: Plaque reversal with MgEDTA in experimental atherosclerosis: Elastin and collagen metabolism. *J Atheros Res* 7:331, 1967.
455. Weeden RP, Mallik DK and Batitman V: Detection and treatment of occupational lead nephropathy. *Arch Intern Med.* (139); 53, 1979.
456. Weinig E and Schwerd W: Il Nocere! Hazards of treatment of lead poisoning with calcium versenate. *100:1788*, 1958.

457. Welcher FJ: The Analytical Uses of Ethylenediamine Tetraacetic Acid 5. Van Nostrand, Princeton.
458. What is chelation therapy for arteriosclerosis? Br Med J 285: 1982.
459. Whitaker, J: You have alternatives to bypass surgery. Health Healing Lett Feb, 1993.
460. Whitaker JA, Austin W and Nelson JD: Edathamil calcium disodium (Versenate) diagnostic test for lead poisoning. Pediatrics 29:384, 1962.
461. Wilder LW, DeJode LR, Milstein SW, et al: Mobilization of atherosclerotic plaque calcium with EDTA utilizing the isolation-perfusion principle. Surgery 52:793, 1962.
462. Wilder LW, DeJode LR, Milstein SW, et al: Mobilization of atherosclerotic plaque calcium with EDTA utilizing the isolation-perfusion principle. Surgery 52:793, 1962.
463. Willett WC, Morris JS, Pressel S, et al: Prediagnostic serum selenium and risk of cancer. Lancet 2(8343):130, 1983.
464. Williams: An Introduction to Bio-inorganic Chemistry. Charles C. Thomas, Springfield, Illinois.
465. Williams JD, Matthews GA and Judd AW: Oral calcium disodium versenate in treatment of lead poisoning. Brit J Indust Med 19:211, 1962.
466. Wills ED: Mechanisms of lipid peroxide formation in tissues: Role of metals and haematin proteins in the catalysis of the oxidation of unsaturated fatty acids. Biochem Biophys Acta 98:238, 1965.
467. Willson RL: Iron, zinc, free radicals and oxygen tissue disorders and cancer control, in Iron Metabolism. Ciba Foundn Symp 51 (new series). Amsterdam, Elsevier, pp 331, 1977.
468. Winder PR and Curtis AC: Edathamil in the treatment of scleroderma and calcinosis cutis. Arch Derm 82:732, 1960.
469. Wirebaugh SR and Geraets DR: Apparent failure of edetic acid chelation therapy for the treatment of coronary atherosclerosis. DICP Ann Pharm 24:22, 1990.
470. Wissler RW, Vesselinovitch D: Regression of atherosclerosis in experimental animals and man. Mod Concepts Cardiovasc Dis 46:28, 1977.
471. Wong G: The chemistry of EDTA (a brief treatment). H Harper and G Gordon (Eds.). Reprints of Medical Literature on Chelation Therapy. Amer Acad Med Prev Los Angeles, 1974.
472. Woods SM, Peters HA and Johnson SAM: Chelation therapy in cutaneous porphyria. A review and report of a five-year recovery. Arch Derm 84:920, 1961.
473. Wynn JE, Riet B, Van't and Borzelleca JF: The toxicity and pharmacodynamics of EGTA: oral administration to rats and comparisons with EDTA. Toxicol Appl Pharmacol 16:807, 1970.
474. Yang WC: Stimulatory effect of EDTA on cardiac mitochondrial respiration. Biochem Biophys Res Commun 2:22, 1960.
475. Yokel RA: Hair as an indicator of excessive aluminum exposure. Clin Chem 28(4):662, 1982.

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